

Metalloproteases, Vascular Remodeling, and Atherothrombotic Syndromes

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Defects in the synthesis and breakdown of the extracellular matrix (ECM) are now seen as key processes in the development of atherosclerosis and its thrombotic complications. Correlations have been observed between circulating levels of ECM biomarkers and the clinical manifestations of and risk factors for atherosclerosis. Several matrix metalloproteinases (MMPs), endopeptidases that can degrade the ECM, such as MMP-9 and MMP-10, play important roles in the pathophysiology of atherothrombosis and contribute to the expansion of abdominal aortic aneurysms. Moreover, they may also be useful biomarkers of atherosclerotic risk and serve as predictors of coronary and cerebrovascular disease recurrence. Although at present the effect of tissue inhibitors of MMPs (TIMPs) on cardiovascular disease prognosis is still uncertain, the ECM could be a promising therapeutic target in atherothrombotic disease, and several MMP inhibitors are currently undergoing clinical trials.

Key words: Atherosclerosis. Extracellular matrix. Vascular remodeling. Biomarkers.

Metalloproteasas, remodelado vascular y síndromes aterotrombóticos

Las alteraciones en la síntesis y/o la degradación de la matriz extracelular (MEC) emergen como procesos clave en el desarrollo de la aterosclerosis y sus complicaciones trombóticas. Se ha observado una asociación entre los biomarcadores circulantes de la MEC y las manifestaciones clínicas y los factores de riesgo ateroscleróticos. Diversas metaloproteasas (MMP), endopeptidasas que degradan la MEC, como MMP-9 y 10, además de desempeñar un papel relevante en la fisiopatología del proceso aterotrombótico y contribuir a la expansión de los aneurismas arteriales, pueden ser de utilidad como biomarcadores de riesgo aterosclerótico y predictores de recurrencia de enfermedad coronaria y cerebrovascular. Aunque actualmente el papel de los inhibidores de MMP (TIMP) en el pronóstico cardiovascular es más incierto, la MEC puede representar una diana terapéutica atractiva en la aterotrombosis, y diversos inhibidores de las MMP se encuentran en fase de investigación clínica.

Palabras clave: Aterosclerosis. Matriz extracelular. Remodelado vascular. Biomarcadores.

INTRODUCTION

Atherosclerosis is a diffuse, systemic disease of the arterial network, the local manifestations of which are associated with clinical problems such as myocardial infarction, stroke, etc. Vulnerable lesions (high risk plaques) are characterized by a large necrotic core, a thin fibrous layer (fibroatheroma), and an inflammatory infiltrate (monocytes/macrophages, T lymphocytes, and mastocytes, etc). Although the rupture of a fibrous plaque

is the main cause of intraluminal thrombosis in acute coronary syndromes and is responsible for 75% of all deaths following acute myocardial infarction, thrombosis has also been observed in eroded plaques and in those with calcified nodules.^{1,2} In high risk patients, plaque vulnerability is a multifocal phenomenon that involves different lesions around the coronary tree. In clinical practice, the signs of vulnerable plaques are difficult to discern. Current efforts directed towards identifying patients at risk are therefore based on three types of markers: *a*) images (eg, magnetic resonance, optical coherence tomography, molecular imaging, etc), for characterizing the composition of atheroma plaques and identifying those that are vulnerable; *b*) functional markers of vascular homeostasis (eg, arterial thickening and the endothelial-dependent vasodilatory response); and *c*) circulating markers (based on the measurement of

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markers of inflammation, oxidative stress, and thrombosis in the blood).³⁻⁶ The use of biomarkers of extracellular matrix (ECM) remodeling has been less studied, even though the ECM provides the structural and functional platform for the blood vessels; changes in its synthesis and/or breakdown must therefore play a key role in the development of atherosclerotic lesions, vascular remodeling, and plaque rupture.^{7,8} This has led to the proposal that certain matrix metalloproteinases (MMP), such as pregnancy-associated plasma protein A (PAPP-A),⁹ may serve as potential biomarkers of atherosclerotic progression. A loss of equilibrium between the molecules and factors that induce the degradation of the ECM and those that favor its synthesis and accumulation could be of prime importance in the development of clinical atherothrombotic syndromes.

PHYSIOLOGY OF THE EXTRACELLULAR MATRIX

The arterial wall is dominated by collagen type I and II, macrophages, and smooth muscle cells; these cell types govern the remodeling of the ECM. Fibronectin, laminin, elastin, and proteoglycans are also involved in ECM synthesis. The balance between ECM synthesis and breakdown is regulated by the equilibrium between the proteases that favor degradation (MMP) and their tissue inhibitors (TIMP) (Figure 1). The MMP are a family of zinc-dependent endopeptidases produced by

monocytes, endothelial cells, and smooth muscle cells, whose function it is to break down numerous components of the ECM as well as other proteins not associated with it. They are synthesized and secreted as inactive proenzymes with a propeptide domain rich in cysteine able to fold and interact with the Zn⁺⁺ of the catalytic domain – thus impeding any enzyme activity. Activation requires the removal of the propeptide domain. Based on their structure, substrate specificity, and membrane binding (Table), these enzymes are classified as collagenases (MMP-1, 8, and 13), stromelysins (MMP-3, 10, and 11), gelatinases (MMP-2 and 9), membrane-type MMP (MT-MMP), and others (matrilysin, metalloelastase, etc).¹⁰ Their activity is regulated transcriptionally, post-translationally, and via interactions with specific inhibitors. A number of growth factors, cytokines, thrombin, and hormones increase the transcription of these enzymes, while heparin, transforming growth factor beta (TGFβ) and corticoids inhibit it.^{11,12} The extracellular activation of latent zymogens (pro-MMP molecules) provides a second control point. The main physiological activator of the MMP is plasmin, which converts inactive forms into active molecules by the proteolysis of the propeptide link and the exposure of the catalytic domain.¹³ Other enzymes, such as thrombin, factor Xa, and the MMP themselves also possess MMP-activating activity. Finally, the activity of the MMP is regulated by TIMP, of which four are known (TIMP-1, 2, 3, and 4). Inhibition is achieved via

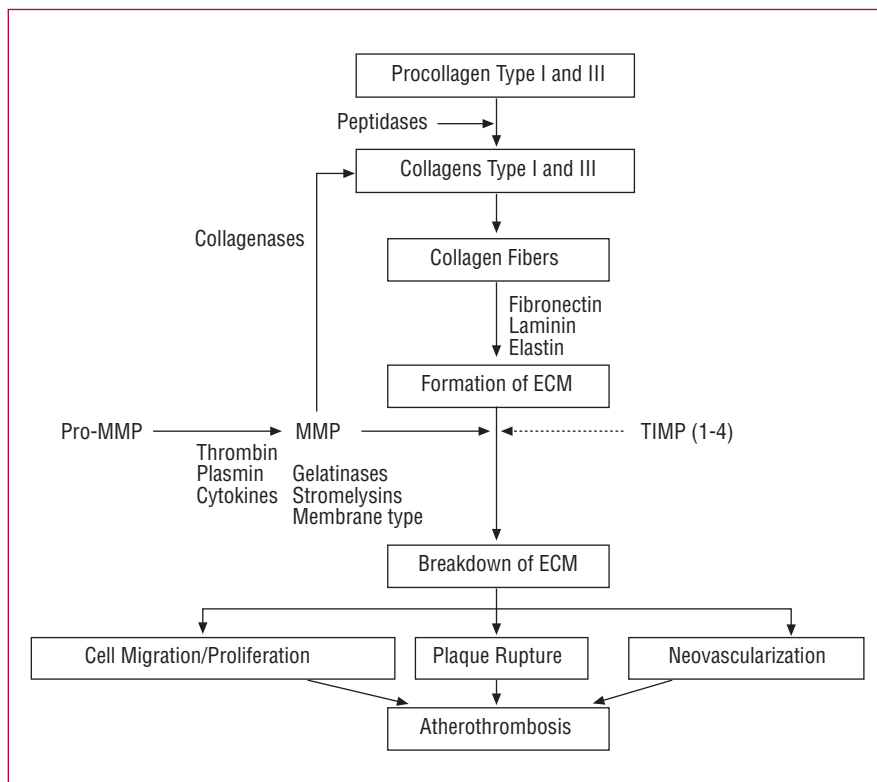


Figure 1. Synthesis and breakdown of the extracellular matrix in atherosclerosis: balance between metalloproteases (MMP) and their inhibitors (TIMP).

TABLE. Characteristics and Specificity of the Main Metalloproteases*

MMP (Type)	Name	ECM Substrate
Collagenases		
MMP-1	Collagenase-1	Collagen I, II, III, VII, VIII, and X, gelatin, proteoglycans, tenascin, entactin
MMP-8	Collagenase-2	Collagen I, II, III, V, VIII, and X, gelatin, aggrecan
MMP-13	Collagenase 3	Collagen I, II, III, IV, IX, X, and XIV, gelatin, tenascin, fibronectin, aggrecan, osteonectin
Gelatinases		
MMP-2	Gelatinase A	Collagen I, IV, V, VII, X, XI, and XIV, gelatin, elastin, fibronectin, laminin, aggrecan, versican, osteonectin, proteoglycans
MMP-9	Gelatinase B	Collagen IV, V, VII, X, XIV, gelatin, elastin, aggrecan, versican, proteoglycans, osteonectin
Stromelysins		
MMP-3	Stromelysin-1	Collagen III, IV, V, and IX, gelatin, aggrecan, versican, proteoglycan, tenascin, fibronectin, laminin, osteonectin
MMP-10	Stromelysin-2	Collagen III, IV, V, gelatin, casein, aggrecan, elastin, proteoglycans
MMP-11	Stromelysin-3	Casein, laminin, fibronectin, gelatin, collagen IV, transferrin
Membrane type		
MMP-14	MT1-MMP	Collagen I, II, and III, casein, elastin, fibronectin, vitronectin, tenascin, proteoglycans, laminin, entactin
MMP-15	MT2-MMP	Tenascin, fibronectin, laminin
MMP-16	MT3-MMP	Collagen III, gelatin, casein, fibronectin
MMP-17	MT4-MMP	ND
MMP-24	MT5-MMP	ND
MMP-25	MT6-MMP	ND
Others		
MMP-7	Matrilysin	Collagen IV and X, gelatin, aggrecan, proteoglycans, fibronectin, laminin, entactin, tenascin, casein, transferrin, integrin b ₄ , osteonectin, elastin
MMP-12	Metalloelastase	Collagen IV, gelatin, elastin, casein, laminin, proteoglycans, fibronectin, vitronectin, entactin
MMP-20	Enamelysin	amelogenin
MMP-23A	MMP-21	ND
MMP-23B	MMP-22	ND
MMP-26	Matrilysin 2	Collagen IV, fibrinogen, fibronectin, casein
MMP-27	ND	ND
MMP-28	Epilysin	Casein

*ECM indicates extracellular matrix; MMP, metalloproteinases; ND, not determined.

the irreversible binding of these molecules to the active enzyme. Thus, in summary, the proteolytic balance depends on the relative concentration of activators and inhibitors¹³ (Figure 1).

THE EXTRACELLULAR MATRIX AND ATHEROTHROMBOSIS

The equilibrium between MMP and TIMP is critical in the maintenance of the integrity of the cardiovascular system.¹³⁻¹⁶ This has led to the proposal that any alteration towards the breakdown of the ECM contributes to the progression of atherosclerosis and plaque instability.¹⁷ Certainly, the participation of the MMP in different mechanisms fundamental to atherothrombotic progression has been reported (Figure 2):

– MMP favor monocyte infiltration of the vascular wall. An increase in the expression of certain MMP, such

as MMP-12, leads to macrophage infiltration, the rupture of the internal elastic lamina, and the acceleration of the atherosclerotic process.¹⁸

– The induction and activation of the MMP, particularly MMP-14 (MT1-MMP), favors the invasion of plaques by vascular smooth muscle cells and fibroblasts. The migration and proliferation of these cell types is important in the development of intimal hyperplasia.^{19,20}

– The activity of MMP-2 and 9, among others, is indispensable for the neovascularization of atherosclerotic plaques. This process, induced by proangiogenic and inflammatory stimuli, appears to be necessary for their growth, and is associated with the vulnerability of advanced lesions.^{21,22}

– The role of MMP in the formation and resolution of thrombi remains somewhat unclear (Figure 3). An abnormal reduction in the activity of ADAMTS-13 favors the appearance of thrombotic microangiopathies,²³ and there is evidence of molecular interactions between

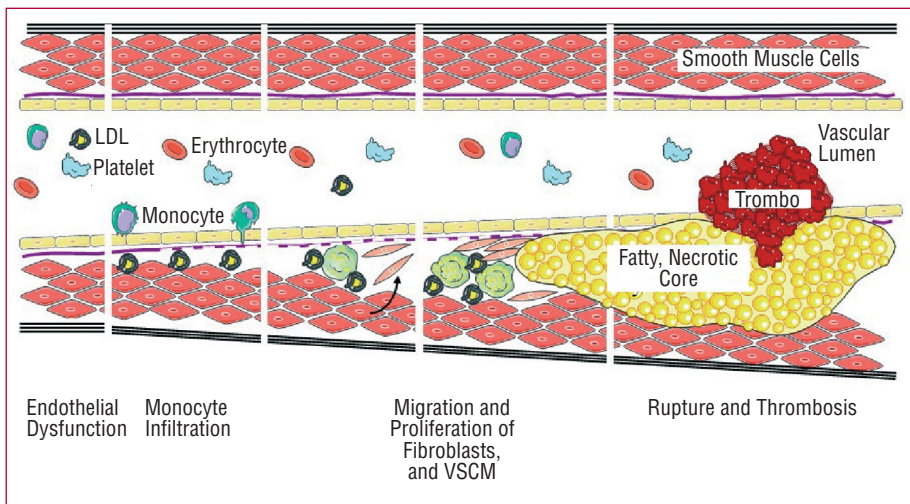


Figure 2. The atherosclerotic process. LDL indicates low density lipoprotein; VSMC, vascular smooth muscle cells.

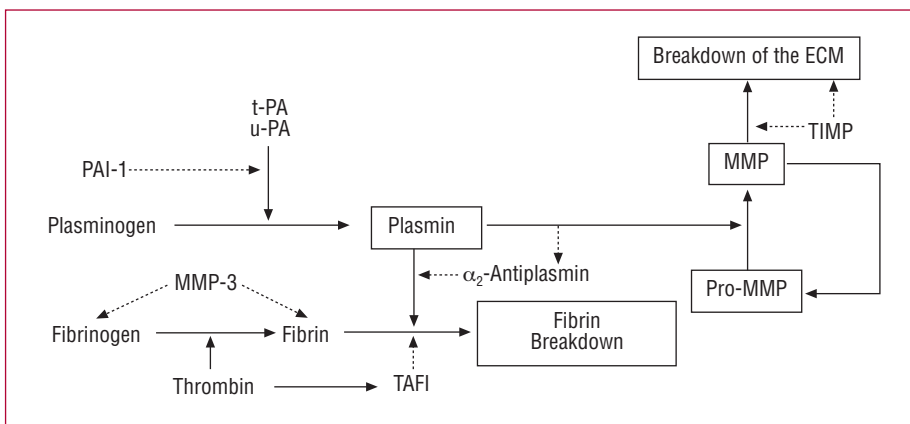


Figure 3. Diagram showing the role of metalloproteases in thrombolysis and fibrinolysis. ECM indicates extracellular matrix; MMP, metalloproteinase; PAI-1, plasminogen activator inhibitor type I; TAFI, thrombin activated fibrinolysis inhibitor; TIMP, tissue inhibitors of metalloproteinases; t-PA, tissue plasminogen activator; u-PA, urokinase type plasminogen activator. Modified from Lijnen.²⁴

MMP-3 and enzymes, substrates, and inhibitors of the fibrinolytic system. This suggests the MMP to have some role in the latter.²⁴ They might therefore be involved in different types of thrombotic disease. Finally, thrombi themselves may be a source of proteolytic activity that might be important in the atherothrombotic process.²⁵

The physiopathological importance of changes in ECM metabolism in the development of atherothrombotic syndromes is supported by numerous pieces of evidence. It is interesting to note that intravascular thrombolysis and acute myocardial infarction are rare consequences of restenosis (one of the limitations of percutaneous revascularization). Restenotic plaques appear as a consequence of the rapid proliferation of vascular smooth muscle cells and the accumulation of ECM at the lesion site. These plaques, however, rarely rupture or express MMP-9 (gelatinase B).²⁶ In contrast, the progress of an atherosclerotic plaque from a fatty streak to an advanced, unstable element is slow, and is associated with an increase in its cellular and ECM contents as well as in proteolytic activity. This latter activity is mainly associated with the macrophages and vascular smooth muscle cells at the plaque shoulders and around the

necrotic core, and is accompanied by the expression and concentration of (mostly) MMP-1 (collagenase), and MMP-9.^{27,28} An increase in the gelatinase MMP-9 has also been observed in the coronary plaques of patients with unstable angina compared to those with stable angina,^{29,30} and has been related to post-infarction ventricular remodeling.^{31,32} Increased concentrations of MMP-9 have also been reported in human brain tissue following ischemic and hemorrhagic ictus, and the induction of MMP-9 has been reported in the cerebral vasculature following fibrinolysis. This indicates that this enzyme may contribute to ischemic cerebral lesions and perihematomal edema as well as to cerebral hemorrhages and neurovascular lesions following fibrinolysis.^{33,34} These local alterations can be detected at the systemic level; patients with angiographic proof of coronary heart disease show an alteration in the balance of fibrinolysis/proteolysis in their peripheral blood.³⁵ In addition, it has been reported that the urine MMP-9 concentration increases, while that of TIMP-1 is reduced, after an acute myocardial infarction.³⁶

The expression and activity of MMP-9 in an atherosclerotic plaque, which is mainly associated with

macrophages, may be the consequence of an increase in the production of the NADPH-dependent superoxide anion since it coincides with the expression of NADPH oxidase and the production of free radicals.³⁷ However, the gelatinases and collagenases are not the only MMP associated with atherosclerosis; our group has recently shown an increase in MMP-10 (stromelysin 2) in advanced carotid plaques obtained by endarterectomy.³⁸ MMP-10 has also been associated with aortic aneurysms, which are characterized by destructive remodeling of the vascular ECM and rupture of the wall.³⁹ Finally, the TIMP-1 concentrations have been reported to increase in calcified areas of human atherosclerotic plaques,⁴⁰ further indicating the inhibition of MMP activity may be related to greater plaque instability.

METALLOPROTEASES AND ATHEROSCLEROTIC RISK FACTORS

The majority of the classic risk factors for atherosclerosis have been related to changes in the concentrations of different ECM biomarkers and the overall Framingham score.^{41,42} The mechanisms behind this, however, are unclear. It may be that the risk factors modulate the vascular structure and stability of plaques, which in turn influence concentrations of MMP and TIMP, or via modifications in collagen production. Neither can it be ruled out that modified biomarker values are an epiphenomenon or an adaptive response to the process of atherosclerosis.

Age and Sex

Advanced age and male sex have been related to elevated concentrations of TIMP-1.⁴¹ Hormone therapy has been reported to reduce MMP-9 concentrations in post-menopausal women.⁴³

Dyslipidemia

In the clinical setting, the Framingham Heart Study reported MMP-9 to be detectable only in 20% of subjects, in whom it was associated with high levels of low-density lipoprotein cholesterol (LDL-C).⁴² In contrast, TIMP-1 was detected in all subjects and increased with the total cholesterol/high-density lipoprotein cholesterol (HDL-C) ratio.⁴¹ In subjects with familial hypercholesterolemia, an increase has been reported in concentrations of MMP-3 and 9, and TIMP-1, particularly in those at greater cardiovascular risk—in whom the serum concentrations of MMP-3 and TIMP-1 have been associated with carotid atherosclerotic lesions.⁴⁴ In addition, the abnormally high levels of MMP-9 in patients with familial hypercholesterolemia and coronary heart disease can be reduced through treatment with statins, although no correlation between the reduction of the MMP concentration and lower cholesterol concentrations is reported.⁴⁵

Under experimental conditions it has been shown that, following angioplasty, the vascular expression of MMP-1 increases in a porcine hypercholesterolemic model, perhaps as a consequence of oxidative stress.⁴⁶ In *in vitro* experiments, oxidized LDL has been shown to increase the production of MMP-1, 3, and 9, while that of TIMP-1 is reduced. In contrast, HDL reduces the production of several MMP types.^{47,48} In monocytes/macrophages it has been reported that oxidized LDL acts synergistically with proinflammatory factors to induce the expression of MMP-1 and MMP-9, while HDL prevents the induction of MMP-1 via these same agents.⁴⁹

Diabetes, Obesity, and Metabolic Syndrome

MMP-9 and TIMP-1 are increased in patients with diabetes and metabolic syndrome.^{50,51} In experimental studies it has been observed that increased glucose levels induce the expression of MMP-1 and 2 by endothelial cells, and of MMP-9 by macrophages, but have no effect on TIMP-1.⁵² In a clinical study involving patients with diabetes, the control of blood sugar and atherosclerotic risk factors reduced TIMP-1 without modifying MMP-9 or TIMP-2.⁵³

The concentration of TIMP-1, but not of MMP-9, has been related to body mass index.⁴¹ Finally, in a study involving obese women, a reduction in MMP-1 was noticed one year after performing stomach reduction surgery.⁵⁴

High Blood Pressure

An increase in circulating MMP-9 – but also of TIMP-1 – has been reported in patients with high blood pressure and general thickening of the artery walls.⁵⁵ High blood pressure is associated with an increase in collagen synthesis and a reduction in its breakdown.⁵⁶

Tobacco and Alcohol

Smokers show increased concentrations of MMP-9 and TIMP-1, the extent of which is partly related to the duration of tobacco exposure.⁵⁷ No association has been reported between the abusive intake of alcohol and circulating TIMP-1 and MMP-2 concentrations,⁵⁸ although an increase in MMP-9 and 2 expression has been reported, especially in the alveolar macrophages. The latter is probably related to lung remodeling in the context of acute respiratory distress syndrome.⁵⁹ An inverse association has been reported between moderate alcohol consumption and circulating TIMP-1 concentrations.⁴¹

Inflammation

Different inflammatory stimuli, such as TNF α and interleukin 1, as well as other factors, regulate the expression of MMP.⁶⁰ In experimental studies it has been

shown that C-reactive protein, an inflammatory biomarker of atherosclerotic risk, induces the expression of MMP-1 by macrophages, and of MMP-10 by endothelial cells, without affecting TIMP-1 concentrations.^{38,61} It has also been reported that circulating MMP-9 concentrations in patients with coronary heart disease, which are higher than those in healthy subjects, are directly associated with the concentrations of inflammation markers such as plasma C-reactive protein, interleukin 6, and fibrinogen. However, no differences in nor association with the MMP-2 levels have been reported.⁶² Similarly, in other clinical studies involving asymptomatic subjects, C-reactive protein have been associated with those of MMP-9 and 10, but not with those of MMP-2 or MMP-3.³⁸ Recently, our group has shown that C-reactive protein induces the expression of MMP-1 and MMP-10 by human endothelial cells, and that asymptomatic subjects with a proinflammatory profile show elevated concentrations of MMP-10.³⁸ Doxycycline, which reduces the concentration of C-reactive protein also reduces the activity of MMP-9.⁶³ It has been proposed that the infection of macrophages or smooth muscle cells with *Chlamydia pneumoniae* can induce the production of MMP since a strong relationship has been seen between the presence of this bacterium and the immunodetection of MMP-9 in atherosclerotic plaques. However, no causal relationship has been established.⁶⁴

PROGNOSTIC VALUE OF EXTRACELLULAR MATRIX BIOMARKERS

Quantification of the turnover of the ECM in cardiovascular tissues faces methodological difficulties. The different procedures available for evaluating fibrosis/degeneration in tissues (eg, endomyocardial biopsy, intravascular ultrasonography, etc) are, apart from being invasive, of limited use. Further, circulating ECM biomarkers are not specific for vascular tissue, and since MMP binds ECM components a local increase does not always correlate with a systemic increase. Currently, it is unsure which of the 40 possible ECM biomarkers has the best prognostic value. Therefore, when establishing criteria for the selection of biomarkers of interest and clinical applicability, certain factors should be taken into account: *a*) it should be demonstrated that markers reflect the remodeling of the ECM, *b*) there should be evidence that high concentrations are found in patients with stable disease, *c*) the biomarker should be stable in plasma or serum, and *d*) the method should be standardized and show little variability. As well as showing good reproducibility, biomarkers should be helpful in arriving at a diagnosis, at a prognosis, and in the therapeutic monitoring of atherosclerosis.^{3,4}

Several studies have reported an association between MMP and TIMP that predicts an adverse prognosis in a wide range of cardiovascular diseases.⁶⁵⁻⁶⁸ The most promising circulating biomarkers of ECM breakdown

are MMP-9 (gelatinase B) and 10 (stromelysin 2). An increase in MMP-9 predicts a narrowing of the arterial lumen, restenosis after the positioning of a stent, and cardiovascular death in patients with coronary heart disease.⁶⁹⁻⁷² It has also been related to the expansion and rupture of aortic aneurysms^{73,74} and increases the risk of hemorrhagic transformation in patients with ictus.⁷⁵ These biomarkers may also offer prognostic information in the primary prevention setting. MMP-9 predicts ischemic heart disease, and/or high blood pressure in patients with no prior cardiovascular disease,⁷⁶ and our group has recently shown that MMP-10 provides a marker of subclinical atherosclerosis; a correlation was seen between plasma MMP-10 levels and the thickness of the intima media of the carotid artery in a large group of patients with no history of cardiovascular disease.⁷⁷

THE EXTRACELLULAR MATRIX AS A THERAPEUTIC TARGET IN ATHEROTHROMBOSIS

Modulation of pericellular proteolysis may be a good target for therapeutic intervention in the context of atherosclerosis.⁷⁸ Perhaps the most representative example of the effect of therapeutic agents on vascular remodeling is provided by thrombolytic treatment. This stimulates the expression of MMP via plasmin, and promotes the degradation of collagen.⁷⁹ Statins also reduce the concentration of MMP-9 and other MMP,^{80,81} as do angiotensin II type I receptor antagonists,⁸² while other agents with cardiovascular action (such as carvedilol and the thiazolidinediones) reduce concentration of MMP-1.^{83,84} Certain antibiotics, such as doxycycline and the tetracyclins, reduce the vascular and systemic expression of several MMP,⁶³ while antioxidants reduce the expression of MMP-1.⁴⁶

Several pharmaceutical companies are interested in developing synthetic inhibitors of MMP, although none are yet available for clinical use.^{85,86} Some experimental studies show that the inhibition of MMP via the use of TIMP may delay the progression of an atherosclerotic plaque.^{71,87} In addition, the inhibition of MMP in the advanced stages of atherosclerosis can protect against the development of unstable plaques, the formation of aneurysms and heart failure.^{88,89}

CONCLUSIONS

Numerous studies have shown that the remodeling of the ECM of artery walls can be monitored by determining the circulating concentration of different MMP and TIMP. These molecules can be considered biomarkers of prognostic potential with respect to the recurrence of ischemic heart disease, the development of heart failure, and the formation of aneurysms in patients with clinical atherosclerosis. They may also be used in asymptomatic subjects with risk factors or in those with subclinical

atherosclerosis. Prospective studies currently underway should allow us to clarify the diagnostic and prognostic potential of these biomarkers in vascular diseases, and to assess their prospects as new therapeutic targets in atherosclerosis.

REFERENCES

- Virmani R, Kolodgie FD, Burke AP, Finn AV, Gold HK, Tulenko TN, et al. Atherosclerotic plaque progression and vulnerability to rupture: angiogenesis as a source of intraplaque hemorrhage. *Arterioscler Thromb Vasc Biol.* 2005;25:2054-61.
- Schaar JA, Muller JE, Falk E, Virmani R, Fuster V, Serruys PW, et al. Terminology for high-risk and vulnerable coronary artery plaques. Report of a meeting on the vulnerable plaque, June 17 and 18, 2003, Santorini, Greece. *Eur Heart J.* 2004;25:1077-82.
- Vasan RS. Biomarkers of cardiovascular disease: molecular basis and practical considerations. *Circulation.* 2006;113:2335-62.
- Tardif JC, Heinonen T, Orloff D, Libby P. Vascular biomarkers and surrogates in cardiovascular disease. *Circulation.* 2006;113:2936-42.
- Naghavi M, Falk E, Hecht HS, Jamieson MJ, Kaul S, Berman D, et al. From vulnerable plaque to vulnerable patient—Part III: Executive summary of the Screening for Heart Attack Prevention and Education (SHAPE) Task Force report. *Am J Cardiol.* 2006;98:2H-15H.
- García-Moll X. Marcadores de inflamación y de antiinflamación en el síndrome coronario agudo: ¿listos para usarlos en la práctica clínica? *Rev Esp Cardiol.* 2005;58:615-7.
- Libby P, Lee RT. Matrix matters. *Circulation.* 2000;102:1874-6.
- Shah PK, Galis ZS. Matrix metalloproteinase hypothesis of plaque rupture: players keep piling up but questions remain. *Circulation.* 2001;104:1878-80.
- Pinon P, Kaski JC. Inflamación, aterosclerosis y riesgo cardiovascular: PAPP-A, Lp-PLAZ y cistatina C. ¿Nuevas aportaciones o información redundante? *Rev Esp Cardiol.* 2006;59:247-58.
- Jones CB, Sane DC, Herrington DM. Matrix metalloproteinases: a review of their structure and role in acute coronary syndrome. *Cardiovasc Res.* 2003;59:812-23.
- Galis ZS, Muszynski M, Sukhova GK, Simon-Morrissey E, Unemori EN, Lark MW, et al. Cytokine-stimulated human vascular smooth muscle cells synthesize a complement of enzymes required for extracellular matrix digestion. *Circ Res.* 1994;75:181-9.
- Galis ZS, Kranzhofer R, Fenton JW 2nd, Libby P. Thrombin promotes activation of matrix metalloproteinase-2 produced by cultured vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol.* 1997;17:483-9.
- Paramo JA, Montero I, Rodríguez JA, Orbe J. Metalloproteasas en aterosclerosis: implicaciones fisiológicas y terapéuticas. *Clin Invest Arterioscl.* 2005;17:133-141.
- Galis ZS, Khatri JJ. Matrix metalloproteinases in vascular remodeling and atherogenesis: the good, the bad, and the ugly. *Circ Res.* 2002;90:251-62.
- Paramo JA, Orbe J, Rodríguez JA. Atheroma plaque stabilization: a new concept based on the dynamic biology of atherosclerosis. *Med Clin (Barc).* 2003;121:583-7.
- Dollery CM, Libby P. Atherosclerosis and proteinase activation. *Cardiovasc Res.* 2006;69:625-35.
- Newby AC. Do metalloproteinases destabilize vulnerable atherosclerotic plaques? *Curr Opin Lipidol.* 2006;17:556-61.
- Liang J, Liu E, Yu Y, Kitajima S, Koike T, Jin Y, et al. Macrophage metalloelastase accelerates the progression of atherosclerosis in transgenic rabbits. *Circulation.* 2006;113:1993-2001.
- Filippov S, Koenig GC, Chun TH, Hotary KB, Ota I, Bugge TH, et al. MT1-matrix metalloproteinase directs arterial wall invasion and neointima formation by vascular smooth muscle cells. *J Exp Med.* 2005;202:663-71.
- Johnson JL. Matrix metalloproteinases: influence on smooth muscle cells and atherosclerotic plaque stability. *Expert Rev Cardiovasc Ther.* 2007;5:265-82.
- Moreno PR, Purushothaman KR, Fuster V, Echeverri D, Trusczyńska H, Sharma SK, et al. Plaque neovascularization is increased in ruptured atherosclerotic lesions of human aorta: implications for plaque vulnerability. *Circulation.* 2004;110:2032-8.
- Herrmann J, Lerman LO, Mukhopadhyay D, Napoli C, Lerman A. Angiogenesis in atherogenesis. *Arterioscler Thromb Vasc Biol.* 2006;26:1948-57.
- Tsai HM. ADAMTS13 and microvascular thrombosis. *Expert Rev Cardiovasc Ther.* 2006;4:813-25.
- Lijnen HR. Matrix metalloproteinases and cellular fibrinolytic activity. *Biochemistry (Mosc).* 2002;67:92-8.
- Swedenborg J, Eriksson P. The intraluminal thrombus as a source of proteolytic activity. *Ann NY Acad Sci.* 2006;1085:133-8.
- Brown DL, Hibbs MS, Kearney M, Isner JM. Differential expression of 92-kDa gelatinase in primary atherosclerotic versus restenotic coronary lesions. *Am J Cardiol.* 1997;79:878-82.
- Galis ZS, Sukhova GK, Lark MW, Libby P. Increased expression of matrix metalloproteinases and matrix degrading activity in vulnerable regions of human atherosclerotic plaques. *J Clin Invest.* 1994;94:2493-503.
- Sukhova GK, Schonbeck U, Rabkin E, Schoen FJ, Poole AR, Billingham RC, et al. Evidence for increased collagenolysis by interstitial collagenases-1 and -3 in vulnerable human atheromatous plaques. *Circulation.* 1999;99:2503-9.
- Lofthus IM, Naylor AR, Goodall S, Crowther M, Jones L, Bell PR, et al. Increased matrix metalloproteinase-9 activity in unstable carotid plaques. A potential role in acute plaque disruption. *Stroke.* 2000;31:40-7.
- Uzui H, Harpf A, Liu M, Doherty TM, Shukla A, Chai NN, et al. Increased expression of membrane type 3-matrix metalloproteinase in human atherosclerotic plaque: role of activated macrophages and inflammatory cytokines. *Circulation.* 2002;106:3024-30.
- Cleutjens JP, Creemers EE. Integration of concepts: cardiac extracellular matrix remodeling after myocardial infarction. *J Card Fail.* 2002;8:S344-8.
- Creemers EE, Cleutjens JP, Smits JF, Daemen MJ. Matrix metalloproteinase inhibition after myocardial infarction: a new approach to prevent heart failure? *Circ Res.* 2001;89:201-10.
- Rosell A, Ortega-Aznar A, Álvarez-Sabin J, Fernández-Cadenas I, Ribo M, Molina CA, et al. Increased brain expression of matrix metalloproteinase-9 after ischemic and hemorrhagic human stroke. *Stroke.* 2006;37:1399-406.
- Wang X, Lee SR, Arai K, Tsuji K, Rebeck GW, Lo EH. Lipoprotein receptor-mediated induction of matrix metalloproteinase by tissue plasminogen activator. *Nat Med.* 2003;9:1313-7.
- Paramo JA, Orbe J, Fernández J. Fibrinolysis/proteolysis balance in stable angina pectoris in relation to angiographic findings. *Thromb Haemost.* 2001;86:636-9.
- Fitzsimmons PJ, Fough R, Lawrence ME, Gantt DS, Rajab MH, Kim H, et al. Urinary levels of matrix metalloproteinase 9 and 2 and tissue inhibitor of matrix metalloproteinase in patients with coronary artery disease. *Atherosclerosis.* 2006;doi:10.1016/j.atherosclerosis.2006.07.027
- Zalba G, Fortuno A, Orbe J, San José G, Moreno MU, Belzunce M, et al. Phagocytic NADPH oxidase-dependent superoxide production stimulates matrix metalloproteinase-9: implications for human atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2007;27:587-93.
- Montero I, Orbe J, Varo N, Beloqui O, Monreal JI, Rodríguez JA, et al. C-reactive protein induces matrix metalloproteinase-1 and -10 in human endothelial cells: implications for clinical and subclinical atherosclerosis. *J Am Coll Cardiol.* 2006;47:1369-78.
- Ogata T, Shibamura H, Tromp G, Sinha M, Goddard KA, Sakalihan N, et al. Genetic analysis of polymorphisms in biologically relevant candidate genes in patients with abdominal aortic aneurysms. *J Vasc Surg.* 2005;41:1036-42.

40. Orbe J, Fernández L, Rodríguez JA, Rabago G, Belzunce M, Monasterio A, et al. Different expression of MMPs/TIMP-1 in human atherosclerotic lesions. Relation to plaque features and vascular bed. *Atherosclerosis*. 2003;170:269-76.
41. Sundstrom J, Evans JC, Benjamin EJ, Levy D, Larson MG, Sawyer DB, et al. Relations of plasma total TIMP-1 levels to cardiovascular risk factors and echocardiographic measures: the Framingham heart study. *Eur Heart J*. 2004;25:1509-16.
42. Sundstrom J, Evans JC, Benjamin EJ, Levy D, Larson MG, Sawyer DB, et al. Relations of plasma matrix metalloproteinase-9 to clinical cardiovascular risk factors and echocardiographic left ventricular measures: the Framingham Heart Study. *Circulation*. 2004;109:2850-6.
43. Koh KK, Ahn JY, Kang MH, Kim DS, Jin DK, Sohn MS, et al. Effects of hormone replacement therapy on plaque stability, inflammation, and fibrinolysis in hypertensive or overweight postmenopausal women. *Am J Cardiol*. 2001;88:1423-6, A8.
44. Beaudoux JL, Giral P, Bruckert E, Bernard M, Foglietti MJ, Chapman MJ. Serum matrix metalloproteinase-3 and tissue inhibitor of metalloproteinases-1 as potential markers of carotid atherosclerosis in infraclinical hyperlipidemia. *Atherosclerosis*. 2003;169:139-46.
45. Koh KK, Son JW, Ahn JY, Jin DK, Kim HS, Choi YM, et al. Comparative effects of diet and statin on NO bioactivity and matrix metalloproteinases in hypercholesterolemic patients with coronary artery disease. *Arterioscler Thromb Vasc Biol*. 2002;22:e19-23.
46. Orbe J, Rodríguez JA, Arias R, Belzunce M, Nespereira B, Pérez-Illarbe M, et al. Antioxidant vitamins increase the collagen content and reduce MMP-1 in a porcine model of atherosclerosis: implications for plaque stabilization. *Atherosclerosis*. 2003;167:45-53.
47. Huang Y, Song L, Wu S, Fan F, Lopes-Virella MF. Oxidized LDL differentially regulates MMP-1 and TIMP-1 expression in vascular endothelial cells. *Atherosclerosis*. 2001;156:119-25.
48. Li D, Liu L, Chen H, Sawamura T, Ranganathan S, Mehta JL. LOX-1 mediates oxidized low-density lipoprotein-induced expression of matrix metalloproteinases in human coronary artery endothelial cells. *Circulation*. 2003;107:612-7.
49. Ardans JA, Economou AP, Martinson JM Jr, Zhou M, Wahl LM. Oxidized low-density and high-density lipoproteins regulate the production of matrix metalloproteinase-1 and -9 by activated monocytes. *J Leukoc Biol*. 2002;71:1012-8.
50. Roberts CK, Won D, Pruthi S, Kurtovic S, Sindhu RK, Vaziri ND, et al. Effect of a short-term diet and exercise intervention on oxidative stress, inflammation, MMP-9, and monocyte chemotactic activity in men with metabolic syndrome factors. *J Appl Physiol*. 2006;100:1657-65.
51. Hayden MR, Sowers JR, Tyagi SC. The central role of vascular extracellular matrix and basement membrane remodeling in metabolic syndrome and type 2 diabetes: the matrix preloaded. *Cardiovasc Diabetol*. 2005;4:9.
52. Death AK, Fisher EJ, McGrath KC, Yue DK. High glucose alters matrix metalloproteinase expression in two key vascular cells: potential impact on atherosclerosis in diabetes. *Atherosclerosis*. 2003;168:263-9.
53. Marx N, Froehlich J, Siam L, Ittner J, Wierse G, Schmidt A, et al. Antidiabetic PPAR gamma-activator rosiglitazone reduces MMP9 serum levels in type 2 diabetic patients with coronary artery disease. *Arterioscler Thromb Vasc Biol*. 2003;23:283-8.
54. Laimer M, Kaser S, Kranebitter M, Sandhofer A, Muhlmann G, Schwelberger H, et al. Effect of pronounced weight loss on the nontraditional cardiovascular risk marker matrix metalloproteinase-9 in middle-aged morbidly obese women. *Int J Obes (Lond)*. 2005;29:498-501.
55. Yasmin J, McEnery CM, Wallace S, Dakham Z, Pulsalkar P, Maki-Petaja K, et al. Matrix metalloproteinase-9 (MMP-9), MMP-2, and serum elastase activity are associated with systolic hypertension and arterial stiffness. *Arterioscler Thromb Vasc Biol*. 2005;25:372.
56. Querejeta R, López B, González A, Sánchez E, Larman M, Martínez Ubago JL, et al. Increased collagen type I synthesis in patients with heart failure of hypertensive origin: relation to myocardial fibrosis. *Circulation*. 2004;110:1263-8.
57. Nakamura T, Ebihara I, Shimada N, Koide H. Effect of cigarette smoking on plasma metalloproteinase-9 concentration. *Clin Chim Acta*. 1998;276:173-7.
58. Ponomarenko Y, Leo MA, Kroll W, Lieber CS. Effects of alcohol consumption on eight circulating markers of liver fibrosis. *Alcohol*. 2002;37:252-5.
59. Burnham EL, Moss M, Ritzenthaler JD, Roman J. Increased fibronectin expression in lung in the setting of chronic alcohol abuse. *Alcohol Clin Exp Res*. 2007;31:675-83.
60. Arenas IA, Xu Y, López-Jaramillo P, Davidge ST. Angiotensin II-induced MMP-2 release from endothelial cells is mediated by TNF-alpha. *Am J Physiol Cell Physiol*. 2004;286:C779-84.
61. Williams TN, Zhang CX, Game BA, He L, Huang Y. C-reactive protein stimulates MMP-1 expression in U937 histiocytes through Fc[gamma]RII and extracellular signal-regulated kinase pathway: an implication of CRP involvement in plaque destabilization. *Arterioscler Thromb Vasc Biol*. 2004;24:61-6.
62. Ferroni P, Basili S, Martini F, Cardarelli CM, Ceci F, di Franco M, et al. Serum metalloproteinase 9 levels in patients with coronary artery disease: a novel marker of inflammation. *J Investig Med*. 2003;51:295-300.
63. Axisa B, Loftus IM, Naylor AR, Goodall S, Jones L, Bell PR, et al. Prospective, randomized, double-blind trial investigating the effect of doxycycline on matrix metalloproteinase expression within atherosclerotic carotid plaques. *Stroke*. 2002;33:2858-64.
64. Arno G, Kaski JC, Smith DA, Akiyu JP, Hughes SE, Baboonian C. Matrix metalloproteinase-9 expression is associated with the presence of *Chlamydia pneumoniae* in human coronary atherosclerotic plaques. *Heart*. 2005;91:521-5.
65. Kalela A, Koivu TA, Sisto T, Kanervisto J, Hoyhtya M, Sillanaukee P, et al. Serum matrix metalloproteinase-9 concentration in angiographically assessed coronary artery disease. *Scand J Clin Lab Invest*. 2002;62:337-42.
66. Noji Y, Kajinami K, Kawashiri MA, Todo Y, Horita T, Nohara A, et al. Circulating matrix metalloproteinases and their inhibitors in premature coronary atherosclerosis. *Clin Chem Lab Med*. 2001;39:380-4.
67. Tayebjee MH, Tan KT, MacFadyen RJ, Lip GY. Abnormal circulating levels of metalloprotease 9 and its tissue inhibitor 1 in angiographically proven peripheral arterial disease: relationship to disease severity. *J Intern Med*. 2005;257:110-6.
68. Wilson EM, Moainie SL, Baskin JM, Lowry AS, Deschamps AM, Mukherjee R, et al. Region- and type-specific induction of matrix metalloproteinases in post-myocardial infarction remodeling. *Circulation*. 2003;107:2857-63.
69. Blankenberg S, Rupprecht HJ, Poirier O, Bickel C, Smieja M, Hafner G, et al. Plasma concentrations and genetic variation of matrix metalloproteinase 9 and prognosis of patients with cardiovascular disease. *Circulation*. 2003;107:1579-85.
70. Inokubo Y, Hanada H, Ishizaka H, Fukushi T, Kamada T, Okumura K. Plasma levels of matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-1 are increased in the coronary circulation in patients with acute coronary syndrome. *Am Heart J*. 2001;141:211-7.
71. Johnson JL, Baker AH, Oka K, Chan L, Newby AC, Jackson CL, et al. Suppression of atherosclerotic plaque progression and instability by tissue inhibitor of metalloproteinase-2: involvement of macrophage migration and apoptosis. *Circulation*. 2006;113:2435-44.
72. Kai H, Ikeda H, Yasukawa H, Kai M, Seki Y, Kuwahara F, et al. Peripheral blood levels of matrix metalloproteinases-2 and -9 are elevated in patients with acute coronary syndromes. *J Am Coll Cardiol*. 1998;32:368-72.
73. Lindholt JS, Vammen S, Fasting H, Henneberg EW, Heickendorff L. The plasma level of matrix metalloproteinase 9 may predict the natural history of small abdominal aortic aneurysms. A preliminary study. *Eur J Vasc Endovasc Surg*. 2000;20:281-5.
74. Longo GM, Xiong W, Greiner TC, Zhao Y, Fiotti N, Baxter BT. Matrix metalloproteinases 2 and 9 work in concert to produce aortic aneurysms. *J Clin Invest*. 2002;110:625-32.

75. Montaner J, Molina CA, Monasterio J, Abilleira S, Arenillas JF, Ribo M, et al. Matrix metalloproteinase-9 pretreatment level predicts intracranial hemorrhagic complications after thrombolysis in human stroke. *Circulation*. 2003;107:598-603.
76. Timms PM, Mannan N, Hitman GA, Noonan K, Mills PG, Syndercombe-Court D, et al. Circulating MMP9, vitamin D and variation in the TIMP-1 response with VDR genotype: mechanisms for inflammatory damage in chronic disorders? *QJM*. 2002;95:787-96.
77. Orbe J, Montero I, Rodríguez JA, Beloqui O, Roncal C, Paramo JA. Independent association of matrix metalloproteinase-10, cardiovascular risk factors and subclinical atherosclerosis. *J Thromb Haemost*. 2007;5:91-7.
78. Bendeck MP. Targeting pericellular proteolysis in vascular disease. *Circ Res*. 2002;91:861-2.
79. Host NB, Hansen SS, Jensen LT, Husum D, Nielsen JD. Thrombolytic therapy of acute myocardial infarction alters collagen metabolism. *Cardiology*. 1994;85:323-33.
80. Luan Z, Chase AJ, Newby AC. Statins inhibit secretion of metalloproteinases-1, -2, -3, and -9 from vascular smooth muscle cells and macrophages. *Arterioscler Thromb Vasc Biol*. 2003;23:769-75.
81. Cipollone F, Fazio M, Iezzi A, Zucchelli M, Pini B, de Cesare D, et al. Suppression of the functionally coupled cyclooxygenase-2/prostaglandin E synthase as a basis of simvastatin-dependent plaque stabilization in humans. *Circulation*. 2003;107:1479-85.
82. Cipollone F, Fazio M, Iezzi A, Pini B, Cucurullo C, Zucchelli M, et al. Blockade of the angiotensin II type 1 receptor stabilizes atherosclerotic plaques in humans by inhibiting prostaglandin E2-dependent matrix metalloproteinase activity. *Circulation*. 2004;109:1482-8.
83. Ohtsuka T, Hamada M, Saeki H, Ogimoto A, Hara Y, Shigematsu Y, et al. Serum levels of matrix metalloproteinases and tumor necrosis factor-alpha in patients with idiopathic dilated cardiomyopathy and effect of carvedilol on these levels. *Am J Cardiol*. 2003;91:1024-7.
84. Haffner SM, Greenberg AS, Weston WM, Chen H, Williams K, Freed MI. Effect of rosiglitazone treatment on nontraditional markers of cardiovascular disease in patients with type 2 diabetes mellitus. *Circulation*. 2002;106:679-84.
85. Galis ZS, Johnson C, Godin D, Magid R, Shipley JM, Senior RM, et al. Targeted disruption of the matrix metalloproteinase-9 gene impairs smooth muscle cell migration and geometrical arterial remodeling. *Circ Res*. 2002;91:852-9.
86. Pyo R, Lee JK, Shipley JM, Curci JA, Mao D, Ziporin SJ, et al. Targeted gene disruption of matrix metalloproteinase-9 (gelatinase B) suppresses development of experimental abdominal aortic aneurysms. *J Clin Invest*. 2000;105:1641-9.
87. Cheng L, Mantile G, Pauly R, Nater C, Felici A, Monticone R, et al. Adenovirus-mediated gene transfer of the human tissue inhibitor of metalloproteinase-2 blocks vascular smooth muscle cell invasiveness in vitro and modulates neointimal development in vivo. *Circulation*. 1998;98:2195-201.
88. Silence J, Collen D, Lijnen HR. Reduced atherosclerotic plaque but enhanced aneurysm formation in mice with inactivation of the tissue inhibitor of metalloproteinase-1 (TIMP-1) gene. *Circ Res*. 2002;90:897-903.
89. Mukherjee R, Brinsa TA, Dowdy KB, Scott AA, Baskin JM, Deschamps AM, et al. Myocardial infarct expansion and matrix metalloproteinase inhibition. *Circulation*. 2003;107:618-25.