

Biochemical biomarkers for multiple sclerosis

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ABSTRACT

Introduction: Multiple sclerosis (MS) is the most frequent demyelinating disease of the central nervous system. Although there is currently no definite cure for MS, new therapies have recently been developed based on a continuous search for new biomarkers.

Development: MS diagnosis relies on the integration of clinical, imaging and laboratory findings as there is still no single pathognomonic clinical feature or diagnostic laboratory biomarker. The most commonly laboratory test used is the presence of immunoglobulin G oligoclonal bands (OCB) in cerebrospinal fluid of MS patients. This test is now included in the 2017 McDonald criteria as a biomarker of dissemination in time. Nevertheless, there are other biomarkers currently in use such as kappa free light chain, which has shown higher sensitivity and specificity for MS diagnosis than OCB. In addition, other potential laboratory tests involved in neuronal damage, demyelination and/or inflammation could be used for detecting MS.

Conclusions: CSF and serum biomarkers have been reviewed for their use in MS diagnosis and prognosis to establish an accurate and prompt MS diagnosis, crucial to implement an adequate treatment and to optimize clinical outcomes over time.

1. Introduction

Multiple sclerosis (MS) is a chronic autoimmune inflammatory neurological disorder, classified as a demyelinating and degenerative disease that affects the central nervous system (CNS) [1]. Its prevalence has increased worldwide in recent years, with 2.5 million cases reported around the world and 700,000 in Europe [2,3]. Specifically, in Spain, the prevalence is considered medium-high, with 180 cases per 100,000

inhabitants [4].

MS is the most frequent demyelinating disease of the CNS and considered the leading cause of non-traumatic disability in young adults. Although it can occur throughout life, symptoms usually start at 25–30 years of age and mostly in women [5–7].

Most MS patients have initially presented a clinical isolated syndrome (CIS), defined as a single demyelinating event affecting the CNS of at least 24 h duration with no association to other organic disease

Abbreviations: APC, antigen-presenting cells; BBB, blood brain barrier; CIS, clinical isolated syndrome; CHI3L1, chitinase-3-like 1; CNS, central nervous system; CSF, cerebrospinal fluid; CXCL12, chemokine ligand 12 or stromal cell-derived factor; CXCL13, chemokine ligand 13 or B lymphocyte chemoattractant; DUSP, dual-specificity MAPK phosphatases; EBV, Epstein Barr virus; ECL, electrochemiluminescence; EDSS, expanded disability status scale; ELISA, enzyme-linked immunoassay; FLC, free light chain; GFAP, glial fibrillary acidic protein; HSP, heat shock protein; κFLC, Kappa free light chain; λFLC, Lambda free light chain; MAPK, mitogen-activated protein kinase; MBP, myelin basic protein; MOG, myelin oligodendrocyte glycoprotein; MRI, magnetic resonance imaging; MS, multiple sclerosis; Nfs, neurofilaments; NfH, Nf composed of heavy chains; NfL, Nf composed of light chains; NfM, Nf composed of medium chains; OCB, oligoclonal bands; OPN, osteopontin; PPMS, primary progressive multiple sclerosis; PRMS, progressive recurrent multiple sclerosis; RRMS, relapsing remitting multiple sclerosis; SPMS, secondary progressive multiple sclerosis; Treg cells, regulatory T cells; VCAM1, vascular cell adhesion molecule 1.

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(fever, infections, metabolic disorders, etc.) [8–10]. Although these patients can convert to Relapsing Remitting MS (RRMS), disease progression is highly variable with the majority characterized by recurrent onset of clinical symptoms with full or partial recovery after the flare.

Therefore, there have been described four MS types: RRMS, Secondary Progressive MS (SPMS) that leads to an irreversible progressive disability, Primary Progressive MS (PPMS) and Progressive Recurrent MS (PRMS) [11].

The disease may debut with flare-ups and different symptoms, depending on the location of the lesions in the CNS, commonly called plaques. A flare-up is the appearance of new neurological symptomatology within a period of at least 24 h or a significant deterioration of preexisting symptoms that had been stable or absent for at least 30 days [12]. These outbreaks can cause lesions at any CNS level, reflecting inflammatory activity and producing different symptoms such as fatigue, blurred vision or eye pain, weakness, coordination problems, sensory symptoms or several of them together. In that case, it is considered a multifocal outbreak [1,13].

Currently there is no definite cure for MS, although new therapies have recently been developed increasing the effectiveness of the treatment and improving the disability-free life expectancy [14]. More research and development of new biomarkers is essential to improve the diagnostic of the disease and therefore to initiate an early treatment to prevent disease progression. Thus, the main aim of this manuscript is to critically review the use of cerebrospinal fluid (CSF) and serum biomarkers to achieve the most accurate MS diagnosis and to evaluate its utility in follow-up and prognosis.

2. Etiology

The main cause of the disease is an unbalance in the autoimmune system [15], but its etiology and underlying pathogenic mechanisms are currently unknown. Although MS is not an inherited disease, a strong genetic component is involved in its etiology [2]. Also, environmental triggers have been observed [16].

2.1. Gene association

Different HLA II alleles could be involved in the development of MS. Populations with the HLA DRB1*1501-DQB1*0602 (HLA DRB2) haplotype are more susceptible to develop MS [17,18]. The full mechanism remains unknown, but it is believed that HLA DR2 has a specific binding gap for CNS self-antigens, which will be presented to T cells. Then the activated T cells may attack the CNS, thereby increasing the production of Th1 lymphocytes in the area, ultimately favoring CNS inflammation. This haplotype is found in 25–30 % of the Northern European and US population. Other genetic risk factors recently identified are interleukin-2 (IL-2) receptor alpha gene and interleukin-7 (IL-7) receptor alpha gene [18,19].

On the other hand, there is a HLA ALA genetic variant, HLA A02, which is the second most abundant one. However, HLA A02 protectively acts against the onset of the disease probably by eliminating viruses related to MS, such as Epstein Barr virus (EBV) [20].

Furthermore, the existence of different allelic variants increases the risk of MS over one allelic variant alone. An overactivity of mitogen-activated protein kinase (MAPK) pathways in microglia would be related to MS presentation and progression, classically related to inflammation [21,22]. MAPK ERK overactivation may cause downregulation of the Wnt/ β -catenin pathway, which leads to a microglial phenotype causing hypomyelination [23,24]. MAPK ERK also causes an overexpression of vascular cell adhesion molecule 1 (VCAM-1), a key adhesion molecule that induces the translocation of leukocytes to inflamed tissue [25]. A dual-specificity MAPK phosphatase (DUSP) is the negative feedback system that regulates MAPK pathways. An overactivity of MAPK pathways could occur when DUSP are downregulated [26]. This downregulation is caused by low serum vitamin D, smoking

habit [27] and prior EBV infection [15,28], all three known as risk factors for MS [29–32], despite the fact that their pathophysiological mechanisms still remain unclear [33,34] (Fig. 1).

2.2. Immunopathology

Homeostasis defects in regulatory T cells (Treg cells), defined as CD4 + CD25 + FoxP3 + T cells, have been associated with some allelic variants in cytokines or co-stimulatory molecules genes [35]. Reduced suppressive effect of Treg cells is known in autoimmune diseases. Also, CD4 + CD25 + Treg cells functional loss has been described in the suppression of T cell immune response. That allows effector CD4 + T cells to migrate into the CNS crossing the blood brain barrier (BBB) and eventually to destroy myelin oligodendrocyte glycoprotein in MS patients [35–38]. In addition, Treg cells are capable of secreting proinflammatory cytokines [39] (Fig. 1).

2.3. Environmental triggers

Other hypothesis is that certain viral or bacterial autoantigens or super-antigens could produce cross-reactions by binding to antigen-presenting cells (APCs). These would travel through the bloodstream and reach the lymphoid organs, where they would activate T lymphocytes. The activated T lymphocytes would trigger CD8 + T lymphocytes and other B-lymphocytes, which could cross the BBB. Then, once inside the CNS, they would be able to generate a cytotoxic effect, producing pro-inflammatory cytokines such as IFN- γ , TNF- α and IL-17. These cytokines would trigger the activation of macrophages and microglia, which, in turn, secrete cytokines such as IL-12 and IL-23 and chemokines. These will then be responsible for inducing the recruitment of other lymphocytes, such as other CD4 + and CD8 + T-lymphocytes, B-lymphocytes and monocytes, to the CNS. CD4 + T cells turn into Th1 lymphocytes through exposure to cytokines, such as IL-12, and towards Th17 by exposure mainly to IL-17 and IL-23. Finally, these cells promote demyelination and damage to oligodendrocytes and neurons [40] (Fig. 1).

In addition, low sun exposure results in low serum vitamin D, which is also associated with increased predisposition to MS development [30,41]. Several studies have demonstrated a correlation between low serum vitamin D and higher disability [41–47]. Also, evidence shows that the risk of subsequent relapses is lower in CIS subjects with higher serum vitamin D [48,49]. For that reason, multiple studies have evaluated the use of vitamin D supplementation as treatment in MS. However, the meta-analysis conducted by James et al. [50], on the risk of MS relapses after vitamin D supplementation, showed no significant association between high-dose vitamin D treatment and risk of MS relapses. Similar results were found by Pozuelo-Moyano et al. [51] who reported no evidence of vitamin D being a beneficial treatment in MS.

3. Diagnostic criteria

Misdiagnosis of MS remains an issue in clinical practice. There are numerous CNS diseases to make a differential diagnosis with [52], such as migraine, fibromyalgia, nonspecific symptoms with abnormal magnetic resonance imaging (MRI), functional neurological disorder and neuromyelitis optica [53]. McDonald criteria revised in 2017 are the most widely employed for MS diagnosis [54].

To establish a diagnosis of MS according to that criteria, an individual must have evidence of CNS damage disseminated in space, plaques need to be present in multiple regions of the nervous system and/or there should be evidence of damage disseminated in time, or occurring at different points in time [54] (Table 1).

Moreover, it has been shown that early therapeutic intervention delays long-term disease progression and improves outcomes, which can be graded using the expanded disability status scale (EDSS). Therefore, an accurate diagnosis is needed especially in those patients with

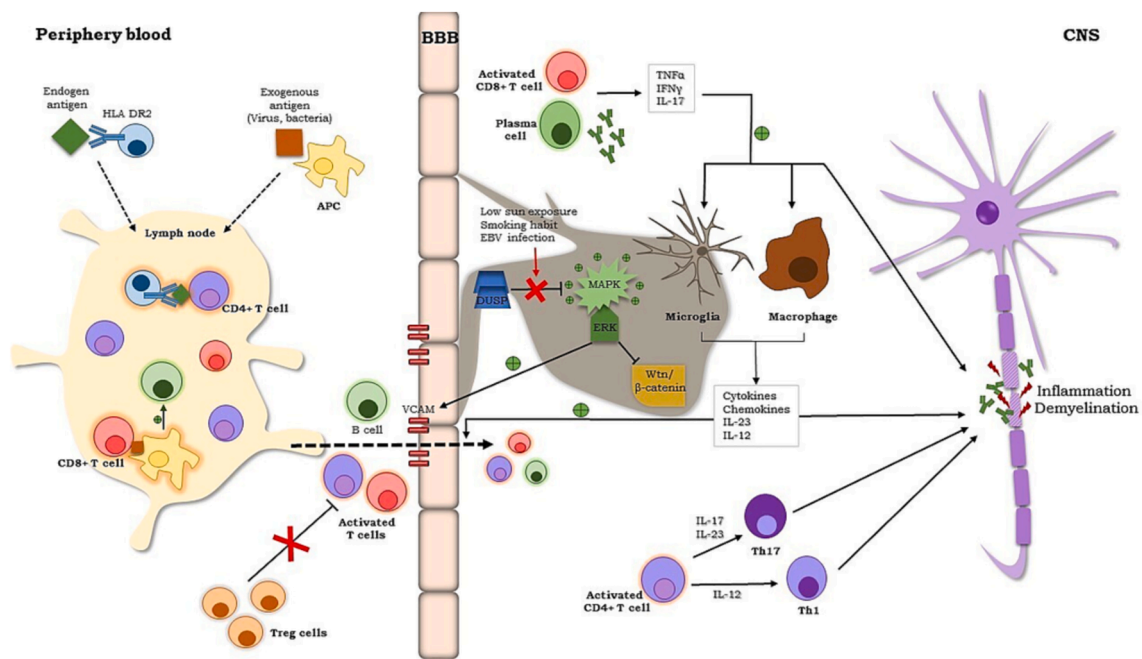


Fig. 1. Pathophysiological model of multiple sclerosis. APC, antigen-presenting cells; BBB, blood brain barrier; CNS, central nervous system; DUSP, dual-specificity MAPK phosphatases; MAPK, mitogen-activated protein kinase; Treg cells, regulatory T cells; VCAM1, vascular cell adhesion molecule 1.

neuromyelitis optica or CIS at high risk of developing RRMS, SPMS, or PPMS [55,56].

McDonald criteria include the use of MRI to establish the presence of disseminated lesions in space or time. In the latest update of these criteria, lesions in the cortical area of the brain were included, in addition to juxtacortical lesions, in order to consider dissemination in space. Furthermore, in addition to asymptomatic MRI lesions, symptomatic ones are currently taken into account when determining dissemination in space or time (except for lesions in the optic nerve in patients with neuromyelitis optica symptoms). According to the 2017 McDonald criteria the presence of IgG oligoclonal bands (OCB) in CSF can substitute the requirement of demonstrating dissemination in time. This improvement in the 2017 criteria compared to those of 2010, enables an early MS diagnosis in patients that meet the criteria for dissemination in space.

MS diagnosis relies on the integration of clinical, imaging, and laboratory findings, since there is still no single pathognomonic clinical feature or diagnostic laboratory biomarker that can effectively detect the disease. The most commonly laboratory test used is the presence of IgG OCB in CSF of MS patients, now included in 2017 McDonald criteria. Nevertheless, there are already in use biomarkers and proposed new serum and CFS biomarkers which could be helpful in diagnosis and prognosis of MS at different stages of the disease (Table 2).

4. Biomarkers used in clinical practice

4.1. IgG intrathecal synthesis. Oligoclonal bands and IgG index

The presence of higher IgG levels and IgG OCB in the CSF absent in serum are suggestive of intrathecal IgG synthesis by plasma cells and monoclonal B lymphocytes in the CNS [57]. Thereby, the detection of IgG OCB in CSF is considered the “gold standard” laboratory test to evidence elevated intrathecal synthesis. However, OCB detection is performed by isoelectric focusing technique followed by agarose gel electrophoresis. This is indeed a complex protocol with a high cost and methodological limitations such as the need of trained personnel and subjective observatory dependent interpretation. The importance of IgG OCB in MS diagnosis relies on its detection in 95 % of MS patients [49],

although they could be found in other chronic inflammatory CNS diseases [58]. Moreover, their presence in CSF of CIS patients is a predictor to MS conversion [59].

Alternatively, IgG intrathecal synthesis can be determined also by the Reiber and Felgenhauer formula [60], Tourtellote formula [61], Schuller formula [62] and IgG or Tibbling Link index [63]. The most widely used formula is the IgG index. This evaluates the amount of IgG in the CSF compared to the levels in serum and is calculated as the ratio of IgG to albumin in CSF compared to the ratio of IgG to albumin in serum [64]. Albumin is included in the index because the albumin quotient (Qalb) CSF/serum, is used as a measurement of BBB dysfunction in MS [65]. An IgG index higher than 0.7 indicates increased intrathecal B cell response [66], and thus probably reveals a MS diagnostic, based on the fact that nearly 70 % of MS patients have increased IgG index [67,68]. However, there are other CNS diseases in which IgG index could increase [58].

4.2. IgM intrathecal synthesis

As IgG, IgM OCBs can be detected resulting from intrathecal production of IgM [69]. Its presence has been linked to increased risk of conversion from CIS to MS and to an aggressive evolution of the disease [64,70,71]. IgM index calculated using the formula $CSF\ IgM \times serum\ albumin / serum\ IgM \times CSF\ albumin$ can be used to establish the intrathecal IgM synthesis. IgM index higher than 0.1 is considered increased and it predicts bad prognosis of the disease [72–74].

4.3. Kappa and Lambda free light chains

Kappa and Lambda immunoglobulins free light chain (κ FLC, λ FLC), are produced by B-lymphocytes during antibody synthesis. An increase in serum in both FLC production has been reported in inflammatory and autoimmune systemic diseases [75]. Whereas in CSF, intrathecal immunoglobulin synthesis is commonly observed in inflammatory disorders of the CNS (also when the origin is infectious) [76]. Different automated nephelometric assays are available for the detection of FLC [77]. The two most studied ones are Siemens® N-latex κ FLC and λ FLC [78] and Binding Site Freelite® FLC [79]. These two assays are

Table 1

McDonald Criteria 2017 summary for the diagnosis of MS.

McDonald Criteria 2017 for the diagnosis of MS	
CLINICAL PRESENTATION	ADDITIONAL CRITERIA TO MAKE MS DIAGNOSIS
...in a person who has experienced a typical attack/CIS at onset	
<ul style="list-style-type: none"> • 2 or more attacks and clinical evidence of 2 or more lesions OR 2 or more attacks and clinical evidence of 1 lesion with clear historical evidence of prior attack involving lesion in different location • 2 or more attacks and clinical evidence of 1 lesion 	None
<ul style="list-style-type: none"> • 1 attack and clinical evidence of 2 or more lesions 	<p>DIS Additional clinical attack implicating different CNS site 1 or more MS typical T2 lesions in 2 or more areas of CNS</p> <p>DIT Additional clinical attack Simultaneous presence of both enhancing and non-enhancing MS typical MRI lesions, or new T2 or enhancing MRI lesion compared to baseline scan CSF oligoclonal bands</p>
<ul style="list-style-type: none"> • 1 attack and clinical evidence of 1 lesion 	<p>DIS Additional attack implicating different CNS site 1 or more MS-typical T2 lesions in 2 or more areas of CNS.</p> <p>AND DIT Additional clinical attack Simultaneous presence of both enhancing and non-enhancing MStypical MRI lesions, or new T2 or enhancing MRI lesion compared to baseline scan (without regard to timing of baseline scan) CSF oligoclonal bands</p>
...in a person who has steady progression of disease since onset	
<ul style="list-style-type: none"> • 1 year of disease progression (retrospective or prospective) 	<p>DIS 1 or more MS-typical T2 lesions 2 or more T2 spinal cord lesions CSF oligoclonal bands</p>

comparable but not interchangeable for patient's follow-up. Thus, the same assay should be employed throughout the whole study period [80–82]. In comparison to OCB detection, FLC assays present important advantages, such as the simplicity of automated methods and the fact that the results are objective and quantifiable [83]. Furthermore, FLC methods are not influenced neither by hemolysis nor by long storage in CSF samples, indicating high stability [84]. Although κ FLC and λ FLC can be elevated in the CSF of MS patients, κ FLC showed better correlation in MS diagnosis [85,86] and its index has higher sensitivity and specificity [87–91]. κ -index is the ratio between CSF and serum κ FLC levels, taking into account the altered permeability of the BBB, through the Qalb, as described by Duranti et al. [92]. In addition, other κ FLC-derived formulas are the ratio between κ FLC and IgG in CSF, and the CSF to serum κ FLC ratio (Q κ FLC). All these κ FLC parameters have been reported to have a diagnostic accuracy for MS diagnosis similar or superior to OCB [88–91,93,94]. Moreover, both κ FLC formula and IgG OCB detection have been proposed as dual assays to improve the diagnostic accuracy for MS disease [93,95]. The prognostic role of κ FLC absolute concentrations in CSF has been established in the conversion of CIS to MS [96–98]. In the same way, the prognostic role of the κ -index [99] and the CSF κ FLC/IgG ratio [100] have been proven. Rosenstein et al. [101], have recently reported that high κ -index at baseline is predictive of progression independent of relapse activity. However, and in spite of the high sensitivity and specificity of κ -index, there is no consensus about its diagnostic cut-off values, being variable between studies [82,87,102–104].

5. Potential biomarkers

5.1. Neurofilaments

When axonal damage occurs, CNS neurofilaments (Nfs) are released [59]. Nfs are very stable cylindrical proteins composed of heavy (NfH), medium (NfM) and light (NfL) chains and α -internexin [105]. They are located in the neuronal cytoplasm, conferring stability to neurons and being extensively expressed in axons, especially NfL [106,107]. After damage, Nfs are released, reaching the interstitial fluid, and consequently, the CFS and the blood. CSF NfL increase under normal conditions with age, and it is associated with cognitive decline and motor impairment [108]. Both NfH and NfL chains have been studied in CSF as biomarkers in MS, being the NfL assay more sensitive [109,110]. Although CSF Nfs are increased not only in MS, but also in other neuronal pathologies, such as Alzheimer's disease, Creutzfeldt-Jakob disease, frontotemporal dementia, human immunodeficiency virus (HIV) associated dementia, amyotrophic lateral sclerosis, atypical parkinsonian disorders and traumatic brain injury [106].

Enzyme-linked immunosorbent assays (ELISA) were the first proposed for NfL quantification in CFS, nevertheless they could not be used for serum measurements due to its low sensitivity. NfL levels in serum are approximately 40 fold lower than in CSF. But it has been demonstrated that serum NfL could be measured in CIS and MS patients by using an electrochemiluminescence (ECL) based assay. Nevertheless its sensitivity was not the optimal [111–114]. A new single molecule array (Simoa) assay has shown 25 fold higher sensitivity than ECL assays, allowing the use of serum for the study of MS and avoiding the invasive procedure of lumbar puncture [115].

Higher NfL levels are detected in RRMS and progressive MS patients compared to healthy control subjects [110,116,117]. Patients with RRMS have shown higher NfL levels when clinical exacerbation occurs or when having contrast-enhancing lesions, while in progressive MS forms there is no correlation between NfL levels and disease activity [118,119]. As well as for CSF NfL, the main utility of serum NfL is the prognosis value. The association between NfL in serum and EDSS has been studied, finding a direct relation between serum NfL and MS severity score [118,120–122]. In addition, it has been shown that NfL decrease after natalizumab [55,110,123], rituximab [124] or fingolimod [125,126] treatment, showing promising utility for treatment follow-up in MS patients. However, there is no consensus in serum NfL thresholds, because of the lack of harmonization between different assays and the within-individual fluctuating levels during relapsing crisis happen [120,122,127–129].

5.2. Tau protein

Tau protein belongs to the microtubule-associated proteins family as a heat stable protein essential for microtubule assembly [130]. It is released upon neuronal damage and can be measured in CSF [131,132]. Its use in MS is controversial as some studies correlate higher CSF tau protein concentrations with disease progression [133] or with the time to next relapse [134]. Meanwhile others do not find differences in tau protein concentrations in MS patients compared to the control group nor a correlation between tau protein and EDSS scores [135,136].

5.3. Glial fibrillary acidic protein

Glial fibrillary acidic protein (GFAP) is the major intermediate cytoskeletal protein in the astrocytes [137,138]. GFAP is released rapidly subsequent to axonal degeneration and it has been widely studied as CSF biomarker in traumatic brain injury [139,140]. Many studies have reported higher levels of GFAP in CFS and in blood in MS patients compared to healthy controls [141–150]. It has also been established that patients with relapsing MS have higher GFAP levels in CSF compared to those with MS in remission [143,145,146,150,151]. A

Table 2
Classification of MS biomarkers.

Biomarker	Biology	Sample	Pathology	Utility	Result in MS
IgG OCB	Intrathecal IgG synthesis	CSF	Inflammation	Diagnosis	↑
IgM index	Intrathecal IgM synthesis	CSF	Inflammation	Prognosis	↑
KFLC	Secreted by B-lymphocytes	CSF	Inflammation	Diagnosis/Prognosis	↑
Neurofilament	Axonal protein	CSF/ blood	Neuronal damage	Diagnosis/Prognosis	↑
Tau protein	Microtubule structural protein	CSF	Neuronal damage	Controversial	Controversial
GFAP	Cytoskeletal protein in the astrocytes	CSF/ blood	Neuronal damage	Diagnosis/Prognosis	↑
S100β	Ca ²⁺ and Zn ²⁺ binding protein secreted by astrocytes	CSF	Neuronal damage	Diagnosis/Prognosis/ Treatment efficacy	↑
MBP	Part of the myelin sheath, synthesized by oligodendroglia	CSF	Demyelination	Diagnosis	↑
CHI3L1	Extracellular monomeric single-chain glycoprotein expressed in astrocytes macrophages, chondrocytes, synovial cells, osteoblasts, and neutrophils	CSF	Neuronal damage/ Inflammation	Diagnosis/Treatment efficacy	↑
Osteopontin	Extracellular matrix protein secreted by activated macrophages, leukocytes and activated T lymphocytes	CSF/ blood	Inflammation	Prognosis	↑
CXCL12	Cytokine	CSF	Inflammation	Diagnosis	↑
CXCL13	Cytokine	CSF/ blood	Inflammation	Prognosis	↑
CD163	Monocyte/macrophage specific membrane marker	CSF/ blood	Inflammation	Prognosis	↑
CD5 + B cells	B cells	Blood	Inflammation	Prognosis	↑ RRMS ↓ SPMS
Tubulin β	Microtubules component	CSF	Neuronal damage	Potential in prognosis	↑
HSP70	Chaperones	CSF	Inflammation	MS inflammation biomarker	↑

Cerebrospinal fluid, CSF; Chemokine ligand, CXCL; Chitinase-3-like protein, CHI3L1; Glial fibrillary acidic protein, GFAP; Heat shock protein 70, HSP70; Immunoglobulin, Ig; Kappa free light chain, KFLC; Oligoclonal bands, OCB; Myelin basic protein, MBP.

correlation between CSF NfL [120,122] and GFAP [147,149] with EDSS has been described suggesting their role in disable progression prediction. However, Jiang et al. [152], have recently reported that GFAP at baseline correlate with lesion volume but not with disease progression measured through EDSS, Timed 25-Foot Walk, 9-Hole Peg Test, and composite confirmed disability progression tests. Serum GFAP has been reported to be higher in MS patients than in healthy controls and also higher in PPMS than in RRMS patients [142,143,145]. In addition, patients with RRMS showed no significant differences in serum GFAP compared to healthy controls. Serum GFAP could also be used to differentiate MS subtypes, being an adequate marker to assess disease progression [143].

5.4. S100 β

S100β is a small Ca²⁺ and Zn²⁺ binding protein, mostly secreted by astrocytes, but also by oligodendrocytes and certain neuronal subpopulations [153,154]. S100β promotes neuronal proliferation, oligodendrocyte differentiation, and astrocyte morphology maintenance [155]. It has been also reported to control the activation of GFAP, the polymerization of tubulin and DNA repair [156]. S100β can either act as a neurotrophic or as a neurotoxic molecule in vitro, depending on the concentration attained [157–160]. S100β is increased in CSF [161] and serum [162] of patients with MS on acute phase, but also during the course of the disease. It could also be increased in acute brain damage including strokes [163], rapid parenchymal destruction [164] or traumatic brain injuries [165]. Petzold et al. [150] demonstrated a significant increasing trend in S100β levels from PPMS to SPMS and then to RRMS. Moreover, S100β was higher in CSF and serum of MS patients at the time of diagnosis of RRMS compared to healthy control patients [162]. However, additional studies should be performed to evaluate whether S100β concentrations are associated with different MS stages and therefore it could potentially be used as a prognostic tool. Moreover, since S100β was recently reported to decrease upon MS treatment with immunosuppressive [166] or natalizumab [167], it can be also considered as a biomarker of treatment efficacy.

5.5. Myelin basic protein

Myelin basic protein (MBP) is part of the myelin sheath together with myelin oligodendrocyte glycoprotein (MOG) and proteolipid protein [168]. MBP is synthesized by oligodendroglia cells in various isoforms that later undergo a relatively large number of posttranslational modifications, such as deamination, citrullination and phosphorylation [169]. Also MBP is generally considered to maintain the compaction of the myelin sheath [170]. Detectable concentrations of MBP in CSF have been found in acute demyelination [171], but not only in MS patients [172,173]. MBP also increases in acute disseminated encephalomyelitis [174,175], encephalitis [176], acute cerebral infarction [177] and neuro-Behcet's disease [178]. It should also be noted that higher CSF MBP concentrations are not related to MS severity or prognosis [179]. In addition, CNS inflammation correlates with increased MBP deamination leading to higher levels of citrullinated MBP [180,181] which could be used as a prognosis biomarker. Its correlation with MS severity has been proven to be strong, although once again, it increases in other neurology pathologies too [182,183].

Anti-myelin antibodies (anti-MOG and anti-MBP) have also been widely studied. It is considered that the MBP increase after a demyelinating event is followed by an immunoactivity phase [184]. The capacity of anti-myelin antibodies to estimate individual risk for progression of MS disease remains unclear. Concurrently, there are studies that suggest an association between serum anti-myelin antibody status and MS prognosis [185]. However, Kuhle et al. [186] did not find significant differences in MS diagnosis or risk progression between patients showing or not anti-MOG antibodies, anti-MBP antibodies, or both.

5.6. Chitinase-3-like 1

Chitinase-3-like 1 (CHI3L1), also known as YKL-40, is an extracellular monomeric single-chain glycoprotein expressed in astrocytes macrophages, chondrocytes, synovial cells, osteoblasts, and neutrophils [127,187,188] all of them involved in tissue remodeling and inflammation [189]. CHI3L1 increases in CSF in neuronal damage, such as in MS [190,191], but also in other CNS neuroinflammatory pathologies

[192]. It has been shown CIS patients exhibit lower CSF CHI3L1 than patients diagnosed with MS [193]. In addition, CSF CHI3L1 directly correlates with the risk of CIS conversion to MS [194] and with a rapid conversion at high concentrations. A meta-analysis recently conducted by Floro et al. [195], reported higher CHI3L1 in the remission stages of MS than during relapse. These findings suggest that CHI3L1 is possibly a reliable biomarker for clinical practice to monitor the course of MS even in the later stages. Schneider et al. [196] recently demonstrated that CHI3L1 correlates with spinal cord atrophy in progressive MS and also with EDSS [197]. Furthermore, CHI3L1 has utility in the evaluation of MS patient's response to IFN-gamma treatment, showing higher concentrations in the non-responder group [198]. However, serum CHI3L1 needs to be further studied.

5.7. Osteopontin

Osteopontin (OPN) is an extracellular matrix protein secreted by activated macrophages, leukocytes and activated T lymphocytes involved in inflammation and autoimmune disorders, including MS [199]. Higher levels of OPN have been reported in CSF and blood of MS patients [200]. Likewise, increased OPN concentrations in CSF and plasma have been associated with MS progression. Orsi et al. [201] described that higher CSF OPN is related with increased 10-year lesion size, suggesting the importance of inflammation in long-term disease progression. In addition, Marastoni et al. [202] performed a study where CSF OPN showed a relationship with disease progression in RRMS patients after being treated with dimethylfumarate.

5.8. Chemokine ligand

Chemokine (C-X-C motif) ligand 13 or B lymphocyte chemo-attractant (CXCL13) is involved in MS pathogenesis [203]. Both serum and CSF CXCL13 expression are increased in MS patients and CIS [204]. Recently, DiSano et al. [205] have evaluated the use of a CXCL13 index, calculated as IgG index or KFLC index, to establish whether or not the production is intrathecal. CXCL13 index has proven to be a reliable biomarker for MS prognosis and follow-up when used alone or in combination with NfL. Moreover, CXCL13 has higher sensitivity than NfL in the prediction of future neuroinflammatory activity in MS patients. This is because it only increases in CSF when neuroinflammation occurs [206,207].

On the other hand, stromal cell-derived factor or C-X-C motif chemokine 12 (CXCL12) is a potent chemo-attractant molecule for different immune cells, including monocytes, T cells, B cells, and plasma cells [208]. Also it is primordial in neuronal development [209,210]. Whereas CSF CXCL12 is elevated in both active and inactive MS, CXCL13 CSF levels were increased in MS active disease only, suggesting it is more specific to evaluate disease progression, as described by Krumbholz et al. [211].

5.9. CD163

CD163 is a monocyte/macrophage specific membrane marker cleaved from the surface of activated macrophages as a soluble form (sCD163) when inflammation of the CNS occurs. sCD163 can be detected in blood and CSF [59]. Fabriek et al. [212] found an up-regulation of plasma sCD163 and a down-regulation of membrane CD163 in MS patients compared to healthy controls. This conclusion suggests a higher release of this biomarker in MS patients. Furthermore, the sCD163 CSF/serum ratio was significantly increased in MS patients reflecting macrophage activation in MS inflammatory lesions [213].

5.10. CD5 + B cells

B cells have been increasingly recognized as major players in the pathogenesis of MS, as they are involved in other autoimmune diseases

[214,215]. CD5 + B cells can be determined in blood samples by flow cytometry and its increased presence has been associated with the development of RRMS [216,217]. Also, it has been demonstrated that the percentage of CD5 + B cells is higher in patients with active disease [218]. Whereas CD5 + B cells have been reported to be decreased in SPMS, some authors suggest that patients with decreased CD5 + B cells and RRMS tend to develop SPMS [214]. In addition, this biomarker has been associated with further elevated risk of early CIS conversion to MS and a higher relapse rate in these patients, independent of IgG OCB presence and MRI findings [219].

5.11. Tubulin β

Tubulin is the major component of microtubules. An alpha (α) and a beta (β) subunits conform the heterodimeric tubulin which exists in six isotypes forms [220]. Tubulin β II is present in the brain and is increased in neuronal development and regeneration. Meanwhile, tubulin β III is also present in dorsal root ganglia and enhanced during axonal growing in the fetal and postnatal period [221,222]. In a preliminary study, Madeddu et al. [151] have suggested that tubulin β II could be a potential candidate for diagnosis. Also, that tubulin β III could be a possible prognostic biomarker of MS. Further studies should be performed to better establish the usefulness of these new biomarkers.

5.12. Heat shock protein 70

Heat shock proteins (HSP) are a group of chaperones that have homeostatic functions in CNS [223]. HSP70 is located in the cytosol protecting cells against lethal stress-induced damage, or in the cell membrane and the intracellular space playing an important role in the immune response [224,225]. Lechner et al. [226], conducted a study to determine whether HSP70 contributes to the neurodegenerative or inflammatory processes in MS. Their results conclude that HSP70 concentrations were significantly higher in patients with CIS or RRMS than in patients with PPMS or SPMS. This may be correlated with the inflammatory process in the first subgroup, which means HSP70 would be a useful biomarker to monitor inflammation in MS.

6. Conclusions

MS is a disabling disease, which needs to be diagnosed as early as possible, requiring a continuous search for new biomarkers and an attempt to achieve a solid consensus on their use. Thus, the latest McDonald's criteria for MS diagnosis included the presence of IgG OCB in CSF as a substitute for dissemination in time, enabling an early MS diagnosis in patients meeting the criteria of dissemination in space, crucial to implement an adequate treatment optimizing clinical outcomes over time. Furthermore, encouraging studies have been undertaken on new potential biomarkers in CSF and blood serum that could be useful in the diagnosis and prognosis of different forms and stages of MS, even if they are not specific for MS but used for other CNS diseases. Despite this, more extensive research is needed to determine the clinical usefulness of these biomarkers and to assess its applicability in everyday practice.

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Author contributions

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

No data was used for the research described in the article.

References

- [1] C. Lucchinetti, W. Brück, J. Parisi, B. Scheithauer, M. Rodriguez, H. Lassmann, Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination, *Ann. Neurol.* 47 (2000) 707–717, [https://doi.org/10.1002/1531-8249\(200006\)47:6<707::aid-ana3>3.0.co;2-q](https://doi.org/10.1002/1531-8249(200006)47:6<707::aid-ana3>3.0.co;2-q).
- [2] B.G. Weinschenker, Epidemiology of multiple sclerosis, *Neurol. Clin.* 14 (1996) 291–308, [https://doi.org/10.1016/S0733-8619\(05\)70257-7](https://doi.org/10.1016/S0733-8619(05)70257-7).
- [3] M. Pugliatti, G. Rosati, H. Carton, T. Riise, J. Drulovic, L. Vécsei, I. Milanov, The epidemiology of multiple sclerosis in Europe, *Eur. J. Neurol.* 13 (2006) 700–722, <https://doi.org/10.1111/j.1468-1331.2006.01342.x>.
- [4] N. Perez-Carmona, E. Fernandez-Jover, A.P. Sempere, Epidemiology of multiple sclerosis in Spain, *Rev. Neurol.* 69 (2019) 32–38, <https://doi.org/10.33588/rn.6901.2018477>.
- [5] S.-M. Orton, B.M. Herrera, I.M. Yee, W. Valdar, S.V. Ramagopalan, A. D. Sadovnick, G.C. Ebers, Canadian Collaborative Study Group, Sex ratio of multiple sclerosis in Canada: a longitudinal study, *Lancet Neurol.* 5 (2006) 932–936, [https://doi.org/10.1016/S1474-4422\(06\)70581-6](https://doi.org/10.1016/S1474-4422(06)70581-6).
- [6] I. Kister, E. Chamot, A.R. Salter, G.R. Cutter, T.E. Bacon, J. Herbert, Disability in multiple sclerosis: a reference for patients and clinicians, *Neurology.* 80 (2013) 1018–1024, <https://doi.org/10.1212/WNL.0b013e3182872855>.
- [7] J. Oh, A. Vidal-Jordana, X. Montalban, Multiple sclerosis: clinical aspects, *Curr. Opin. Neurol.* 31 (2018) 752–759, <https://doi.org/10.1097/WCO.0000000000000622>.
- [8] D. Miller, F. Barkhof, X. Montalban, A. Thompson, M. Filippi, Clinically isolated syndromes suggestive of multiple sclerosis, part I: natural history, pathogenesis, diagnosis, and prognosis, *Lancet Neurol.* 4 (2005) 281–288, [https://doi.org/10.1016/S1474-4422\(05\)70071-5](https://doi.org/10.1016/S1474-4422(05)70071-5).
- [9] D.H. Miller, B.G. Weinschenker, M. Filippi, B.L. Banwell, J.A. Cohen, M. S. Freedman, S.L. Galetta, M. Hutchinson, R.T. Johnson, L. Kappos, J. Kira, F. D. Lublin, H.F. McFarland, X. Montalban, H. Panitch, J.R. Richert, S.C. Reingold, C.H. Polman, Differential diagnosis of suspected multiple sclerosis: a consensus approach, *Mult. Scler. Houndmills Basingstoke Engl.* 14 (2008) 1157–1174, <https://doi.org/10.1177/1352458508096878>.
- [10] F.D. Lublin, S.C. Reingold, J.A. Cohen, G.R. Cutter, P.S. Sorensen, A.J. Thompson, J.S. Wolinsky, L.J. Balcer, B. Banwell, F. Barkhof, B. Bebo, P.A. Calabresi, M. Clanet, G. Comi, R.J. Fox, M.S. Freedman, A.D. Goodman, M. Ingles, L. Kappos, B.C. Kieseier, J.A. Lincoln, C. Lubetzki, A.E. Miller, X. Montalban, P. W. O'Connor, J. Petkau, C. Pozzilli, R.A. Rudick, M.P. Sormani, O. Stüve, E. Waubant, C.H. Polman, Defining the clinical course of multiple sclerosis: the 2013 revisions, *Neurology.* 83 (2014) 278–286, <https://doi.org/10.1212/WNL.0000000000000560>.
- [11] R. Dobson, G. Giovannoni, Multiple sclerosis - a review, *Eur. J. Neurol.* 26 (2019) 27–40, <https://doi.org/10.1111/ene.13819>.
- [12] C.H. Polman, S.C. Reingold, B. Banwell, M. Clanet, J.A. Cohen, M. Filippi, K. Fujihara, E. Havrdova, M. Hutchinson, L. Kappos, F.D. Lublin, X. Montalban, P. O'Connor, M. Sandberg-Wollheim, A.J. Thompson, E. Waubant, B. Weinschenker, J.S. Wolinsky, Diagnostic criteria for multiple sclerosis: 2010 Revisions to the McDonald criteria, *Ann. Neurol.* 69 (2011) 292–302, <https://doi.org/10.1002/ana.22366>.
- [13] C.F. Lucchinetti, R.H. Gavrilova, I. Metz, J.E. Parisi, B.W. Scheithauer, S. Weigand, K. Thomsen, J. Mandrekar, A. Altintas, B.J. Erickson, F. König, C. Giannini, H. Lassmann, L. Linbo, S.J. Pittock, W. Brück, Clinical and radiographic spectrum of pathologically confirmed tumefactive multiple sclerosis, *Brain J. Neurol.* 131 (2008) 1759–1775, <https://doi.org/10.1093/brain/awn098>.
- [14] S.L. Hauser, B.A.C. Cree, Treatment of multiple sclerosis: A review, *Am. J. Med.* 133 (2020) 1380–1390.e2, <https://doi.org/10.1016/j.amjmed.2020.05.049>.
- [15] A. Ascherio, K.L. Munger, Environmental risk factors for multiple sclerosis. Part I: the role of infection, *Ann. Neurol.* 61 (2007) 288–299, <https://doi.org/10.1002/ana.21117>.
- [16] L. Alfredsson, T. Olsson, Lifestyle and environmental factors in multiple sclerosis, *Cold Spring Harb. Perspect. Med.* 9 (4) (2019) a028944, <https://doi.org/10.1101/cshperspect.a028944>.
- [17] L.F. Barcellos, J.R. Oksenberg, A.B. Begovitch, E.R. Martin, S. Schmidt, E. Vittinghoff, D.S. Goodin, D. Pelletier, R.R. Lincoln, P. Bucher, A. Swerdlin, M. A. Pericak-Vance, J.L. Haines, S.L. Hauser, Multiple Sclerosis Genetics Group, HLA-DR2 dose effect on susceptibility to multiple sclerosis and influence on disease course, *Am. J. Hum. Genet.* 72 (2003) 710–716, <https://doi.org/10.1086/367781>.
- [18] International Multiple Sclerosis Genetics Consortium, Wellcome Trust Case Control Consortium 2, S. Sawcer, G. Hellenthal, M. Pirinen, C.C.A. Spencer, N.A. Patsopoulos, L. Moutsianas, A. Dilthey, Z. Su, C. Freeman, S.E. Hunt, S. Edkins, E. Gray, D.R. Booth, S.C. Potter, A. Goris, G. Band, A.B. Oturai, A. Strange, J. Saarela, C. Bellenguez, B. Fontaine, M. Gillman, B. Hemmer, R. Gwilliam, F. Zipp, A. Jayakumar, R. Martin, S. Leslie, S. Hawkins, E. Giannoulatou, S. D'alfonso, H. Blackburn, F. Martinelli Boneschi, J. Liddle, H.F. Harbo, M.L. Perez, A. Spurkland, M.J. Waller, M.P. Mycko, M. Ricketts, M. Comabella, N. Hammond, I. Kockum, O. T. McCann, M. Ban, P. Whittaker, A. Kempainen, P. Weston, C. Hawkins, S. Widaa, J. Zajicek, S. Dronov, N. Robertson, S.J. Bumpstead, L.F. Barcellos, R. Ravindrarajah, R. Abraham, L. Alfredsson, K. Ardlie, C. Aubin, A. Baker, K. Baker, S.E. Baranzini, L. Bergamaschi, R. Bergamaschi, A. Bernstein, A. Berthele, M. Boggild, J.P. Bradfield, D. Brassat, S.A. Broadley, D. Buck, H. Butzkueven, R. Capra, W.M. Carroll, P. Cavalla, E.G. Celius, S. Cepok, R. Chiavacci, F. Clerget-Darpoux, K. Clysters, G. Comi, M. Cossburn, I. Cournu-Rebeix, M.B. Cox, W. Cozen, B.A.C. Cree, A.H. Cross, D. Cusi, M.J. Daly, E. Davis, P.I.W. de Bakker, M. Debooverie, M.B. D'hooghe, K. Dixon, R. Dobosi, B. Dubois, D. Ellinghaus, I. Elovaaara, F. Esposito, C. Fontenille, S. Foote, A. Franke, D. Galimberti, A. Ghezzi, J. Glessner, R. Gomez, O. Gout, C. Graham, S.F.A. Grant, F.R. Guerini, H. Hakonarson, P. Hall, A. Hamsten, H.-P. Hartung, R.N. Heard, S. Heath, J. Hobart, M. Hoshi, C. Infante-Duarte, G. Ingram, W. Ingram, T. Islam, M. Jagodic, M. Kabesch, A.G. Kermod, T.J. Kilpatrick, C. Kim, N. Klopp, K. Koivisto, M. Larsson, M. Lathrop, J.S. Lechner-Scott, M.A. Leone, V. Leppä, U. Liljedahl, I.L. Bomfim, R. R. Lincoln, J. Link, J. Liu, A.R. Lorentzen, S. Lupoli, F. Pobywajlo, H.L. Quach, P.P. Ramsay, M. Reunanen, R. Reynolds, J.D. Rioux, M. Rodegher, S. Roessler, J.P. Rubio, I.-M. Rückert, M. Salvetti, E. Salvi, A. Santaniello, C.A. Schaefer, S. Schreiber, C. Schulze, R.J. Scott, F. Sellebjerg, K.W. Selmaj, D. Sexton, L. Shen, B. Simms-Acuna, S. Skidmore, P.M.A. Sleiman, C. Smestad, P.S. Sorensen, H.B. Søndergaard, J. Stankovich, R.C. Strange, A.-M. Sulonen, E. Sundqvist, A.-C. Syvänen, F. Taddeo, B. Taylor, J.M. Blackwell, P. Tienari, E. Bramer, A. Tourbah, M.A. Brown, E. Tronczynska, J.P. Casas, N. Tubridy, A. Corvin, J. Vickery, J. Jankowski, P. Villoslada, H.S. Markus, K. Wang, C.G. Mathew, J. Watson, C.N.A. Palmer, H.-E. Wichmann, R. Plomin, E. Willoughby, A. Rautanen, J. Winkelmann, M. Wittig, R.C. Trembath, J. Yaouanq, A.C. Viswanathan, H. Zhang, N.W. Wood, R. Zuvich, P. Deloukas, C. Langford, A. Duncanson, J.R. Oksenberg, M.A. Pericak-Vance, J.L. Haines, T. Olsson, J. Hillert, A.J. Ivinson, P.L. De Jager, L. Peltonen, G. J. Stewart, D.A. Hafler, S.L. Hauser, G. McVean, P. Donnelly, A. Compston, Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis, *Nature.* 476 (2011) 214–219, <https://doi.org/10.1038/nature10251>.
- [19] D.A. Hafler, A. Compston, S. Sawcer, E.S. Lander, M.J. Daly, P.L. De Jager, P.I. W. de Bakker, S.B. Gabriel, D.B. Mirel, A.J. Ivinson, M.A. Pericak-Vance, S. G. Gregory, J.D. Rioux, J.L. McCauley, J.L. Haines, L.F. Barcellos, B. Cree, J. R. Oksenberg, S.L. Hauser, International Multiple Sclerosis Genetics Consortium, Risk alleles for multiple sclerosis identified by a genomewide study, *N. Engl. J. Med.* 357 (2007) 851–862, <https://doi.org/10.1056/NEJMoa073493>.
- [20] J. Link, A.R. Lorentzen, I. Kockum, K. Duvefelt, B.A. Lie, E.G. Celius, H.F. Harbo, J. Hillert, B. Brynedal, Two HLA class I genes independently associated with multiple sclerosis, *J. Neuroimmunol.* 226 (2010) 172–176, <https://doi.org/10.1016/j.jneuroim.2010.07.006>.
- [21] J.C. Lee, J.T. Laydon, P.C. McDonnell, T.F. Gallagher, S. Kumar, D. Green, D. McNulty, M.J. Blumenthal, J.R. Heys, S.W. Landvatter, J.E. Strickler, M. M. McLaughlin, I.R. Siemens, S.M. Fisher, G.P. Livi, J.R. White, J.L. Adams, P. R. Young, A protein kinase involved in the regulation of inflammatory cytokine biosynthesis, *Nature.* 372 (1994) 739–746, <https://doi.org/10.1038/372739a0>.
- [22] J.A. Nick, S.K. Young, P.G. Arndt, J.G. Lieber, B.T. Surat, K.R. Poch, N.J. Avdi, K. C. Malcolm, C. Taube, P.M. Henson, G.S. Worthen, Selective suppression of neutrophil accumulation in ongoing pulmonary inflammation by systemic inhibition of p38 mitogen-activated protein kinase, *J. Immunol. Baltim. Md* 169 (9) (2002) 5260–5269, <https://doi.org/10.4049/jimmunol.169.9.5260>.
- [23] J. Van Steenwinkel, A.-L. Schang, M.L. Krishnan, V. Degos, A. Delahaye-Duriez, C. Bokobza, Z. Csaba, F. Verdonk, A. Montané, S. Sigaut, O. Hennebert, S. Lebon, L. Schwendimann, T. Le Charpentier, R. Hassan-Abdi, G. Ball, P. Aljabar, A. Saxena, R.K. Holloway, W. Birchmeier, O. Baud, D. Rowitch, V. Miron, F. Chretien, C. Leconte, V.C. Besson, E.G. Petretto, A.D. Edwards, H. Hagberg, N. Soussi-Yanicostas, B. Fleiss, P. Gressens, Decreased microglial Wnt/ β -catenin signalling drives microglial pro-inflammatory activation in the developing brain, *Brain.* 142 (2019) 3806–3833, <https://doi.org/10.1093/brain/awz319>.
- [24] T.L. Biechele, R.M. Kulikauskas, R.A. Toroni, O.M. Lucero, R.D. Swift, R.G. James, N.C. Robin, D.W. Dawson, R.T. Moon, A.J. Chien, Wnt/ β -catenin signaling and AXIN1 regulate apoptosis triggered by inhibition of the mutant kinase BRAFV600E in human melanoma, *Sci. Signal.* 5 (2012) ra3, <https://doi.org/10.1126/scisignal.2002274>.
- [25] J.W. Peterson, L. Bø, S. Mörk, A. Chang, R.M. Ransohoff, B.D. Trapp, VCAM-1-positive microglia target oligodendrocytes at the border of multiple sclerosis lesions, *J. Neuropathol. Exp. Neurol.* 61 (2002) 539–546, <https://doi.org/10.1093/jnen/61.6.539>.
- [26] G.J.A. ten Bosch, J. Bolk, B.A. 't Hart, J.D. Laman, Multiple sclerosis is linked to MAPK overactivity in microglia, *J. Mol. Med. Berl. Ger.* 99 (8) (2021) 1033–1042, <https://doi.org/10.1007/s00109-021-02080-4>.
- [27] B. Arneht, Multiple sclerosis and smoking, *Am. J. Med.* 133 (2020) 783–788, <https://doi.org/10.1016/j.amjmed.2020.03.008>.
- [28] M. Adamczyk-Sowa, B. Gębka-Kepińska, M. Kepiński, Multiple sclerosis - risk factors, *Wiadomości Lek. Wars. Pol.* 1960 (73) (2020) 2677–2682.
- [29] Y. Zhang, D.Y.M. Leung, B.N. Richers, Y. Liu, L.K. Remigio, D.W. Riches, E. Goleva, Vitamin D inhibits monocyte/macrophage proinflammatory cytokine

- production by targeting MAPK phosphatase-1, *J. Immunol. Baltim. Md* 188 (5) (2012) 2127–2135, <https://doi.org/10.4049/jimmunol.1102412>.
- [30] J. Smolders, Ø. Torkildsen, W. Camu, T. Holmøy, An update on vitamin D and disease activity in multiple sclerosis, *CNS Drugs* 33 (2019) 1187–1199, <https://doi.org/10.1007/s40263-019-00674-8>.
- [31] M.L. Roberts, N.R. Cooper, Activation of a ras-MAPK-dependent pathway by Epstein-Barr virus latent membrane protein 1 is essential for cellular transformation, *Virology* 240 (1998) 93–99, <https://doi.org/10.1006/viro.1997.8901>.
- [32] K.-M. Lin, S.-J. Lin, J.-H. Lin, P.-Y. Lin, P.-L. Teng, H.-E. Wu, T.-H. Yeh, Y.-P. Wang, M.-R. Chen, C.-H. Tsai, Dysregulation of dual-specificity phosphatases by Epstein-Barr virus LMP1 and its impact on lymphoblastoid cell line survival, *J. Virol.* 94 (2020) e01837–e10919, <https://doi.org/10.1128/JVI.01837-19>.
- [33] E.M. Mohammed, Environmental influencers, MicroRNA, and multiple sclerosis, *J. Cent. Nerv. Syst. Dis.* 12 (2020), 1179573519894955, <https://doi.org/10.1177/1179573519894955>.
- [34] L. Belbasis, V. Bellou, E. Evangelou, J.P.A. Ioannidis, I. Tzoulaki, Environmental risk factors and multiple sclerosis: an umbrella review of systematic reviews and meta-analyses, *Lancet Neurol.* 14 (2015) 263–273, [https://doi.org/10.1016/S1474-4422\(14\)70267-4](https://doi.org/10.1016/S1474-4422(14)70267-4).
- [35] A. Noori-Zadeh, S.A. Mesbah-Namin, S. Bistoon-beigloo, S. Bakhtiyari, H.-A. Abbaszadeh, S. Darabi, M. Rajabibazi, A. Abdanipour, Regulatory T cell number in multiple sclerosis patients: A meta-analysis, *Mult. Scler. Relat. Disord.* 5 (2016) 73–76, <https://doi.org/10.1016/j.msard.2015.11.004>.
- [36] J. Haas, A. Hug, A. Viehöver, B. Fritzsche, C.S. Falk, A. Filser, T. Vetter, L. Milkova, M. Korporal, B. Fritz, B. Storch-Hagenlocher, P.H. Krammer, E. Suri-Payer, B. Wildemann, Reduced suppressive effect of CD4+CD25high regulatory T cells on the T cell immune response against myelin oligodendrocyte glycoprotein in patients with multiple sclerosis, *Eur. J. Immunol.* 35 (2005) 3343–3352, <https://doi.org/10.1002/eji.200526065>.
- [37] G. Beriou, C.M. Costantino, C.W. Ashley, L. Yang, V.K. Kuchroo, C. Baecher-Allan, D.A. Hafler, IL-17-producing human peripheral regulatory T cells retain suppressive function, *Blood* 113 (2009) 4240–4249, <https://doi.org/10.1182/blood-2008-10-183251>.
- [38] M. Ayyoub, F. Deknuydt, I. Raimbaut, C. Dousset, L. Leveque, G. Biele, D. Valmori, Human memory FOXP3+ Tregs secrete IL-17 ex vivo and constitutively express the TH17 lineage-specific transcription factor ROR γ t, *Proc. Natl. Acad. Sci. U. S. A.* 106 (2009) 8635–8640, <https://doi.org/10.1073/pnas.0900621106>.
- [39] D.V. Sawant, D.A.A. Vignali, Once a Treg, always a Treg? *Immunol. Rev.* 259 (2014) 173–191, <https://doi.org/10.1111/imr.12173>.
- [40] I. Lazibat, M. Rubinić-Majdak, S. Županić, Multiple sclerosis: New aspects of immunopathogenesis, *Acta Clin. Croat.* 57 (2018) 352–361, <https://doi.org/10.20471/acc.2018.57.02.17>.
- [41] S. Duan, Z. Lv, X. Fan, L. Wang, F. Han, H. Wang, S. Bi, Vitamin D status and the risk of multiple sclerosis: A systematic review and meta-analysis, *Neurosci. Lett.* 570 (2014) 108–113, <https://doi.org/10.1016/j.neulet.2014.04.021>.
- [42] T.F. Runia, W.C.J. Hop, Y.B. de Rijke, D. Buljevac, R.Q. Hintzen, Lower serum vitamin D levels are associated with a higher relapse risk in multiple sclerosis, *Neurology* 79 (2012) 261–266, <https://doi.org/10.1212/WNL.0b013e31825fdec7>.
- [43] A.A. Harandi, S. Shahbeigi, H. Pakdaman, S.-M. Fereshtehnejad, E. Nikravesh, R. Jalilzadeh, Association of serum 25(OH) vitamin D3 concentration with severity of multiple sclerosis, *Iran. J. Neurol.* 11 (2012) 54–58.
- [44] B. Weinstock-Guttman, R. Zivadinov, J. Qu, D. Cookfair, X. Duan, E. Bang, N. Bergsland, S. Hussein, M. Cherneva, L. Willis, M. Heinen-Brown, M. Ramanathan, Vitamin D metabolites are associated with clinical and MRI outcomes in multiple sclerosis patients, *J. Neurol. Neurosurg. Psychiatry* 82 (2011) 189–195, <https://doi.org/10.1136/jnnp.2010.227942>.
- [45] E.M. Mowry, L.B. Krupp, M. Milazzo, D. Chabas, J.B. Strober, A.L. Belman, J. C. McDonald, J.R. Oksenberg, P. Bacchetti, E. Waubant, Vitamin D status is associated with relapse rate in pediatric-onset multiple sclerosis, *Ann. Neurol.* 67 (2010) 618–624, <https://doi.org/10.1002/ana.21972>.
- [46] J. Smolders, P. Menheere, A. Kessels, J. Damoiseaux, R. Hupperts, Association of vitamin D metabolite levels with relapse rate and disability in multiple sclerosis, *Mult. Scler. Houndmills Basingstoke Engl.* 14 (2008) 1220–1224, <https://doi.org/10.1177/1352458508094399>.
- [47] I.A.F. Mei, A.-L. Ponsonby, T. Dwyer, L. Blizzard, B.V. Taylor, T. Kilpatrick, H. Butzkueven, A.J. McMichael, Vitamin D levels in people with multiple sclerosis and community controls in Tasmania, Australia, *J. Neurol.* 254 (5) (2007) 581–590, <https://doi.org/10.1007/s00415-006-0315-8>.
- [48] A. Ascherio, K.L. Munger, R. White, K. Kochert, K.C. Simon, C.H. Polman, M. S. Freedman, H.-P. Hartung, D.H. Miller, X. Montalbán, G. Edan, F. Barkhof, D. Pleimes, E.-W. Radü, R. Sandbrink, L. Kappos, C. Pohl, Vitamin D as an early predictor of multiple sclerosis activity and progression, *JAMA Neurol.* 71 (2014) 306–314, <https://doi.org/10.1001/jamaneurol.2013.5993>.
- [49] J. Kuhle, G. Disanto, R. Dobson, R. Adutori, L. Bianchi, J. Topping, J.P. Bestwick, U.-C. Meier, M. Marta, G.D. Costa, T. Runia, E. Evdoshenko, N. Lazareva, E. Thouvenot, P. Iaffaldano, V. Drenzo, M. Khademi, F. Piehl, M. Comabella, M. Sombekke, J. Killestein, H. Hegen, S. Rauch, S. D'Alfonso, J.C. Alvarez-Cermeño, P. Kleinová, D. Horáková, R. Roesler, F. Lauda, S. Llufrú, T. Avsar, U. Uygungölu, A. Altintas, S. Saip, T. Menge, C. Rajda, R. Bergamaschi, N. Moll, M. Khalil, R. Marignier, I. Dujmovic, H. Larsson, C. Malmstrom, E. Scarfepi, C. Fenoglio, S. Wergeland, A. Laroni, V. Annibaldi, S. Romano, A.D. Martínez, A. Carra, M. Salvetti, A. Uccelli, Ø. Torkildsen, K.M. Myhr, D. Galimberti, K. Rejdak, J. Lycke, J.L. Frederiksen, J. Drulovic, C. Confavreux, D. Brassat, C. Enzinger, S. Fuchs, I. Bosca, J. Pelletier, C. Picard, E. Colombo, D. Franciotta, T. Derfuss, RLP Lindberg, Ö. Yaldizli, L. Vécsei, B.C. Kieseier, H.P. Hartung, P. Villoslada, A. Siva, A. Saiz, H. Tumanli, E. Havrdová, L.M. Villar, M. Leone, N. Barizzone, F. Deisenhammer, C. Teunissen, X. Montalban, M. Tintoré, T. Olsson, M. Trojano, S. Lehmann, G. Castelnovo, S. Lapin, R. Hintzen, L. Kappos, R. Furlan, V. Martinelli, G. Comi, S.V. Ramagopalan, G. Giovannoni, Conversion from clinically isolated syndrome to multiple sclerosis: A large multicentre study, *Mult. Scler. Houndmills Basingstoke Engl.* 21 (8) (2015) 1013–1024, <https://doi.org/10.1177/1352458514568827>.
- [50] E. James, R. Dobson, J. Kuhle, D. Baker, G. Giovannoni, S.V. Ramagopalan, The effect of vitamin D-related interventions on multiple sclerosis relapses: a meta-analysis, *Mult. Scler. Houndmills Basingstoke Engl.* 19 (2013) 1571–1579, <https://doi.org/10.1177/1352458513489756>.
- [51] B. Pozuelo-Moyano, J. Benito-León, A.J. Mitchell, J. Hernández-Gallego, A systematic review of randomized, double-blind, placebo-controlled trials examining the clinical efficacy of vitamin D in multiple sclerosis, *Neuroepidemiology* 40 (2013) 147–153, <https://doi.org/10.1159/000345122>.
- [52] J.J. Cavanagh, M. Levy, Differential diagnosis of multiple sclerosis, *Presse Medicale Paris Fr.* 1983 50 (2021), <https://doi.org/10.1016/j.jpm.2021.104092>.
- [53] B.S. Travers, B.-K.-T. Tsang, J.L. Barton, Multiple sclerosis: Diagnosis, disease-modifying therapy and prognosis, *Aust. J. Gen. Pract.* 51 (2022) 199–206, <https://doi.org/10.31128/AJGP-07-21-6103>.
- [54] A.J. Thompson, B.L. Banwell, F. Barkhof, W.M. Carroll, T. Coetzee, G. Comi, J. Correale, F. Fazekas, M. Filippi, M.S. Freedman, K. Fujihara, S.L. Galetta, H. P. Hartung, L. Kappos, F.D. Lublin, R.A. Marrie, A.E. Miller, D.H. Miller, X. Montalban, E.M. Mowry, P.S. Sorensen, M. Tintoré, A.L. Traboulsee, M. Trojano, B.M.J. Uitendhaag, S. Vukusic, E. Waubant, B.G. Weinstenker, S. C. Reingold, J.A. Cohen, Diagnosis of multiple sclerosis: 2017 revisions of the McDonald criteria, *Lancet Neurol.* 17 (2018) 162–173, [https://doi.org/10.1016/S1474-4422\(17\)30470-2](https://doi.org/10.1016/S1474-4422(17)30470-2).
- [55] M. Gunnarsson, C. Malmström, M. Axelsson, P. Sundström, C. Dahle, M. Vrethem, T. Olsson, F. Piehl, N. Norgren, L. Rosengren, A. Svenningsson, J. Lycke, Axonal damage in relapsing multiple sclerosis is markedly reduced by natalizumab, *Ann. Neurol.* 69 (2011) 83–89, <https://doi.org/10.1002/ana.22247>.
- [56] J.L. Jones, J.M. Anderson, C.-L. Phuah, E.J. Fox, K. Selmaj, D. Margolin, S.L. Lake, J. Palmer, S.J. Thompson, A. Wilkins, D.J. Webber, D.A. Compston, A.J. Coles, Improvement in disability after alemtuzumab treatment of multiple sclerosis is associated with neuroprotective autoimmunity, *Brain J. Neurol.* 133 (2010) 2232–2247, <https://doi.org/10.1093/brain/awq176>.
- [57] H. Link, Y.-M. Huang, Oligoclonal bands in multiple sclerosis cerebrospinal fluid: An update on methodology and clinical usefulness, *J. Neuroimmunol.* 180 (2006) 17–28, <https://doi.org/10.1016/j.jneuroim.2006.07.006>.
- [58] A. Petzold, Intrathecal oligoclonal IgG synthesis in multiple sclerosis, *J. Neuroimmunol.* 262 (2013) 1–10, <https://doi.org/10.1016/j.jneuroim.2013.06.014>.
- [59] P.-P. Axisa, D.A. Hafler, Multiple Sclerosis: genetics, biomarkers, treatments, *Curr. Opin. Neurol.* 29 (2016) 345–353, <https://doi.org/10.1097/WCO.0000000000000319>.
- [60] H. Reiber, K. Felgenhauer, Protein transfer at the blood cerebrospinal fluid barrier and the quantitation of the humoral immune response within the central nervous system, *Clin. Chim. Acta Int. J. Clin. Chem.* 163 (1987) 319–328, [https://doi.org/10.1016/0009-8981\(87\)90250-6](https://doi.org/10.1016/0009-8981(87)90250-6).
- [61] W. Tourtelotte, On cerebrospinal fluid immunoglobulin-G (IgG) quotients in multiple sclerosis and other diseases. A review and a new formula to estimate the amount of IgG synthesized per day by the central nervous system, *J. Neurol. Sci.* 10 (1970) 279–304, [https://doi.org/10.1016/0022-510x\(70\)90156-5](https://doi.org/10.1016/0022-510x(70)90156-5).
- [62] E. Schuller, H.J. Sagar, Central nervous system IgG synthesis in multiple sclerosis. Application of a new formula, *Acta Neurol. Scand.* 67 (1983) 365–371, <https://doi.org/10.1111/j.1600-0404.1983.tb03154.x>.
- [63] H. Link, G. Tibbling, Principles of albumin and IgG analyses in neurological disorders. III. Evaluation of IgG synthesis within the central nervous system in multiple sclerosis, *Scand. J. Clin. Lab. Invest.* 37 (1977) 397–401, <https://doi.org/10.1080/00365517709091498>.
- [64] L. Villar, N. García-Barragán, M. Espiño, E. Roldán, M. Sádaba, J. Gómez-Rial, P. González-Porqué, J. Álvarez-Cermeño, Influence of oligoclonal IgM specificity in multiple sclerosis disease course, *Mult. Scler. J.* 14 (2008) 183–187, <https://doi.org/10.1177/1352458507082046>.
- [65] M.A. Rocca, M.P. Amato, N. De Stefano, C. Enzinger, J.J. Geurts, I.-K. Penner, A. Rovira, J.F. Sumowski, P. Valsasina, M. Filippi, MAGNIMS Study Group, Clinical and imaging assessment of cognitive dysfunction in multiple sclerosis, *Lancet Neurol.* 14 (2015) 302–317, [https://doi.org/10.1016/S1474-4422\(14\)70250-9](https://doi.org/10.1016/S1474-4422(14)70250-9).
- [66] M. Comabella, X. Montalban, Body fluid biomarkers in multiple sclerosis, *Lancet Neurol.* 13 (2014) 113–126, [https://doi.org/10.1016/S1474-4422\(13\)70233-3](https://doi.org/10.1016/S1474-4422(13)70233-3).
- [67] M. Bonnan, Intrathecal IgG synthesis: a resistant and valuable target for future multiple sclerosis treatments, *Mult. Scler. Int.* 2015 (2015), 296184, <https://doi.org/10.1155/2015/296184>.
- [68] P. Maggi, M. Absinta, M. Grammatico, L. Vuolo, G. Emmi, G. Carlucci, G. Spagni, A. Barilaro, A.M. Repice, L. Emmi, D. Prisco, V. Martinelli, R. Scotti, N. Sadeghi, G. Perrotta, P. Sati, B. Dachy, D.S. Reich, M. Filippi, L. Massacesi, Central vein sign differentiates Multiple Sclerosis from central nervous system inflammatory vasculopathies, *Ann. Neurol.* 83 (2018) 283–294, <https://doi.org/10.1002/ana.25146>.
- [69] T. Ziemssen, F. Ziemssen, The role of the humoral immune system in multiple sclerosis (MS) and its animal model experimental autoimmune encephalomyelitis

- (EAE), *Autoimmun. Rev.* 4 (2005) 460–467, <https://doi.org/10.1016/j.autrev.2005.03.005>.
- [70] D. Ferraro, A.M. Simone, R. Bedin, V. Galli, F. Vitetta, L. Federzoni, R. D'Amico, E. Merelli, P.F. Nichelli, P. Sola, Cerebrospinal fluid oligoclonal IgM bands predict early conversion to clinically definite multiple sclerosis in patients with Clinically Isolated Syndrome, *J. Neuroimmunol.* 257 (2013) 76–81, <https://doi.org/10.1016/j.jneuroim.2013.01.011>.
- [71] P. Perini, F. Ranzato, M. Calabrese, L. Battistin, P. Gallo, Intrathecal IgM production at clinical onset correlates with a more severe disease course in multiple sclerosis, *J. Neurol. Neurosurg. Psychiatry.* 77 (2006) 953–955, <https://doi.org/10.1136/jnnp.2005.086116>.
- [72] M. Fonderico, T. Biagioli, L. Lanzilao, A. Bellinvia, R. Fratangelo, L. Pastò, E. Prestipino, L. Razzolini, L. Tudisco, A. Ginestroni, L. Vuolo, E. Fainardi, C. Ballerini, E. Portaccio, M.P. Amato, Prognostic role of intrathecal IgM synthesis in multiple sclerosis: Results from a clinical series, *Mult. Scler. Houndmills Basingstoke Engl.* 27 (2021) 198–207, <https://doi.org/10.1177/1352458520907913>.
- [73] E. Monreal, S. Sainz de la Maza, L. Costa-Frossard, P. Walo-Delgado, J. Zamora, J. I. Fernández-Velasco, N. Villarrubia, M. Espiño, D. Lourido, P. Lapuente, I. Toboso, J.C. Álvarez-Cermeño, J. Masjuan, L.M. Villar, Predicting aggressive multiple sclerosis with intrathecal IgM synthesis among patients with a clinically isolated syndrome, *Neurol. Neuroimmunol. Neuroinflammation.* 8 (5) (2021) e1047, <https://doi.org/10.1212/NXI.0000000000001047>.
- [74] M.K. Sharief, G. Keir, E.J. Thompson, Intrathecal synthesis of IgM in neurological diseases: a comparison between detection of oligoclonal bands and quantitative estimation, *J. Neurol. Sci.* 96 (1990) 131–142, [https://doi.org/10.1016/0022-510x\(90\)90126-8](https://doi.org/10.1016/0022-510x(90)90126-8).
- [75] J.A. Brebner, R.A. Stockley, Polyclonal free light chains: a biomarker of inflammatory disease or treatment target?, *F1000 Med. Rep.* 5 (2013) 4, <https://doi.org/10.3410/M5-4>.
- [76] C. DeCarli, M.A. Menegus, R.A. Rudick, Free light chains in multiple sclerosis and infections of the CNS, *Neurology.* 37 (1987) 1334–1338, <https://doi.org/10.1212/wnl.37.8.1334>.
- [77] A.R. Bradwell, H.D. Carr-Smith, G.P. Mead, L.X. Tang, P.J. Showell, M. T. Drayson, R. Drew, Highly sensitive, automated immunoassay for immunoglobulin free light chains in serum and urine, *Clin. Chem.* 47 (2001) 673–680.
- [78] H.T. Velthuis, I. Knop, P. Stam, M. van den Broek, H.K. Bos, S. Hol, E. Teunissen, K.S. Fischechick, H. Althaus, B. Schmidt, C. Wagner, R. Melsert, N Latex FLC - new monoclonal high-performance assays for the determination of free light chain kappa and lambda, *Clin. Chem. Lab. Med.* 49 (2011) 1323–1332, <https://doi.org/10.1515/CCLM.2011.624>.
- [79] N.M.A. White-Al Habeeb, T. Earle, M. Spencer, I.M. Blasutig, Evaluation of the N-latex serum free light chain assay on the Siemens BNII analyzer and agreement with The Binding Site FreeLite assay on the SPAPlus, *Clin. Biochem.* 51 (2018) 90–96, <https://doi.org/10.1016/j.clinbiochem.2017.05.009>.
- [80] M. Daves, A. Piccin, V. Roccaforte, G. Lippi, Comparison of FreeLite and N-Latex serum free light chain assays: a critical review, *Biochem. Medica.* 31 (2021), 030701, <https://doi.org/10.11613/BM.2021.030701>.
- [81] L. Caponi, N. Romiti, E. Koni, A.D. Fiore, A. Paolicchi, M. Franzini, Inter-assay variability in automated serum free light chain assays and their use in the clinical laboratory, *Crit. Rev. Clin. Lab. Sci.* 57 (2020) 73–85, <https://doi.org/10.1080/10408363.2019.1670133>.
- [82] H. Hegen, J. Walde, D. Milosavljevic, F. Aboulenein-Djamshidian, M. Senel, H. Tumani, F. Deisenhammer, S. Presslauer, Free light chains in the cerebrospinal fluid. Comparison of different methods to determine intrathecal synthesis, *Clin. Chem. Lab. Med. CCLM* 57 (2019) 1574–1586, <https://doi.org/10.1515/cclm-2018-1300>.
- [83] D. Alves Martins, J. Lopes, A. Martins da Silva, C.I. Morais, J. Vasconcelos, I. Lima, C. Carneiro, E. Neves, Kappa free light chains: Diagnostic performance in multiple sclerosis and utility in a clinical laboratory, *Clin. Chim. Acta.* 528 (2022) 56–64, <https://doi.org/10.1016/j.cca.2022.01.017>.
- [84] F.F. Koenen, U. Wurster, T. Witte, K.F. Jendretzky, S. Gingele, H. Tumani, K.-W. Sühs, M. Stangel, P. Schwenkenbecher, T. Skripuletz, The impact of immunomodulatory treatment on kappa free light chains as biomarker in neuroinflammation, *Cells* 9 (2020) 842, <https://doi.org/10.3390/cells9040842>.
- [85] G. Passerini, G. Dalla Costa, F. Sangalli, L. Moiola, B. Colombo, M. Locatelli, G. Comi, R. Furlan, V. Martinelli, Free light chains and intrathecal B cells activity in multiple sclerosis: A prospective study and meta-analysis, *Mult. Scler. Int.* 2016 (2016) e2303857, <https://doi.org/10.1155/2016/2303857>.
- [86] B. Kaplan, S. Golderman, G. Yahalom, R. Yeskaraev, T. Ziv, B.M. Aizenbud, B.-A. Sela, A. Livneh, Free light chain monomer-dimer patterns in the diagnosis of multiple sclerosis, *J. Immunol. Methods.* 390 (2013) 74–80, <https://doi.org/10.1016/j.jim.2013.01.010>.
- [87] C. Leurs, H. Twaalfhoven, B. Lissenberg-Witte, V. van Pesch, I. Dujmovic, J. Drulovic, M. Castellazzi, T. Bellini, M. Pugliatti, J. Kuhle, L. Villar, J. Alvarez-Cermeño, R. Alvarez-Lafuente, H. Hegen, F. Deisenhammer, L. Walchhofer, E. Thouvenot, M. Comabella, X. Montalban, L. Vécsei, C. Rajda, D. Galimberti, E. Scarpini, A. Altintas, K. Rejdak, J. Frederiksen, G. Pihl-Jensen, P. Jensen, M. Khalil, M. Voortman, F. Fazekas, A. Saiz, D. La Puma, M. Vercaemmen, L. Vanopdenbosch, B. Uitdehaag, J. Killestein, C. Bridel, C. Teunissen, Kappa free light chains is a valid tool in the diagnostics of MS: A large multicenter study, *Mult. Scler. J.* 26 (2020) 912–923, <https://doi.org/10.1177/1352458519845844>.
- [88] M. Senel, F. Mojib-Yezdani, U. Braisch, F. Bachhuber, J. Lewerenz, A.C. Ludolph, M. Otto, H. Tumani, CSF free light chains as a marker of intrathecal immunoglobulin synthesis in multiple sclerosis: A blood-CSF barrier related evaluation in a large cohort, accessed November 22, 2022, *Front. Immunol.* 10 (2019), <https://www.frontiersin.org/articles/10.3389/fimmu.2019.00641>.
- [89] K.M. Gurtner, E. Shosha, S.C. Bryant, B.D. Andreguetto, D.L. Murray, S.J. Pittock, M.A.V. Willrich, CSF free light chain identification of demyelinating disease: comparison with oligoclonal banding and other CSF indexes, *Clin. Chem. Lab. Med. CCLM.* 56 (2018) 1071–1080, <https://doi.org/10.1515/cclm-2017-0901>.
- [90] I. Crespi, M.G. Sulas, R. Mora, P. Naldi, D. Vecchio, C. Comi, R. Cantello, G. Bellomo, Combined use of kappa free light chain index and isoelectrofocusing of cerebro-spinal fluid in diagnosing multiple sclerosis: Performances and costs, *Clin. Lab.* 63 (2017) 551–559, <https://doi.org/10.7754/Clin.Lab.2016.160930>.
- [91] G. Hassan-Smith, L. Durant, A. Tsentemidou, L.K. Assi, J.M. Faint, S. Kalra, M. R. Douglas, S.J. Curnow, High sensitivity and specificity of elevated cerebrospinal fluid kappa free light chains in suspected multiple sclerosis, *J. Neuroimmunol.* 276 (2014) 175–179, <https://doi.org/10.1016/j.jneuroim.2014.08.003>.
- [92] F. Duranti, M. Pieri, D. Centonze, F. Buttari, S. Bernardini, M. Dessi, Determination of kFLC and K Index in cerebrospinal fluid: A valid alternative to assess intrathecal immunoglobulin synthesis, *J. Neuroimmunol.* 263 (2013) 116–120, <https://doi.org/10.1016/j.jneuroim.2013.07.006>.
- [93] A. Emersic, V. Anadolli, M. Kršnik, U. Rot, Intrathecal immunoglobulin synthesis: The potential value of an adjunct test, *Clin. Chim. Acta.* 489 (2019) 109–116, <https://doi.org/10.1016/j.cca.2018.12.006>.
- [94] D. Zeman, Free light chains in the cerebrospinal fluid. Do we still need oligoclonal IgG? *Clin. Chem. Lab. Med. CCLM.* 56 (2018) 1011–1014, <https://doi.org/10.1515/cclm-2018-0096>.
- [95] P. Schwenkenbecher, F.F. Koenen, U. Wurster, T. Witte, S. Gingele, K.-W. Sühs, M. Stangel, T. Skripuletz, Reiber's diagram for kappa free light chains: the new standard for assessing intrathecal synthesis? *Diagnostics* 9 (2019) 194, <https://doi.org/10.3390/diagnostics9040194>.
- [96] M.M. Voortman, T. Stojakovic, L. Pirpamer, M. Jehna, C. Langkammer, H. Schrnagl, M. Reindl, S. Ropele, T. Seifert-Held, J.-J. Archelos, S. Fuchs, C. Enzinger, F. Fazekas, M. Khalil, Prognostic value of free light chains lambda and kappa in early multiple sclerosis, *Mult. Scler. Houndmills Basingstoke Engl.* 23 (2017) 1496–1505, <https://doi.org/10.1177/1352458516681503>.
- [97] L.M. Villar, M. Espiño, L. Costa-Frossard, A. Muriel, J. Jiménez, J.C. Alvarez-Cermeño, High levels of cerebrospinal fluid free kappa chains predict conversion to multiple sclerosis, *Clin. Chim. Acta Int. J. Clin. Chem.* 413 (2012) 1813–1816, <https://doi.org/10.1016/j.cca.2012.07.007>.
- [98] J.R. Rinker, K. Trinkaus, A.H. Cross, Elevated CSF free kappa light chains correlate with disability prognosis in multiple sclerosis, *Neurology.* 67 (2006) 1288–1290, <https://doi.org/10.1212/01.wnl.0000238107.31364.21>.
- [99] G. Makshakov, V. Nazarov, O. Kochetova, E. Surkova, S. Lapin, E. Evdoshenko, G. Saruhan-Direskenli, Diagnostic and prognostic value of the cerebrospinal fluid concentration of immunoglobulin free light chains in clinically isolated syndrome with conversion to multiple sclerosis, *PLoS One.* 10 (11) (2015) e0143375, <https://doi.org/10.1371/journal.pone.0143375>.
- [100] D. Vecchio, G. Bellomo, R. Serino, E. Virgilio, M. Lamona, U. Dianzani, R. Cantello, C. Comi, I. Crespi, Intrathecal kappa free light chains as markers for multiple sclerosis, *Sci. Rep.* 10 (2020) 20329, <https://doi.org/10.1038/s41598-020-77029-7>.
- [101] I. Rosenstein, M. Axelsson, L. Novakova, C. Malmeström, K. Blennow, H. Zetterberg, J. Lycke, Intrathecal kappa free light chain synthesis is associated with worse prognosis in relapsing-remitting multiple sclerosis, *J. Neurol.* (2023), <https://doi.org/10.1007/s00415-023-11817-9>.
- [102] L. Gaetani, M. Di Carlo, G. Brachelente, F. Valletta, P. Eusebi, A. Mancini, L. Gentili, A. Borrelli, P. Calabresi, P. Sarchielli, C. Ferri, A. Villa, M. Di Filippo, Cerebrospinal fluid free light chains compared to oligoclonal bands as biomarkers in multiple sclerosis, *J. Neuroimmunol.* 339 (2020), 577108, <https://doi.org/10.1016/j.jneuroim.2019.577108>.
- [103] S. Toscano, F. Patti, CSF biomarkers in multiple sclerosis: beyond neuroinflammation, *Neuroinflammation. Neuroinflammation.* 8 (2021) 14–41, <https://doi.org/10.20517/2347-8659.2020.12>.
- [104] D. Ferraro, R. Bedin, P. Natali, D. Franciotta, K. Smolik, M. Santangelo, P. Immovilli, V. Camera, F. Vitetta, M. Gastaldi, T. Trenti, S. Meletti, P. Sola, Kappa Index versus CSF oligoclonal bands in predicting multiple sclerosis and infectious/inflammatory CNS disorders, *Diagnostics* 10 (2020) 856, <https://doi.org/10.3390/diagnostics10100856>.
- [105] A. Yuan, M.V. Rao Veeranna, R.A. Nixon, Neurofilaments at a glance, *J. Cell Sci.* 125 (14) (2012) 3257–3263, <https://doi.org/10.1242/jcs.104729>.
- [106] L. Gaetani, K. Blennow, P. Calabresi, M.D. Filippo, L. Parnetti, H. Zetterberg, Neurofilament light chain as a biomarker in neurological disorders, *J. Neurol. Neurosurg. Psychiatry* 90 (2019) 870–881, <https://doi.org/10.1136/jnnp-2018-320106>.
- [107] G. Krishnamoorthy, A. Saxena, L.T. Mars, H.S. Domingues, R. Mentele, A. Ben-Nun, H. Lassmann, K. Dormair, F.C. Kuschus, R.S. Liblau, H. Wekerle, Myelin-specific T cells also recognize neuronal autoantigen in a transgenic mouse model of multiple sclerosis, *Nat. Med.* 15 (2009) 626–632, <https://doi.org/10.1038/nm.1975>.
- [108] S. Lerche, I. Wurster, B. Rösen, M. Zimmermann, G. Machetanz, S. Wiethoff, M. Dehnert, L. Rietschel, B. Riebenbauer, C. Deuschle, E. Stransky, I. Liepl-Scharfene, T. Gasser, K. Brockmann, CSF NFL in a longitudinally assessed PD Cohort: Age effects and cognitive trajectories, *Mov. Disord. Off. J. Mov. Disord. Soc.* 35 (2020) 1138–1144, <https://doi.org/10.1002/mds.28056>.
- [109] J. Kuhle, K. Plattner, J.P. Bestwick, R.L. Lindberg, S.V. Ramagopalan, N. Norgren, A. Nissim, A. Malaspina, D. Leppert, G. Giovannoni, L. Kappos, A comparative study of CSF neurofilament light and heavy chain protein in MS, *Mult. Scler.*

- Houndmills Basingstoke Engl. 19 (2013) 1597–1603, <https://doi.org/10.1177/1352458513482374>.
- [110] J. Kuhle, C. Malmestrom, M. Axelsson, K. Plattner, O. Yaldizli, T. Derfuss, G. Giovannoni, L. Kappos, J. Lycke, Neurofilament light and heavy subunits compared as therapeutic biomarkers in multiple sclerosis, *Acta Neurol. Scand.* 128 (2013) e33–e36, <https://doi.org/10.1111/ane.12151>.
- [111] J.N. Lycke, J.E. Karlsson, O. Andersen, L.E. Rosengren, Neurofilament protein in cerebrospinal fluid: a potential marker of activity in multiple sclerosis, *J. Neurol. Neurosurg. Psychiatry.* 64 (1998) 402–404, <https://doi.org/10.1136/jnnp.64.3.402>.
- [112] C.E. Teunissen, E. Iacobaeus, M. Khademi, L. Brundin, N. Norgren, M.J.A. Koel-Simmeling, M. Schepens, F. Bouwman, H.A.M. Twaalfhoven, H.J. Blom, C. Jakobs, C.D. Dijkstra, Combination of CSF N-acetylaspartate and neurofilaments in multiple sclerosis, *Neurology.* 72 (15) (2009) 1322–1329, <https://doi.org/10.1212/WNL.0b013e3181a0fe3f>.
- [113] G. Disanto, R. Adiatori, R. Dobson, V. Martinelli, G. Dalla Costa, T. Runia, E. Evdoshenko, E. Thouvenot, M. Trojano, N. Norgren, C. Teunissen, L. Kappos, G. Giovannoni, J. Kuhle, International Clinically Isolated Syndrome Study Group, Serum neurofilament light chain levels are increased in patients with a clinically isolated syndrome, *J. Neurol. Neurosurg. Psychiatry.* 87 (2016) 126–129, <https://doi.org/10.1136/jnnp-2014-309690>.
- [114] J. Kuhle, C. Barro, G. Disanto, A. Mathias, C. Soneson, G. Bonnier, Ö. Yaldizli, A. Regeniter, T. Derfuss, M. Canales, M. Schlupe, R. Du Pasquier, G. Krueger, C. Granziera, Serum neurofilament light chain in early relapsing remitting MS is increased and correlates with CSF levels and with MRI measures of disease severity, *Mult. Scler. Houndmills Basingstoke Engl.* 22 (2016) 1550–1559, <https://doi.org/10.1177/1352458515623365>.
- [115] J. Kuhle, C. Barro, U. Andreasson, T. Derfuss, R. Lindberg, Å. Sandelius, V. Liman, N. Norgren, K. Blennow, H. Zetterberg, Comparison of three analytical platforms for quantification of the neurofilament light chain in blood samples: ELISA, electrochemoluminescence immunoassay and Simoa, *Clin. Chem. Lab. Med.* 54 (2016) 1655–1661, <https://doi.org/10.1515/cclm-2015-1195>.
- [116] C.E. Teunissen, M. Khalil, Neurofilaments as biomarkers in multiple sclerosis, *Mult. Scler. Houndmills Basingstoke Engl.* 18 (2012) 552–556, <https://doi.org/10.1177/1352458512443092>.
- [117] Y.K. Semra, O.A. Seidi, M.K. Sharief, Heightened intrathecal release of axonal cytoskeletal proteins in multiple sclerosis is associated with progressive disease and clinical disability, *J. Neuroimmunol.* 122 (2002) 132–139, [https://doi.org/10.1016/S0165-5728\(01\)00455-6](https://doi.org/10.1016/S0165-5728(01)00455-6).
- [118] J. Salzer, A. Svenningsson, P. Sundström, Neurofilament light as a prognostic marker in multiple sclerosis, *Mult. Scler. Houndmills Basingstoke Engl.* 16 (2010) 287–292, <https://doi.org/10.1177/1352458509359725>.
- [119] J. Burman, H. Zetterberg, M. Fransson, A.S. Loskog, R. Raininko, J. Fagius, Assessing tissue damage in multiple sclerosis: a biomarker approach, *Acta Neurol. Scand.* 130 (2014) 81–89, <https://doi.org/10.1111/ane.12239>.
- [120] L. Novakova, H. Zetterberg, P. Sundström, M. Axelsson, M. Khademi, M. Gunnarsson, C. Malmestrom, A. Svenningsson, T. Olsson, F. Piehl, K. Blennow, J. Lycke, Monitoring disease activity in multiple sclerosis using serum neurofilament light protein, *Neurology.* 89 (2017) 2230–2237, <https://doi.org/10.1212/WNL.0000000000004683>.
- [121] C. Barro, P. Benkert, G. Disanto, C. Tsagkas, M. Amann, Y. Naegelin, D. Leppert, C. Gobbi, C. Granziera, Ö. Yaldizli, Z. Michalak, J. Wuerfel, L. Kappos, K. Parmar, J. Kuhle, Serum neurofilament as a predictor of disease worsening and brain and spinal cord atrophy in multiple sclerosis, *Brain J. Neurol.* 141 (2018) 2382–2391, <https://doi.org/10.1093/brain/awy154>.
- [122] G. Disanto, C. Barro, P. Benkert, Y. Naegelin, S. Schädelin, A. Giardiello, C. Zecca, K. Blennow, H. Zetterberg, D. Leppert, L. Kappos, C. Gobbi, J. Kuhle, *Swiss Multiple Sclerosis Cohort Study Group*, Serum Neurofilament light: A biomarker of neuronal damage in multiple sclerosis, *Ann. Neurol.* 81 (6) (2017) 857–870.
- [123] J. Romme Christensen, R. Ratzler, L. Börnsen, M. Lyksborg, E. Garde, T.B. Dyrby, H.R. Siebner, P.S. Sorensen, F. Sellebjerg, Natalizumab in progressive MS: results of an open-label, phase 2A, proof-of-concept trial, *Neurology.* 82 (2014) 1499–1507, <https://doi.org/10.1212/WNL.0000000000000361>.
- [124] M. Axelsson, C. Malmestrom, M. Gunnarsson, H. Zetterberg, P. Sundström, J. Lycke, A. Svenningsson, Immunosuppressive therapy reduces axonal damage in progressive multiple sclerosis, *Mult. Scler. Houndmills Basingstoke Engl.* 20 (2014) 43–50, <https://doi.org/10.1177/1352458513490544>.
- [125] J. Kuhle, G. Disanto, J. Lorscheider, T. Stites, Y. Chen, F. Dahlke, G. Francis, A. Shrinivasan, E.-W. Radue, G. Giovannoni, L. Kappos, Fingolimod and CSF neurofilament light chain levels in relapsing-remitting multiple sclerosis, *Neurology.* 84 (2015) 1639–1643, <https://doi.org/10.1212/WNL.0000000000001491>.
- [126] D. Pelletier, D.A. Hafler, Fingolimod for multiple sclerosis, *N. Engl. J. Med.* 366 (2012) 339–347, <https://doi.org/10.1056/NEJMc1101691>.
- [127] E. Cantó, C. Barro, C. Zhao, S.J. Caillier, Z. Michalak, R. Bove, D. Tomić, A. Santaniello, D.A. Häring, J. Hollenbach, R.G. Henry, B.A.C. Cree, L. Kappos, D. Leppert, S.L. Hauser, P. Benkert, J.R. Oksenberg, J. Kuhle, Association between serum neurofilament light chain levels and long-term disease course among patients with multiple sclerosis followed up for 12 Years, *JAMA Neurol.* 76 (2019) 1359–1366, <https://doi.org/10.1001/jamaneuro.2019.2137>.
- [128] D. Ferraro, C. Guicciardi, S. De Biasi, M. Pinti, R. Bedin, V. Camera, F. Vitetta, M. Nasi, S. Meletti, P. Sola, Plasma neurofilaments correlate with disability in progressive multiple sclerosis patients, *Acta Neurol. Scand.* 141 (2020) 16–21, <https://doi.org/10.1111/ane.13152>.
- [129] Z.L.E. van Kempen, R.J. Kryscio, G.D. Costa, Serum neurofilament light as a prognostic marker for MS disability: Are we there yet? *Neurology.* 94 (2020) 1013–1014, <https://doi.org/10.1212/WNL.0000000000009576>.
- [130] M.D. Weingarten, A.H. Lockwood, S.Y. Hwo, M.W. Kirschner, A protein factor essential for microtubule assembly, *Proc. Natl. Acad. Sci. U. S. A.* 72 (1975) 1858–1862, <https://doi.org/10.1073/pnas.72.5.1858>.
- [131] M. Sjögren, H. Vanderstichele, H. Agren, O. Zachrisson, M. Edsbacke, C. Wikkelso, I. Skoog, A. Wallin, L.O. Wahlund, J. Marcusson, K. Nägga, N. Andreasen, P. Davidsson, E. Vanmechelen, K. Blennow, Tau and Abeta42 in cerebrospinal fluid from healthy adults 21–93 years of age: establishment of reference values, *Clin. Chem.* 47 (2001) 1776–1781.
- [132] L.I. Binder, A. Frankfurter, L.I. Rebhun, The distribution of tau in the mammalian central nervous system, *J. Cell Biol.* 101 (1985) 1371–1378, <https://doi.org/10.1083/jcb.101.4.1371>.
- [133] K. Rotasy, E. Withut, D. Pohl, P. Lange, B. Ciesielczyk, R. Diem, J. Gärtner, M. Otto, Tau, phospho-tau, and S-100B in the cerebrospinal fluid of children with multiple sclerosis, *J. Child Neurol.* 20 (2005) 822–825, <https://doi.org/10.1177/08830738050200100801>.
- [134] A. Martínez-Yélamos, A. Saiz, J. Bas, J.J. Hernandez, F. Graus, T. Arbizu, Tau protein in cerebrospinal fluid: a possible marker of poor outcome in patients with early relapsing-remitting multiple sclerosis, *Neurosci. Lett.* 363 (2004) 14–17, <https://doi.org/10.1016/j.neulet.2004.03.039>.
- [135] K. Hein (née Maier), A. Köhler, R. Diem, M.B. Sättler, I. Demmer, P. Lange, M. Bähr, M. Otto, Biological markers for axonal degeneration in CSF and blood of patients with the first event indicative for multiple sclerosis, *Neurosci. Lett.* 436 (1) (2008) 72–76, <https://doi.org/10.1016/j.neulet.2008.02.064>.
- [136] I. Guimarães, M.I. Cardoso, M.J. Sá, Tau protein seems not to be a useful routine clinical marker of axonal damage in multiple sclerosis, *Mult. Scler. Houndmills Basingstoke Engl.* 12 (2006) 354–356, <https://doi.org/10.1191/1352458506ms1288sr>.
- [137] M. Storoni, M.M. Verbeek, Z. Illes, R. Marignier, C.E. Teunissen, M. Grabowska, C. Confavreux, G.T. Plant, A. Petzold, Serum GFAP levels in optic neuropathies, *J. Neurol. Sci.* 317 (2012) 117–122, <https://doi.org/10.1016/j.jns.2012.02.012>.
- [138] T. Mitsu, R. Takano, K. Fujihara, T. Takahashi, S. Sato, Y. Itoyama, Marked increase in cerebrospinal fluid glial fibrillary acidic protein in neuromyelitis optica: an astrocytic damage marker, *J. Neurol. Neurosurg. Psychiatry.* 80 (2009) 575–577, <https://doi.org/10.1136/jnnp.2008.150698>.
- [139] R. Diaz-Arrostia, K.K.W. Wang, L. Papa, M.D. Sorani, J.K. Yue, A.M. Puccio, P. J. McMahon, T. Inoue, E.L. Yuh, H.F. Lingsma, A.I.R. Maas, A.B. Valadka, D. O. Okonkwo, G.T. Manley and the TRACK-TBI Investigat, I.S.S. Casey, M. Cheong, S.R. Cooper, K. Dams-O'Connor, W.A. Gordon, A.J. Hricik, D.K. Menon, S. Mukherjee, D.M. Schnyer, T.K. Sinha, M.J. Vassar, Acute biomarkers of traumatic brain injury: Relationship between plasma levels of ubiquitin C-terminal hydrolase-L1 and glial fibrillary acidic protein, *J. Neurotrauma.* 31 (1) (2014) 19–25, <https://doi.org/10.1089/neu.2013.3040>.
- [140] J.J. Bazarian, P. Biberthaler, R.D. Welch, L.M. Lewis, P. Barzo, V. Bogner-Flatz, P. Gunnar Brolinson, A. Büki, J.Y. Chen, R.H. Christenson, D. Hack, J.S. Huff, S. Johar, J.D. Jordan, B.A. Leidel, T. Lindner, E. Ludington, D.O. Okonkwo, J. Ornato, W.F. Peacock, K. Schmidt, J.A. Tyndall, A. Vossough, A.S. Jagoda, Serum GFAP and UCH-L1 for prediction of absence of intracranial injuries on head CT (ALERT-TBI): a multicentre observational study, *Lancet Neurol.* 17 (2018) 782–789, [https://doi.org/10.1016/S1474-4422\(18\)30231-X](https://doi.org/10.1016/S1474-4422(18)30231-X).
- [141] X. Ayrignac, E. Le Bars, C. Duflos, C. Hirtz, A. Maleska Maceski, C. Carra-Dallière, M. Charif, F. Pinna, P. Prin, N. Menjot de Champfleur, J. Deverdun, T. Kober, B. Marechal, M.J. Fartaria, R. Corredor Jerez, P. Labauge, S. Lehmann, Serum GFAP in multiple sclerosis: correlation with disease type and MRI markers of disease severity, *Sci. Rep.* 10 (2020) 10923, <https://doi.org/10.1038/s41598-020-67934-2>.
- [142] H. Högel, E. Rissanen, C. Barro, M. Matilainen, M. Nylund, J. Kuhle, L. Airas, Serum glial fibrillary acidic protein correlates with multiple sclerosis disease severity, *Mult. Scler. J.* 26 (2020) 210–219, <https://doi.org/10.1177/1352458518819380>.
- [143] A. Huss, M. Otto, M. Senel, A.C. Ludolph, A. Abdelhak, H. Tumani, A Score Based on NFL and Glial Markers May Differentiate Between Relapsing-Remitting and Progressive MS Course, accessed November 22, 2022, *Front. Neurol.* 11 (2020), <https://www.frontiersin.org/articles/10.3389/fneur.2020.00608>.
- [144] X. Yang, Q. Huang, H. Yang, S.i. Liu, B. Chen, T. Liu, J. Yang, H. Yao, S. Lin, X. Chen, H. Zhuang, Y. long, C. Gao, Astrocytic damage in glial fibrillary acidic protein astrocytopathy during initial attack, *Mult. Scler. Relat. Disord.* 29 (2019) 94–99, <https://doi.org/10.1016/j.msard.2019.01.036>.
- [145] A. Abdelhak, A. Huss, J. Kassubek, H. Tumani, M. Otto, Serum GFAP as a biomarker for disease severity in multiple sclerosis, *Sci. Rep.* 8 (2018) 14798, <https://doi.org/10.1038/s41598-018-33158-8>.
- [146] M.A. Mané-Martínez, B. Olsson, L. Bau, E. Matas, Á. Cobo-Calvo, U. Andreasson, K. Blennow, L. Romero-Pinel, S. Martínez-Yélamos, H. Zetterberg, Glial and neuronal markers in cerebrospinal fluid in different types of multiple sclerosis, *J. Neuroimmunol.* 299 (2016) 112–117, <https://doi.org/10.1016/j.jneuroim.2016.08.004>.
- [147] M. Axelsson, C. Malmestrom, S. Nilsson, S. Haghighi, L. Rosengren, J. Lycke, Glial fibrillary acidic protein: a potential biomarker for progression in multiple sclerosis, *J. Neurol.* 258 (2011) 882–888, <https://doi.org/10.1007/s00415-010-5863-2>.
- [148] S. Haghighi, O. Andersen, A. Odén, L. Rosengren, Cerebrospinal fluid markers in MS patients and their healthy siblings, *Acta Neurol. Scand.* 109 (2004) 97–99, <https://doi.org/10.1034/j.1600-0404.2003.00197.x>.

- [149] C. Malmström, S. Haghghi, L. Rosengren, O. Andersen, J. Lycke, Neurofilament light protein and glial fibrillary acidic protein as biological markers in MS, *Neurology*. 61 (2003) 1720–1725, <https://doi.org/10.1212/01.WNL.0000098880.19793.B6>.
- [150] A. Petzold, M.J. Eikelenboom, D. Gveric, G. Keir, M. Chapman, R.H.C. Lazeron, M. L. Cuzner, C.H. Polman, B.M.J. Uitdehaag, E.J. Thompson, G. Giovannoni, Markers for different glial cell responses in multiple sclerosis: clinical and pathological correlations, *Brain*. 125 (2002) 1462–1473, <https://doi.org/10.1093/brain/awf165>.
- [151] R. Madeddu, C. Farace, P. Tolu, G. Solinas, Y. Asara, M.A. Sotgiu, L.G. Delogu, J. C. Prados, S. Sotgiu, A. Montella, Cytoskeletal proteins in the cerebrospinal fluid as biomarker of multiple sclerosis, *Neurol. Sci.* 34 (2013) 181–186, <https://doi.org/10.1007/s10072-012-0974-4>.
- [152] X. Jiang, C. Shen, C.E. Teunissen, M. Wessels, H. Zetterberg, G. Giovannoni, C. M. Singh, B. Caba, C. Elliott, E. Fisher, C. de Moor, S. Belachew, A.R. Gafson, Glial fibrillary acidic protein and multiple sclerosis progression independent of acute inflammation, *Mult. Scler. Houndmills Basingstoke Engl.* (2023), 13524585231176732, <https://doi.org/10.1177/13524585231176732>.
- [153] R. Gerlach, G. Demel, H.-G. König, U. Gross, J.H.M. Prehn, A. Raabe, V. Seifert, D. Kögel, Active secretion of S100B from astrocytes during metabolic stress, *Neuroscience* 141 (2006) 1697–1701, <https://doi.org/10.1016/j.neuroscience.2006.05.008>.
- [154] S. Hachem, A. Aguirre, V. Vives, A. Marks, V. Gallo, C. Legraverend, Spatial and temporal expression of S100B in cells of oligodendrocyte lineage, *Glia*. 51 (2005) 81–97, <https://doi.org/10.1002/glia.20184>.
- [155] R. Donato, G. Sorci, F. Riuzzi, C. Arcuri, R. Bianchi, F. Brozzi, C. Tubaro, I. Giambanco, S100b's double life: Intracellular regulator and extracellular signal, *Biochim. Biophys. Acta BBA - Mol. Cell Res.* 1793 (6) (2009) 1008–1022, <https://doi.org/10.1016/j.bbamer.2008.11.009>.
- [156] J. Baudier, D. Mochly-Rosen, A. Newton, S.H. Lee, D.E. Koshland, R.D. Cole, Comparison of S100b protein with calmodulin: interactions with melittin and microtubule-associated tau proteins and inhibition of phosphorylation of tau proteins by protein kinase C, *Biochemistry*. 26 (10) (1987) 2886–2893, <https://doi.org/10.1021/bi00384a033>.
- [157] L. Zhang, W. Liu, D. Alizadeh, D. Zhao, O. Farrukh, J. Lin, S.A. Badie, B. Badie, S100B attenuates microglia activation in gliomas: Possible role of STAT3 pathway, *Glia*. 59 (2011) 486–498, <https://doi.org/10.1002/glia.21118>.
- [158] C. Reali, F. Scintu, R. Pillai, R. Donato, F. Michetti, V. Sogos, S100b counteracts effects of the neurotoxicant trimethyltin on astrocytes and microglia, *J. Neurosci. Res.* 81 (2005) 677–686, <https://doi.org/10.1002/jnr.20584>.
- [159] D.S. Gonçalves, G. Lenz, J. Karl, C.A. Gonçalves, R. Rodnight, Extracellular S100b protein modulates ERK in astrocyte cultures, *NeuroReport*. 11 (2000) 807.
- [160] R.H. Selinfreund, S.W. Barger, W.J. Pledger, L.J. Van Eldik, Neurotrophic protein S100 beta stimulates glial cell proliferation, *Proc. Natl. Acad. Sci.* 88 (1991) 3554–3558, <https://doi.org/10.1073/pnas.88.9.3554>.
- [161] F. Michetti, A. Massaro, M. Murazio, The nervous system-specific S-100 antigen in cerebrospinal fluid of multiple sclerosis patients, *Neurosci. Lett.* 11 (1979) 171–175, [https://doi.org/10.1016/0304-3940\(79\)90122-8](https://doi.org/10.1016/0304-3940(79)90122-8).
- [162] A. Barateiro, V. Afonso, G. Santos, J.J. Cerqueira, D. Brites, J. van Horsen, A. Fernandes, S100B as a potential biomarker and therapeutic target in multiple sclerosis, *Mol. Neurobiol.* 53 (2016) 3976–3991, <https://doi.org/10.1007/s12035-015-9336-6>.
- [163] M.T. Wunderlich, A.D. Ebert, T. Kratz, M. Goertler, S. Jost, M. Herrmann, Early neurobehavioral outcome after stroke is related to release of neurobiochemical markers of brain damage, *Stroke*. 30 (1999) 1190–1195, <https://doi.org/10.1161/01.str.30.6.1190>.
- [164] B.J. Steinhoff, H. Tumani, M. Otto, K. Mursch, J. Wiltfang, G. Herrendorf, H.-J. Bittermann, K. Felgenhauer, W. Paulus, E. Markakis, Cisternal S100 protein and neuron-specific enolase are elevated and site-specific markers in intractable temporal lobe epilepsy, *Epilepsy Res.* 36 (1999) 75–82, [https://doi.org/10.1016/S0920-1211\(99\)00026-1](https://doi.org/10.1016/S0920-1211(99)00026-1).
- [165] M. Herrmann, S. Jost, S. Kutz, A.D. Ebert, T. Kratz, M.T. Wunderlich, H. Synowitz, Temporal profile of release of neurobiochemical markers of brain damage after traumatic brain injury is associated with intracranial pathology as demonstrated in cranial computerized tomography, *J. Neurotrauma*. 17 (2000) 113–122, <https://doi.org/10.1089/neu.2000.17.113>.
- [166] H. Bartosik-Psujek, M. Psujek, J. Jaworski, Z. Stelmasiak, Total tau and S100b proteins in different types of multiple sclerosis and during immunosuppressive treatment with mitoxantrone, *Acta Neurol. Scand.* 123 (2011) 252–256, <https://doi.org/10.1111/j.1600-0404.2010.01393.x>.
- [167] K.E. O'Connell, T. Mok, B. Sweeney, A.M. Ryan, K.K. Dev, The use of cytokine signature patterns: separating drug naïve, interferon and natalizumab-treated multiple sclerosis patients, *Autoimmunity*. 47 (2014) 505–511, <https://doi.org/10.3109/08916934.2014.930734>.
- [168] M.L. Cuzner, W.T. Norton, Biochemistry of demyelination, *Brain Pathol. Zurich Switz.* 6 (1996) 231–242, <https://doi.org/10.1111/j.1750-3639.1996.tb00852.x>.
- [169] C. Zhang, A.K. Walker, R. Zand, M.A. Moscarello, J.M. Yan, P.C. Andrews, Myelin basic protein undergoes a broader range of modifications in mammals than in lower vertebrates, *J. Proteome Res.* 11 (2012) 4791–4802, <https://doi.org/10.1021/pr201196e>.
- [170] Y. Hu, I. Doudevski, D. Wood, M. Moscarello, C. Husted, C. Genain, J. A. Zasadzinski, J. Israelachvili, Synergistic interactions of lipids and myelin basic protein, *Proc. Natl. Acad. Sci. U. S. A.* 101 (2004) 13466–13471, <https://doi.org/10.1073/pnas.0405665101>.
- [171] S.R. Cohen, M.J. Brune, R.M. Herndon, G.M. McKhann, Cerebrospinal fluid myelin basic protein and multiple sclerosis, *Adv. Exp. Med. Biol.* 100 (1978) 513–519, https://doi.org/10.1007/978-1-4684-2514-7_37.
- [172] S.D. Levin, N.R. Hoyle, J.K. Brown, D.G. Thomas, Cerebrospinal fluid myelin basic protein immunoreactivity as an indicator of brain damage in children, *Dev. Med. Child Neurol.* 27 (1985) 807–813, <https://doi.org/10.1111/j.1469-8749.1985.tb03806.x>.
- [173] T.W. Noseworthy, B.J. Anderson, A.F. Noseworthy, A. Shustack, R.G. Johnston, K. C. Petruk, T.A. McPherson, Cerebrospinal fluid myelin basic protein as a prognostic marker in patients with head injury, *Crit. Care Med.* 13 (1985) 743–746, <https://doi.org/10.1097/00003246-198509000-00010>.
- [174] A. Re, R. Giachetti, Acute disseminated encephalomyelitis (ADEM) after autologous peripheral blood stem cell transplant for non-Hodgkin's lymphoma, *Bone Marrow Transplant.* 24 (1999) 1351–1354, <https://doi.org/10.1038/sj.bmt.1702047>.
- [175] H. Koshihara, K. Oguchi, Y. Takei, K. Kitazawa, K. Higuchi, S. Ohara, Meningeal inflammation and demyelination in a patient clinically diagnosed with acute disseminated encephalomyelitis, *J. Neurol. Sci.* 346 (2014) 323–327, <https://doi.org/10.1016/j.jns.2014.08.037>.
- [176] C. Jacque, A. Delassalle, G. Rancurel, M. Raoul, B. Lesourd, J.C. Legrand, Myelin basic protein in CSF and blood. Relationship between its presence and the occurrence of a destructive process in the brains of encephalitic patients, *Arch. Neurol.* 39 (1982) 557–560, <https://doi.org/10.1001/archneur.1982.00510210027006>.
- [177] T. Strand, C. Alling, B. Karlsson, I. Karlsson, B. Winblad, Brain and plasma proteins in spinal fluid as markers for brain damage and severity of stroke, *Stroke*. 15 (1984) 138–144, <https://doi.org/10.1161/01.str.15.1.138>.
- [178] M. Ohta, H. Nishitani, F. Matsubara, G. Inaba, Myelin basic protein in spinal fluid from patients with neuro-Behcet's disease, *N. Engl. J. Med.* 302 (1980) 1093, <https://doi.org/10.1056/NEJM198005083021922>.
- [179] H. Wekerle, H. Lassmann, The immunology of inflammatory demyelinating disease, *McAlpines Mult. Scler.* (2006) 491–555, <https://doi.org/10.1016/B978-0-443-07271-0.50013-6>.
- [180] A.P. Nicholas, T. Sambandam, J.D. Echols, S.R. Barnum, Expression of citrullinated proteins in murine experimental autoimmune encephalomyelitis, *J. Comp. Neurol.* 486 (2005) 254–266, <https://doi.org/10.1002/cne.20527>.
- [181] R. Rajmakers, J. Vogelzangs, J.L. Croxford, P. Wesseling, W.J. van Venrooij, G.J. M. Pruijn, Citrullination of central nervous system proteins during the development of experimental autoimmune encephalomyelitis, *J. Comp. Neurol.* 486 (2005) 243–253, <https://doi.org/10.1002/cne.20529>.
- [182] M.A. Moscarello, D.D. Wood, C. Ackerley, C. Boulias, Myelin in multiple sclerosis is developmentally immature, *J. Clin. Invest.* 94 (1994) 146–154, <https://doi.org/10.1172/JCI117300>.
- [183] D.D. Wood, J.M. Bilbao, P. O'Connors, M.A. Moscarello, Acute multiple sclerosis (Marburg type) is associated with developmentally immature myelin basic protein, *Ann. Neurol.* 40 (1996) 18–24, <https://doi.org/10.1002/ana.410400106>.
- [184] V. Martinsen, P. Kursula, Multiple sclerosis and myelin basic protein: insights into protein disorder and disease, *Amino Acids*. 54 (2022) 99–109, <https://doi.org/10.1007/s00726-021-03111-7>.
- [185] T. Berger, P. Rubner, F. Schautzer, R. Egg, H. Ulmer, I. Mayringer, E. Dilitz, F. Deisenhammer, M. Reindl, Antimyelin antibodies as a predictor of clinically definite multiple sclerosis after a first demyelinating event, *N. Engl. J. Med.* 349 (2003) 139–145, <https://doi.org/10.1056/NEJMoa022328>.
- [186] J. Kuhle, C. Pohl, M. Mehling, G. Edan, M.S. Freedman, H.-P. Hartung, C. H. Polman, D.H. Miller, X. Montalban, F. Barkhof, L. Bauer, S. Dahms, R. Lindberg, L. Kappos, R. Sandbrink, Lack of association between antimyelin antibodies and progression to multiple sclerosis, *N. Engl. J. Med.* 356 (2007) 371–378, <https://doi.org/10.1056/NEJMoa063602>.
- [187] F. Llorens, K. Thüne, W. Tahir, E. Kanata, D. Diaz-Lucena, K. Xanthopoulos, E. Kovatsi, C. Pleschka, P. Garcia-Esparcia, M. Schmitz, D. Ozbay, S. Correia, A. Correia, I. Milosevic, O. Andréoletti, N. Fernández-Borges, I.M. Vorberg, M. Zlatzer, T. Sklavadiadis, J.M. Torres, S. Krasemann, R. Sánchez-Valle, I. Ferrer, I. Rerz, YKL-40 in the brain and cerebrospinal fluid of neurodegenerative dementias, *Mol. Neurodegener.* 12 (2017) 83, <https://doi.org/10.1186/s13024-017-0226-4>.
- [188] C.G. Lee, C.A. Da Silva, C.S. Dela Cruz, F. Ahangari, B. Ma, M.-J. Kang, C.-H. He, S. Takyar, J.A. Elias, Role of chitin and chitinase/chitinase-like proteins in inflammation, tissue remodeling, and injury, *Annu. Rev. Physiol.* 73 (2011) 479–501, <https://doi.org/10.1146/annurev-physiol-012110-142250>.
- [189] D. Bonne-Barkay, S.J. Bissel, J. Kofler, A. Starkey, G. Wang, C.A. Wiley, Astrocyte and macrophage regulation of YKL-40 expression and cellular response in neuroinflammation, *Brain Pathol. Zurich Switz.* 22 (2012) 530–546, <https://doi.org/10.1111/j.1750-3639.2011.00550.x>.
- [190] J. Burman, R. Raininko, K. Blennow, H. Zetterberg, M. Axelsson, C. Malmström, YKL-40 is a CSF biomarker of intrathecal inflammation in secondary progressive multiple sclerosis, *J. Neuroimmunol.* 292 (2016) 52–57, <https://doi.org/10.1016/j.jneuroim.2016.01.013>.
- [191] S. Modvig, M. Degn, H. Roed, T.L. Sørensen, H.B.W. Larsson, A.R. Langkilde, J. L. Frederiksen, F. Sellebjerg, Cerebrospinal fluid levels of chitinase 3-like 1 and neurofilament light chain predict multiple sclerosis development and disability after optic neuritis, *Mult. Scler. Houndmills Basingstoke Engl.* 21 (2015) 1761–1770, <https://doi.org/10.1177/1352458515574148>.
- [192] F. Baldacci, S. Lista, G. Palermo, F.S. Giorgi, A. Vergallo, H. Hampel, The neuroinflammatory biomarker YKL-40 for neurodegenerative diseases: advances in development, *Expert Rev. Proteomics*. 16 (2019) 593–600, <https://doi.org/10.1080/14789450.2019.1628643>.

- [193] P. Kušnierová, D. Zeman, P. Hradělek, O. Zapletalová, D. Stejskal, C. Liguori, Determination of chitinase 3-like 1 in cerebrospinal fluid in multiple sclerosis and other neurological diseases, *PLoS ONE*. 15 (5) (2020) e0233519, <https://doi.org/10.1371/journal.pone.0233519>.
- [194] M. Comabella, M. Fernández, R. Martín, S. Rivera-Vallvé, E. Borrás, C. Chiva, E. Julià, A. Rovira, E. Cantó, J.C. Alvarez-Cermeño, L.M. Villar, M. Tintoré, X. Montalban, Cerebrospinal fluid chitinase 3-like 1 levels are associated with conversion to multiple sclerosis, *Brain, J. Neurol.* 133 (2010) 1082–1093, <https://doi.org/10.1093/brain/awq035>.
- [195] S. Floro, T. Carandini, A.M. Pietroboni, M.A. De Riz, E. Scarpini, D. Galimberti, Role of chitinase 3-like 1 as a biomarker in multiple sclerosis, *Neurol. Neuroimmunol. Neuroinflammation*. 9 (4) (2022) e1164, <https://doi.org/10.1212/NXI.0000000000001164>.
- [196] R. Schneider, B. Bellenberg, B. Gisevius, S. Hirschberg, R. Sankowski, M. Prinz, R. Gold, C. Lukas, A. Haghikia, Chitinase 3-like 1 and neurofilament light chain in CSF and CNS atrophy in MS, *Neurol. Neuroimmunol. Neuroinflammation*. 8 (1) (2021) e906, <https://doi.org/10.1212/NXI.0000000000000906>.
- [197] E. Cantó, M. Tintoré, L.M. Villar, C. Costa, R. Nurdinov, J.C. Álvarez-Cermeño, G. Arrambide, F. Reverter, F. Deisenhammer, H. Hegen, M. Khademi, T. Olsson, H. Tumani, E. Rodríguez-Martín, F. Piehl, A. Bartos, D. Zimova, J. Kotoucová, J. Kuhle, L. Kappos, J.A. García-Merino, A.J. Sánchez, A. Saiz, Y. Blanco, R. Hintzen, N. Jafari, D. Brassat, F. Lauda, R. Roesler, K. Rejdak, E. Papuc, C. de Andrés, S. Rauch, M. Khalil, C. Enzinger, D. Galimberti, E. Scarpini, C. Teunissen, A. Sánchez, A. Rovira, X. Montalban, M. Comabella, Chitinase 3-like 1: prognostic biomarker in clinically isolated syndromes, *Brain J. Neurol.* 138 (2015) 918–931, <https://doi.org/10.1093/brain/awv017>.
- [198] C. Matute-Blanch, J. Río, L.M. Villar, L. Midaglia, S. Malhotra, J.C. Álvarez-Cermeño, A. Vidal-Jordana, X. Montalban, M. Comabella, Chitinase 3-like 1 is associated with the response to interferon-beta treatment in multiple sclerosis, *J. Neuroimmunol.* 303 (2017) 62–65, <https://doi.org/10.1016/j.jneuroim.2016.12.006>.
- [199] M. Braitch, C.S. Constantinescu, The role of osteopontin in experimental autoimmune encephalomyelitis (EAE) and multiple sclerosis (MS), *Inflamm. Allergy Drug Targets* 9 (2010) 249–256, <https://doi.org/10.2174/187152810793358778>.
- [200] C. Sinclair, M. Mirakhor, J. Kirk, M. Farrell, S. McQuaid, Up-regulation of osteopontin and α B-crystallin in the normal-appearing white matter of multiple sclerosis: an immunohistochemical study utilizing tissue microarrays, *Neuropathol. Appl. Neurobiol.* 31 (2005) 292–303, <https://doi.org/10.1111/j.1365-2990.2004.00638.x>.
- [201] G. Orsi, T. Cseh, Z. Hayden, G. Perlaki, S.A. Nagy, O. Giyab, D.A. Olsen, J. S. Madsen, T. Berki, Z. Illes, Microstructural and functional brain abnormalities in multiple sclerosis predicted by osteopontin and neurofilament light, *Mult. Scler. Relat. Disord.* 51 (2021), 102923, <https://doi.org/10.1016/j.msard.2021.102923>.
- [202] D. Marastoni, A.I. Pisani, G. Schiavi, V. Mazziotti, M. Castellaro, A. Tamanti, F. Bosello, F. Crescenzo, G.K. Ricciardi, S. Montemezzi, F.B. Pizzini, M. Calabrese, CSF TNF and osteopontin levels correlate with the response to dimethyl fumarate in early multiple sclerosis, *Ther. Adv. Neurol. Disord.* 15 (2022), 17562864221092124, <https://doi.org/10.1177/17562864221092124>.
- [203] D. Baker, M. Marta, G. Pryce, G. Giovannoni, K. Schmierer, Memory B Cells are Major Targets for Effective Immunotherapy in Relapsing Multiple Sclerosis, *EBioMedicine*. 16 (2017) 41–50, <https://doi.org/10.1016/j.ebiom.2017.01.042>.
- [204] M. Stilund, M.C. Gjelstrup, T. Petersen, H.J. Møller, P.V. Rasmussen, T. Christensen, M. Reindl, Biomarkers of inflammation and axonal degeneration/damage in patients with newly diagnosed multiple sclerosis: contributions of the soluble CD163 CSF/serum ratio to a biomarker panel, *PLoS ONE* 10 (4) (2015) e0119681, <https://doi.org/10.1371/journal.pone.0119681>.
- [205] K.D. DiSano, F. Gilli, A.R. Pachner, Intrathecally produced CXCL13: A predictive biomarker in multiple sclerosis, *Mult. Scler. J. - Exp. Transl. Clin.* 6 (2020), 2055217320981396, <https://doi.org/10.1177/2055217320981396>.
- [206] R. Magliozzi, O. Howell, A. Vora, B. Serafini, R. Nicholas, M. Puopolo, R. Reynolds, F. Aloisi, Meningeal B-cell follicles in secondary progressive multiple sclerosis associate with early onset of disease and severe cortical pathology, *Brain, J. Neurol.* 130 (2007) 1089–1104, <https://doi.org/10.1093/brain/awm038>.
- [207] B. Serafini, B. Rosicarelli, R. Magliozzi, E. Stigliano, F. Aloisi, Detection of ectopic B-cell follicles with germinal centers in the meninges of patients with secondary progressive multiple sclerosis, *Brain Pathol. Zurich Switz.* 14 (2004) 164–174, <https://doi.org/10.1111/j.1750-3639.2004.tb00049.x>.
- [208] A. Guyon, CXCL12 chemokine and its receptors as major players in the interactions between immune and nervous systems, *Front. Cell. Neurosci.* 8 (2014) 65, <https://doi.org/10.3389/fncel.2014.00065>.
- [209] R.S. Klein, J.B. Rubin, Immune and nervous system CXCL12 and CXCR4: parallel roles in patterning and plasticity, *Trends Immunol.* 25 (2004) 306–314, <https://doi.org/10.1016/j.it.2004.04.002>.
- [210] F. Lazarini, T.N. Tham, P. Casanova, F. Arenzana-Seisdedos, M. Dubois-Dalcq, Role of the alpha-chemokine stromal cell-derived factor (SDF-1) in the developing and mature central nervous system, *Glia*. 42 (2003) 139–148, <https://doi.org/10.1002/glia.10139>.
- [211] M. Krumbholz, D. Theil, S. Cepok, B. Hemmer, P. Kivisäkk, R.M. Ransohoff, M. Hofbauer, C. Farina, T. Derfuss, C. Hartle, J. Newcombe, R. Hohlfeld, E. Meinl, Chemokines in multiple sclerosis: CXCL12 and CXCL13 up-regulation is differentially linked to CNS immune cell recruitment, *Brain, J. Neurol.* 129 (2006) 200–211, <https://doi.org/10.1093/brain/awh680>.
- [212] B.O. Fabrick, H.J. Møller, R.P.M. Vloet, L.M. van Winsen, R. Hanemaaijer, C. E. Teunissen, B.M.J. Uitdehaag, T.K. van den Berg, C.D. Dijkstra, Proteolytic shedding of the macrophage scavenger receptor CD163 in multiple sclerosis, *J. Neuroimmunol.* 187 (2007) 179–186, <https://doi.org/10.1016/j.jneuroim.2007.04.016>.
- [213] M. Stilund, A.-K. Reuschlein, T. Christensen, H.J. Møller, P.V. Rasmussen, T. Petersen, S. Nataf, Soluble CD163 as a marker of macrophage activity in newly diagnosed patients with multiple sclerosis, *PLoS One*. 9 (6) (2014) e98588, <https://doi.org/10.1371/journal.pone.0098588>.
- [214] M. Niino, T. Fukazawa, N. Minami, I. Amino, J. Tashiro, N. Fujiki, S. Doi, S. Kikuchi, CD5-positive B cell subsets in secondary progressive multiple sclerosis, *Neurosci. Lett.* 523 (2012) 56–61, <https://doi.org/10.1016/j.neulet.2012.06.041>.
- [215] P. Youinou, L.E. Mackenzie, A. Lamour, R.A. Mageed, P.M. Lydyard, Human CD5-positive B cells in lymphoid malignancy and connective tissue diseases, *Eur. J. Clin. Invest.* 23 (1993) 139–150, <https://doi.org/10.1111/j.1365-2362.1993.tb00753.x>.
- [216] P. Bongioanni, C. Fioretti, R. Vanacore, F. Bianchi, F. Lombardo, F. Ambrogi, G. Meucci, Lymphocyte subsets in multiple sclerosis A study with two-colour fluorescence analysis, *J. Neurol. Sci.* 139 (1996) 71–77, [https://doi.org/10.1016/0022-510X\(96\)00030-5](https://doi.org/10.1016/0022-510X(96)00030-5).
- [217] T.F. Scott, J. McKolanis, W. Rothfus, E. Cottingham, Lymphocyte subsets in relapsing-remitting Multiple Sclerosis: A longitudinal study of B lymphocytes and T lymphocytes, *Neurol. Res.* 16 (1994) 385–388, <https://doi.org/10.1080/01616412.1994.11740258>.
- [218] O.A. Seidi, Y.K. Semra, M.K. Sharief, Expression of CD5 on B lymphocytes correlates with disease activity in patients with multiple sclerosis, *J. Neuroimmunol.* 133 (2002) 205–210, [https://doi.org/10.1016/S0165-5728\(02\)00360-0](https://doi.org/10.1016/S0165-5728(02)00360-0).
- [219] L.M. Villar, M. Espiño, E. Roldán, N. Marín, L. Costa-Frossard, A. Muriel, J. C. Alvarez-Cermeño, Increased peripheral blood CD5+ B cells predict earlier conversion to MS in high-risk clinically isolated syndromes, *Mult. Scler. Houndmills Basingstoke Engl.* 17 (2011) 690–694, <https://doi.org/10.1177/1352458510396922>.
- [220] R.F. Ludueña, Multiple forms of tubulin: different gene products and covalent modifications, *Int. Rev. Cytol.* 178 (1998) 207–275, [https://doi.org/10.1016/S0074-7696\(08\)62138-5](https://doi.org/10.1016/S0074-7696(08)62138-5).
- [221] N.B. Laferriere, T.H. MacRae, D.L. Brown, Tubulin synthesis and assembly in differentiating neurons, *Biochem. Cell Biol. Biochim. Biol. Cell.* 75 (2) (1997) 103–117.
- [222] K.F. Sullivan, D.W. Cleveland, Identification of conserved isotype-defining variable region sequences for four vertebrate beta tubulin polypeptide classes, *Proc. Natl. Acad. Sci. U. S. A.* 83 (1986) 4327–4331, <https://doi.org/10.1073/pnas.83.12.4327>.
- [223] S.W. Ryu, R. Stewart, D.C. Pectol, N.A. Ender, O. Wimalaratne, J.-H. Lee, C. P. Zanini, A. Harvey, J.M. Huibregtse, P. Mueller, T.T. Paull, K.J. Walters, Proteome-wide identification of HSP70/HSC70 chaperone clients in human cells, *PLoS Biol.* 18 (7) (2020) e3000606, <https://doi.org/10.1371/journal.pbio.3000606>.
- [224] A.G. Pockley, B. Henderson, G. Multhoff, Extracellular cell stress proteins as biomarkers of human disease, *Biochem. Soc. Trans.* 42 (2014) 1744–1751, <https://doi.org/10.1042/BST20140205>.
- [225] J. Radons, The human HSP70 family of chaperones: where do we stand? *Cell Stress Chaperones*. 21 (2016) 379–404, <https://doi.org/10.1007/s12192-016-0676-6>.
- [226] P. Lechner, D. Buck, L. Sick, B. Hemmer, G. Multhoff, Serum heat shock protein 70 levels as a biomarker for inflammatory processes in multiple sclerosis, *Mult. Scler. J. - Exp. Transl. Clin.* 4 (2018), 2055217318767192, <https://doi.org/10.1177/2055217318767192>.