TITLE: Evaluation of the nutritional aspects and cholesterol oxidation products of pork liver and fish patés.

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ABSTRACT

A comparative study between traditional patés elaborated with pork liver and fish patés (salmon, anchovy and cod) was carried out. The nutritional value and their security related to cholesterol oxidation products (COP) content were evaluated. Salmon paté showed similar fat content (24-28%) and energetic value (300Kcal/100g) to pork liver patés, whereas patés made with anchovy and cod showed less fat (13-16%) and calories (200-236 Kcal/100g).

PUFA/SFA ratios were much higher in all fish patés (1.55-4.95) than in liver pork patés (0.36-0.44). No great differences were found in \( \omega-6/\omega-3 \) ratio between salmon and pork liver patés (11.34-18.4), being even much higher this ratio in anchovy (32.32) and cod patés (62.77). EPA and DHA supply was around 0.63 for salmon, 0.21 for anchovy and 0.07 for cod patés. Cholesterol amounts were lower in fish patés (31-37mg/100g) than in pork liver patés (77-102mg/100g). Total COP ranged 0.38-2.83ppm, without clear differences between pork liver and fish patés.

**Keywords**: COP, salmon, anchovy, cod, canned foodstuffs

**Running title**: Nutritional and safety analysis of pork liver and fish patés
INTRODUCTION

Paté is a cooked product with important gastronomic tradition and appreciated sensory properties. Traditionally they have been elaborated with goose liver (“foie-grass”) or pork liver. However, during the last years a great variety of new products have been launched on the market, with special interest for those including fish, due to the nutritional advantages shown by these products.

Epidemiological studies carried out by Dyerberg and Bang (1979) in the 70’s showed the low incidence of cardiovascular pathologies in the Skimo population, great fish consumers. This fact was related to the abundance of w-3 long chain fatty acids in the fish fat, being eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) the most relevant ones. Further works demonstrated that the beneficial effects of these fatty acids are due to a complex interaction of a number of mechanisms such as the reduction of the plasma levels of triglycerides, the decrease in the platelet aggregation, their antiarrhythmic effect and their beneficial effect on the endothelial dysfunction (Harris, 1989; Von Shacky, 2000; Kang & Leaf, 2000; DeCaterina, Liao, & Libby, 2000; Goodfellow, Bellamy, Ramsey, Jones & Lewis, 2000).

Consequently, many studies show the positive effect of the w-3 fatty acids in relation to the prevalence of rheumatic arthritis, cancer, development of metastasis, hypertension and cardiac arrhythmia (Caygill & Hill, 1995; Simopoulos, 1997). Furthermore, an increased intake of w-3 polyunsaturated fatty acids from fish may have substantial implications for public health and health economy by decreasing the risk of coronary events and sudden cardiac death (Schmidt, Skou, Christensen & Dyerberg, 2000).

Other epidemiological studies show that increasing fish intake from one to two servings per week to five to six servings per week does not substantially reduce the risk of coronary heart disease among men who are initially free of cardiovascular disease.
(Ascherio, Rimm, Stampfer, Giovannucci & Willet, 1995). However, this higher intake has been positively correlated with a reduction on the risk of mortality in patients who have already had a myocardial stroke (Hu, Manson & Willet, 2001). Furthermore, there is evidence that the incorporation of a dietary supplement containing EPA and DHA reduces significantly the risk factors for cardiovascular disease (Yam, Bott-Kanner, Genin, Shinitzky & Klainman, 2002).

Other type of compounds clearly involved in the development of cardiovascular diseases, in this case with a negative effect, are cholesterol oxides (Smith, 1996). Some experiments in humans show their ability to be absorbed from the diet (Emanuel, Hassel, Addis, Bergmann & Zavoral, 1991; Linseisen & Wolfram, 1998). Many undesirable biological effects as cytotoxicity, mutagenicity, carcinogenicity, angiototoxicity, atherogenicity and cell membrane damage have been attributed to them (Guardiola, Codony, Addis, Rafecas & Boatella, 1996; Tai, Chen & Chen, 1999).

Yan (1999) reviewing the COP formation in foodstuffs concluded about the need of re-examine processing conditions in order not only to produce organoleptically superior products but also products with low levels of COP. Tai, Chen and Chen (2000) in a review about the presence of cholesterol oxides in different food concluded that these compounds are formed in most processed food containing cholesterol, being difficult to prevent their formation. Heating, which is the main technological step for the adequate elaboration and preservation of cooked products, including paté, is one of the main causes of their synthesis.

The presence of unsaturated fatty acids enhances cholesterol oxidation, through the development of free radicals and peroxides during heating (Korytowski, Bachowski & Girotti, 1992; Osada, Kodama, Cui, Yamada & Sugano, 1993). As fish fat is characterised by higher proportions of long chain unsaturated fatty acids than other food
products, is more susceptible to oxidation (Tai et al., 2000). Osada et al. (1993) demonstrated that levels of cholesterol oxides in processed fish products were higher to those found in other processed food products, such as dairy products or egg derivatives. The objective of this work was to study the lipid fraction of different samples of pork liver paté and fish paté, taking into account two points of view: the potential nutritional advantages of fish patés related to pork liver patés and the evaluation of the intensity of the cholesterol oxidation processes in these products.
MATERIAL and METHODS

Commercial samples

Six different types of patés were purchased from local supermarkets. Three of them corresponded to traditional products elaborated with liver pork. The rest were patés elaborated with different fish species: salmon, anchovy and cod.

Ingredients used in the patés (according to the respective labelling):

Pork liver patés. Trade 1: pork liver, pork backfat, pork fat and meat, water, milk protein, rice flour, salt, stabilisers (carragenates, disodium diphosphate), spices, taste enhancer (monosodium glutamate), potassium sorbate, E-217, sodium nitrite, potassium nitrate and flavourings. Trade 2: pork backfat, liver and meat, water, wheat flour, salt, species, stabilizer (E-450), sugar, taste enhancer (E-621), antioxidant (E-330) and conservant (E-250). Trade 3: pork liver, pork backfat, pork meat, water, wheat flour, salt, milk protein, antioxidant (E-330) and conservant (E-250).

Fish patés. Salmon paté: salmon (40%), skimmed milk, proteins and vegetable oils, dewlap, salt, spices, stabilizers (E-407). Anchovy paté: anchovy (25%), milk, mashed potatoes, water, vegetable oil, protein, flour, flavourings, gelifiers (E-407, E-412, E-410 y E-415) and colorant (E-127). Cod paté: cod (37%), milk, water, vegetable oil, potato starch, salt, garlic, milk proteins, vegetable fiber, flavourings, stabiliser (carragenaan) and spices.

Chemical Analysis

Proximal analysis

Moisture was determined by desiccation (AOAC, 2002a). Protein was analysed using the Kjeldahl method for the determination of nitrogen (AOAC, 2002b), using 6.25 as the factor to transform nitrogen in protein. Fat was determined by the Soxhlet method.
with petroleum ether (AOAC, 2002c). Ashes were determined by incineration using the method of the AOAC (AOAC, 2002d).

Analysis of fatty acids
Quantitative fat extraction was made for subsequent analysis of fatty acids with a chloroform/methanol mixture using the method of Folch, Lees and Sloane-Stanley (1957). The fatty acid profile was determined by gas chromatography previous methylation with BF$_3$/methanol (AOAC, 2002e). Chromatographic conditions used were as described by Muguerza, Gimeno, Ansorena, Bloukas and Astiasarán (2001).

0.5μl of the sample were injected in a gas chromatograph with a FID detector (Perkin-Elmer Autosystem XL) fitted with a capillary column SP$^{TM}$-2560 (100 m x 0.25 mm x 0.2 μm). The temperature of both the injection port and detector was 220°C. The oven temperature was 165°C during 80 minutes, followed by an increase to 220°C at a rate of 4°C/minute and 50 minutes at 220°C. The carrier gas was hydrogen (20 psi). The identification of the peaks was made by comparison of their retention times to those of pure standard compounds (Sigma, St. Louis, MO, USA) and the quantification of individual fatty acids was based on heptadecanoic acid methyl ester as internal standard.

Cholesterol determination
The determination of cholesterol was made by gas chromatography, according to the method described by Kovacs, Anderson and Ackman (1979). A Perkin-Elmer Autosystem gas chromatograph equipped with an HP1 column (30 m x 0.25 mm x 0.1 μm) was used. The oven temperature was 265°C. The temperature of both the injection port and detector was 285°C. The sample size was 0.5 μl. Cholesterol was identified by comparing its relative and absolute retention times with those of cholestane (Sigma, St. Louis, MO, USA) as an internal standards. A Perkin-Elmer Turbochrom programme was used for quantification.
Cholesterol oxides determination

Cholesterol oxides were determined using the method described by Echarte et al. (2001). Cold saponification was performed, a further extraction of the cholesterol oxides with ether, a purification with silica cartridges and a derivatization to obtain the trimethyl silyl ethers. 1μl of sample was injected in a gas chromatograph HP 6890GC System (Hewlett-Packard) coupled to a 5973 Mass Selective Detector (Hewlett-Packard). The column used was HP-5MS (30m x 250μm x 0.25μm) and helium as carrier gas (1ml/minute). Oven temperature was initially 80ºC, held for 1 minute, and programmed to 250ºC at a rate of 10ºC/minute, and final column temperature of 280ºC at a rate of 4ºC/minute, and held for 20 minutes. The injector temperature was 250ºC and the inlet pressure was 23.2psig; mass range was 50/550; solvent delay was 20 minutes. Identification of the peaks was made by comparison of the mass spectra obtained for every pure compound to those of the sample. Quantification was made using 19-hydroxycolesterol as internal standard (Sigma, St. Louis, MO, USA).

Data Analysis

Four samples were analysed from every trade of paté. Each parameter was determined four times in each sample. Data shown in the tables are the mean (n=16) with standard deviations. Statistical test one-way ANOVA and a Tuckey’s b posteriori test were used to analyse statistical differences between samples (p≤0.05). Pearson correlation was determined among different parameters. Software used was SPSS 9.0 for Windows.
DISCUSSION

Patés elaborated with liver and fat from pork or goose (foie-gras) are usually, according to traditional food composition tables, highly energetic products (around 300-400 kcal/100g) with relatively high percentages of lipids (30-40%) (McCance & Widdowson’s, 1992; Mataix, 1994; Senser & Scherz, 1999; Moreiras, Carbajal, Cabrera & Cuadrado, 2001). Samples from the three commercial trades containing pork liver analysed in this work showed lipid concentrations of about 24-28%, leading to calorific values that ranged between 285 and 320 kcal/100g. Comparing these data with the obtained for fish patés it can be seen that only salmon patés showed similar amounts of fat (26%), giving rise to the highest calorific value between the fish patés. Patés made with anchovy or cod showed 10-13% less fat percentages (16.10 and 13.72%, respectively) and around 70-100 kcal less per 100g (236 and 200 Kcal/100g, respectively) than the rest of patés. These differences could be due to the different proportions of fish employed in the formulation, the different fat content of the species and the different amounts of other fat supplying ingredients (vegetable oils). Labels of fish patés show that they were elaborated with 40% salmon, 25% anchovy and 37% cod, respectively. Furthermore, these species differ substantially in their fat content. Composition tables stand up values of fat around 12% for salmon, 5% for anchovy and 0.3% for cod (Belitz & Grosch, 1988; MacCance & Widdoson’s, 1992).

It is known that every fish species have similar protein amount, being inversely correlated their content in fat and water. No significant differences were observed among the three types of fish patés in the protein amount, ranging values from 7.51 to 8.62%. This protein comes not only from the fish, but also from other proteic ingredients (milk and additives). Meat samples showed a greater variability in their protein content, ranging values from 8.38 to 14.5%. This last value is similar to that
observed in the composition tables (14%). The water content did not show significant
differences between meat and fish patés except to the cod paté which showed the
highest amount. Carbohydrates (which were determined by difference) were present in
greater amounts in fish patés.
Values found for the total mineral content (ashes) ranged between 2.21 and 2.72% in
meat samples. This type of food is considered as a good source of some minerals: Na
(present as a consequence of using ClNa in the elaboration), Fe (supplied by the liver as
the main ingredient), Ca, Mg and Zn. The values obtained for the fish patés were
similar, except for the anchovy paté, whose ashes’ content was double. This higher
content for ashes was also detected in other commercial anchovy’s patés (Aquerreta,
Astiasarán, Mohino & Bello, 2002), and could be explained by the presence of the
bones of this little fish in the final product.
Together with the calorific value, the relatively high supply of cholesterol is the main
drawback of the presence of meat products in the diet. Values obtained for the pork liver
patés ranged between 77.61mg/100g and 101.92mg/100g, clearly lower than those
found in the literature for this type of products (255mg/100g). Chizzolini, Zanardi,
Dorigoni and Ghidinim (1999) in a review about the cholesterol content of meat and
meat products established that the cholesterol content ranged from 60 to 99 mg/100g for
meat and from 37 to 110mg/100g for meat products. Fish patés did not show significant
differences in their cholesterol content, with values ranging from 31.38 to
36.87mg/100g. These similar results among fish patés in spite of the different fat
content of the species can be explained by the supply of cholesterol by other ingredients
such as milk. Salmon pate, the one with the highest fish fat supply was elaborated with
skimmed milk, whereas anchovy and cod patés were elaborated with whole milk which
obviously supply cholesterol. The lower amounts of cholesterol in fish patés in relation
to pork liver patés, specially salmon patés which show similar fat content, could be explained by the fact that vegetable oils contribute to the total fat content but not to the total cholesterol content. Pork liver patés did not include vegetable oils.

The last epidemiological studies conclude that the type of fat supplied in the diet is an important factor in relation with health. It has been proved that the substitution of saturated fat by unsaturated fat is more effective in the decrease of risk of cardiovascular disease than the only reduction of total fat intake (Hu et al., 2001). Furthermore, the influence of the different types of saturated fatty acids (SFA) on the cholesterol levels and risk of cardiovascular disease has been determined.

Traditionally in the nutritional evaluation of the lipid fraction of food stearic acid (C18:0) had been excluded from the SFA fraction, because it does not raise plasma cholesterol levels as the rest of the SFA do (Tholstrup, Marckmann, Jespersen, & Sandstrom, 1994; Candela, Astiasarán, & Bello, 1997; Zapelena, Aquerreta, Astiasarán & Bello, 1995). However, it decreases the HDL fraction, so it seems that the exclusion is not completely justified (Hu et al., 2001) and in our work it has been included in the total amount of SFA fraction.

Palmitic acid was the most abundant one in the saturated fraction in every pork liver paté, followed by the stearic acid (table 2), highly contributing to the total amount of this fraction, whose values ranged between 6.61g/100g and 8.07g/100g (table 5). Myristic and lauric acids, presumably those with the strongest influence on rising the cholesterol levels (Kris-Etherton & Yu, 1997) supplied about 0.3g to the total SFA fraction. The values obtained in the fish patés for the SFA fraction were lower, particularly in the anchovy patés (1.4g/100g) and in the cod patés (1.8g/100g).

Oleic acid was the most abundant fatty acid in the pork liver patés. This monounsaturated fatty acid was the main responsible of MUFA being this the most
abundant fraction in these patés (8.6 – 10.4 g/100g). Similar data was found for salmon paté (9.83g MUFA/100g), whereas cod and specially anchovy patés showed much lower amounts of oleic acid. The intake of MUFA has been inversely associated to risk of CHD, although the association was weaker than for polyunsaturated fat (Hu et al., 1997).

Fish patés showed higher total PUFA amounts than pork liver patés. The main reason was probably the use of seed vegetable oils, which are a good source of omega-6 PUFA, particularly linoleic acid. Great differences were found in the linoleic content of fish and meat products. In relation with the potential omega-3 supply of fish patés only salmon patés showed significant differences. Total omega-3 PUFA of salmon reached 0.75g/100g sample, in contrast with ranges between 0.09 and 0.26 found in the rest of patés. In pork liver patés the main omega-3 PUFA was linolenic acid, which is also found in high amounts in salmon patés probably as a consequence of the use of dewlap in its formulation or even by the use of a seed vegetable oil with a higher proportion of this acid. EPA and DHA showed the highest amounts in salmon (0.63g/100g) and anchovy (0.21g/100g) patés, the ones with the highest fish fat supply. However, cod patés (0.07g/100g) did not show significant differences in the content of these fatty acids in relation to the pork liver patés (0.02-0.04g/100g). Omega-6/omega-3 ratios were high in all samples and specially in anchovy and cod paté, which was due to the combination of a high amount of linoleic acid supplied by vegetable oils and the low amounts of omega-3 fatty acids, especially in the case of the cod paté. Aquerreta et al. (2002) observed that experimental fish patés elaborated without vegetable oils showed lower omega-6/omega-3 ratios than commercial fish patés.
All this values gave rise to clear higher ratios PUFA/SFA for the fish patés, particularly in the anchovy paté (4.95), with regard to the pork liver patés, that showed values lower than 0.45.

The influence of the trans fatty acids on cardiovascular health has been widely demonstrated (Hu et al., 2001; Salmerón et al., 2001). High intakes of this type of fatty acids have been associated with a stronger risk of cardiovascular diseases, mediated by a number of different mechanisms: increment of LDL cholesterol, decrease of HDL cholesterol, increment of lipoprotein A levels, increment of plasma triglycerides, etc.

Neither the pork liver patés nor the fish patés showed concentrations higher than 0.3g/100g. This amounts can be considered as low taking into account that for example, a medium portion of French fries contains 5-6g trans fatty acids/100g or a donuts contains 2g trans fatty acids /100g (Katan, 2000).

Technological conditions applied during the elaboration of the patés (specially heating intensity) and the different fatty acid profiles of the samples would explain the differences in the intensity of COP formation. Total COP ranged from 38 to 283μg/100g (0.38 to 2.83 ppm). Percentages of oxidation ranged between 0.05 and 0.73, being the last one that of cod paté (table 4). The amounts of COP found in the different analysed samples were not correlated with their cholesterol content neither with the unsaturated fatty acids content. Not many references have been found in the bibliography about COP content in commercial meat products. In dry-cured ham with 12-24 months of ageing total COP ranged 2.8-5.8 ppm (Vestergaard & Parolari, 1999). Zunin, Boggia and Evangelisti (2001) found that COP values amounted to 0.3-3 μg/g in canned tuna. Novelli et al. (1998) found a very large range of total COP amount in samples of salame Milano and mortadela (0-30.93 ppm). These authors stated up that the considered as most toxic COP, 25-hydroxycholesterol and colestanetriol, were
rarely observed in the analysed samples. Wang, Yang-Nian. and Chin-Wen (1995) analysing lipid and cholesterol oxidation in Chinese-style sausages neither detected 25-hydroxycholesterol nor cholestanetriol. In the analysed patés 25-hydroxycholesterol was only detected in one of the pork liver paté, whereas cholestanetriol appeared in all the fish samples and in one of the pork liver paté, always at amounts lower than 0.27 ppm. 7-ketocholesterol, considered to be a good indicator of oxidation (Zunin, Calcagno and Evangelisti, 1998; Penazzi et al., 1995), was present in all analysed samples, except for those of anchovy pate. This compound and the 7α-hydroxycholesterol showed significantly correlations (0.780 and 0.926) with total COP.

In summary, it can be stated up that the potential nutritional advantages of fish patés in relation with the classical pork liver patés depends largely on the species and the rest of the ingredients. Only significant increments in EPA and DHA were observed in salmon patés in relation to pork liver patés, being omega-6 / omega-3 ratio even much higher in anchovy and cod patés than in pork liver patés. Although cholesterol amounts were lower in fish patés than in pork liver patés, no clear differences were observed in the COP amounts.
ACKNOWLEDGEMENTS

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Official Analytical Chemists.


RESULTS

Table 1. General composition (g/100g product), cholesterol content (mg/100g product) and calorific value (Kcal/100g) of the analysed samples.

<table>
<thead>
<tr>
<th></th>
<th>Pork Liver</th>
<th>Fish Patés</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Paté 1</td>
<td>Paté 2</td>
</tr>
<tr>
<td>Moisture</td>
<td>57.06 ± 0.46&lt;sup&gt;a&lt;/sup&gt;</td>
<td>52.46 ± 0.31&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>Protein</td>
<td>9.93 ± 0.65&lt;sup&gt;b&lt;/sup&gt;</td>
<td>14.50 ± 1.03&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Fat</td>
<td>26.18 ± 0.88&lt;sup&gt;c&lt;/sup&gt;</td>
<td>28.05 ± 0.52&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
<tr>
<td>Ashes</td>
<td>2.21 ± 0.21&lt;sup&gt;a&lt;/sup&gt;</td>
<td>2.72 ± 0.24&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Carbohydrates (*)</td>
<td>4.62</td>
<td>2.27</td>
</tr>
<tr>
<td>Calorific value</td>
<td>294</td>
<td>320</td>
</tr>
</tbody>
</table>

Different letters in the same row show significant differences among samples (p<0.05).
(*) Calculated by difference.
<table>
<thead>
<tr>
<th></th>
<th>Pork Liver</th>
<th>Paté 1</th>
<th>Paté 2</th>
<th>Paté 3</th>
<th>Salmon</th>
<th>Anchovy</th>
<th>Cod</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lauric 12:0</td>
<td>0.02 (0.00)^b</td>
<td>0.03 (0.00)^c</td>
<td>0.02 (0.00)^b</td>
<td>0.02 (0.00)^b</td>
<td>0.01 (0.00)^a</td>
<td>0.02 (0.00)^b</td>
<td></td>
</tr>
<tr>
<td>Mirystic 14:0</td>
<td>0.29 (0.02)^c</td>
<td>0.31 (0.01)^c</td>
<td>0.26 (0.01)^b</td>
<td>0.31 (0.02)^c</td>
<td>0.04 (0.00)^a</td>
<td>0.06 (0.00)^a</td>
<td></td>
</tr>
<tr>
<td>Palmitic 16:0</td>
<td>5.04 (0.4)^d</td>
<td>4.88 (0.05)^d</td>
<td>4.12 (0.06)^c</td>
<td>3.69 (0.16)^b</td>
<td>0.80 (0.07)^a</td>
<td>1.08 (0.03)^b</td>
<td></td>
</tr>
<tr>
<td>Estearic 18:0</td>
<td>2.61 (0.24)^d</td>
<td>2.51 (0.05)^d</td>
<td>2.11 (0.02)^c</td>
<td>1.78 (0.06)^b</td>
<td>0.50 (0.06)^a</td>
<td>0.49 (0.01)^d</td>
<td></td>
</tr>
<tr>
<td>Arachidic 20:0</td>
<td>0.05 (0.01)^ab</td>
<td>0.05 (0.01)^ab</td>
<td>0.05 (0.01)^ab</td>
<td>0.06 (0.01)^b</td>
<td>0.03 (0.00)^a</td>
<td>0.04 (0.00)^d</td>
<td></td>
</tr>
<tr>
<td>Behenic 22:0</td>
<td>0.06 (0.03)^a</td>
<td>0.06 (0.01)^a</td>
<td>0.06 (0.01)^a</td>
<td>0.12 (0.04)^a</td>
<td>0.09 (0.07)^c</td>
<td>0.11 (0.02)^a</td>
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</tr>
<tr>
<td>Palmitoleic 16:1</td>
<td>0.54 (0.04)^c</td>
<td>0.54 (0.01)^c</td>
<td>0.44 (0.01)^b</td>
<td>0.57 (0.03)^c</td>
<td>0.03 (0.00)^a</td>
<td>0.05 (0.00)^a</td>
<td></td>
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<tr>
<td>Oleic 18:1</td>
<td>9.22 (0.74)^d</td>
<td>9.81 (0.17)^d</td>
<td>8.08 (0.07)^c</td>
<td>8.42 (0.30)^c</td>
<td>3.41 (0.37)^a</td>
<td>5.33 (0.24)^b</td>
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<tr>
<td>Erucic 22:1</td>
<td>0.04 (0.02)^ab</td>
<td>0.05 (0.00)^b</td>
<td>0.05 (0.02)^b</td>
<td>0.24 (0.01)^c</td>
<td>0.04 (0.00)^ab</td>
<td>0.02 (0.00)^a</td>
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<tr>
<td>Linoleic 18:2 W_6</td>
<td>2.65 (0.26)^a</td>
<td>3.11 (0.05)^a</td>
<td>2.52 (0.07)^d</td>
<td>8.51 (0.35)^d</td>
<td>7.06 (0.71)^c</td>
<td>5.65 (0.29)^b</td>
<td></td>
</tr>
<tr>
<td>Linolenic 18:3 W_3</td>
<td>0.20 (0.03)^c</td>
<td>0.22 (0.02)^c</td>
<td>0.12 (0.01)^b</td>
<td>0.12 (0.07)^b</td>
<td>0.01 (0.00)^a</td>
<td>0.02 (0.00)^a</td>
<td></td>
</tr>
<tr>
<td>Arachidonic 20:4 W_6</td>
<td>0.04 (0.00)^a</td>
<td>0.04 (0.00)^a</td>
<td>0.02 (0.01)^b</td>
<td>0.00 (0.00)^a</td>
<td>0.05 (0.00)^a</td>
<td>0.00 (0.00)^a</td>
<td></td>
</tr>
<tr>
<td>Eicosapentaenoic EPA 20:5 W_3</td>
<td>0.01 (0.00)^a</td>
<td>0.01 (0.00)^a</td>
<td>0.01 (0.00)^a</td>
<td>0.28 (0.01)^c</td>
<td>0.05 (0.00)^b</td>
<td>0.02 (0.00)^a</td>
<td></td>
</tr>
<tr>
<td>Docosahexaenoic DHA 22:6 W_3</td>
<td>0.03 (0.00)^b</td>
<td>0.03 (0.00)^b</td>
<td>0.01 (0.00)^a</td>
<td>0.35 (0.02)^c</td>
<td>0.16 (0.00)^d</td>
<td>0.05 (0.00)^b</td>
<td></td>
</tr>
<tr>
<td>Palmitelaidic trans 16:1</td>
<td>0.02 (0.00)^a</td>
<td>0.11 (0.00)^b</td>
<td>0.01 (0.00)^a</td>
<td>0.02 (0.00)^b</td>
<td>0.00 (0.00)^a</td>
<td>0.00 (0.00)^a</td>
<td></td>
</tr>
<tr>
<td>Elaidic trans 18:1</td>
<td>0.05 (0.01)^b</td>
<td>0.04 (0.00)^b</td>
<td>0.04 (0.01)^b</td>
<td>0.03 (0.01)^b</td>
<td>0.01 (0.00)^a</td>
<td>0.01 (0.00)^a</td>
<td></td>
</tr>
<tr>
<td>Linolelaidic trans 18:2</td>
<td>0.02 (0.01)^c</td>
<td>0.01 (0.00)^bc</td>
<td>0.01 (0.00)^bc</td>
<td>0.01(0.00)^abc</td>
<td>0.01 (0.00)^ab</td>
<td>0.00 (0.00)^a</td>
<td></td>
</tr>
<tr>
<td>Brassidic trans 22:1</td>
<td>0.12 (0.01)^b</td>
<td>0.13 (0.01)^b</td>
<td>0.03 (0.02)^a</td>
<td>0.03 (0.01)^b</td>
<td>0.01 (0.00)^a</td>
<td>0.01 (0.00)^a</td>
<td></td>
</tr>
</tbody>
</table>

Different letters in the same row show significant differences among samples (p<0.05).
Table 3. Different fatty acid fractions and important nutritional ratios.

<table>
<thead>
<tr>
<th></th>
<th>Pork Liver</th>
<th>Fish Patés</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Paté 1</td>
<td>Paté 2</td>
</tr>
<tr>
<td>Σ SFA</td>
<td>8.07</td>
<td>7.85</td>
</tr>
<tr>
<td>Σ MUFA</td>
<td>9.8</td>
<td>10.4</td>
</tr>
<tr>
<td>Σ PUFA</td>
<td>2.91</td>
<td>3.42</td>
</tr>
<tr>
<td>PUFA / SFA</td>
<td>0.36</td>
<td>0.44</td>
</tr>
<tr>
<td>Σ W₆</td>
<td>2.69</td>
<td>3.15</td>
</tr>
<tr>
<td>Σ W₃</td>
<td>0.24</td>
<td>0.26</td>
</tr>
<tr>
<td>W₆ / W₃</td>
<td>11.20</td>
<td>12.11</td>
</tr>
<tr>
<td>Σ Trans</td>
<td>0.21</td>
<td>0.29</td>
</tr>
</tbody>
</table>

*SFA: saturated fatty acids; MUFA: monounsaturated fatty acids; PUFA: polyunsaturated fatty acids.*
Table 4. Content of cholesterol (mg/100g product) and cholesterol oxides (μg/100g product).

<table>
<thead>
<tr>
<th></th>
<th>Pork liver</th>
<th>Fish Patés</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Paté 1</td>
<td>Paté 2</td>
</tr>
<tr>
<td>Cholesterol</td>
<td>77.61 ± 2.98&lt;sup&gt;b&lt;/sup&gt;</td>
<td>101.9 ± 11.9&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>7α-Hydroxycholesterol</td>
<td>0&lt;sup&gt;a&lt;/sup&gt;</td>
<td>38 ± 9&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>7β-Hydroxycholesterol</td>
<td>0&lt;sup&gt;a&lt;/sup&gt;</td>
<td>30 ± 6&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>25-Hydroxycholesterol</td>
<td>0&lt;sup&gt;a&lt;/sup&gt;</td>
<td>41 ± 1&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>7-Ketocholesterol</td>
<td>38 ± 7&lt;sup&gt;b&lt;/sup&gt;</td>
<td>32 ± 3&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Cholestanetriol</td>
<td>0&lt;sup&gt;a&lt;/sup&gt;</td>
<td>22 ± 2&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>α-Epoxycholesterol</td>
<td>0&lt;sup&gt;a&lt;/sup&gt;</td>
<td>24 ± 2&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Total Oxides</td>
<td>38</td>
<td>187</td>
</tr>
<tr>
<td>%Oxidation</td>
<td>0.05</td>
<td>0.18</td>
</tr>
</tbody>
</table>

Different letters in the same row show significant differences among samples (p<0.05).

%Oxidation = [(μg COP * 10<sup>-3</sup>) / mg cholesterol] * 100